

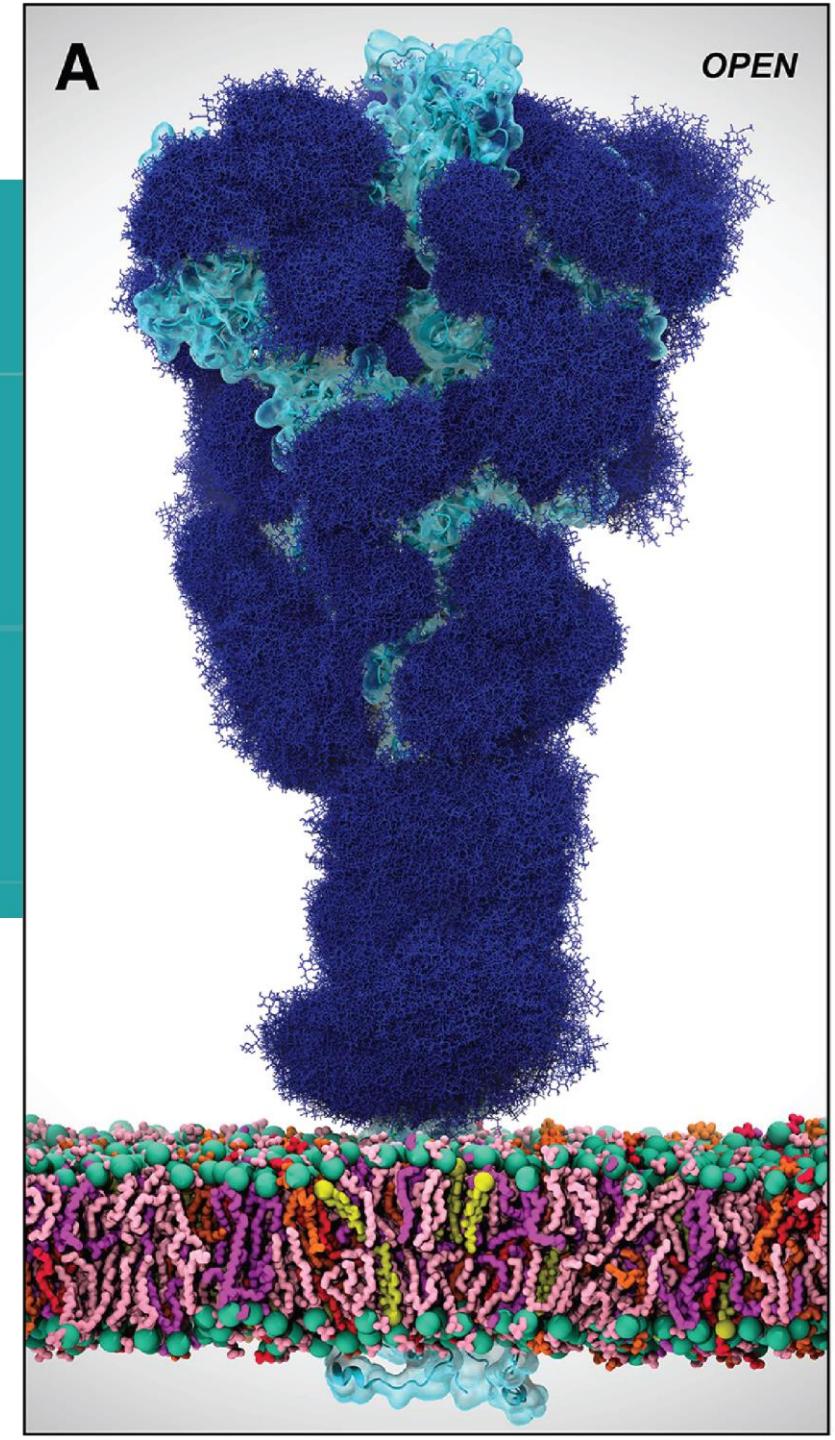
# Reasons to be excited about current efforts in glycoproteomics

Nicholas M. Riley

SUMS Seminar Series, October 1, 2020



@riley\_nm1

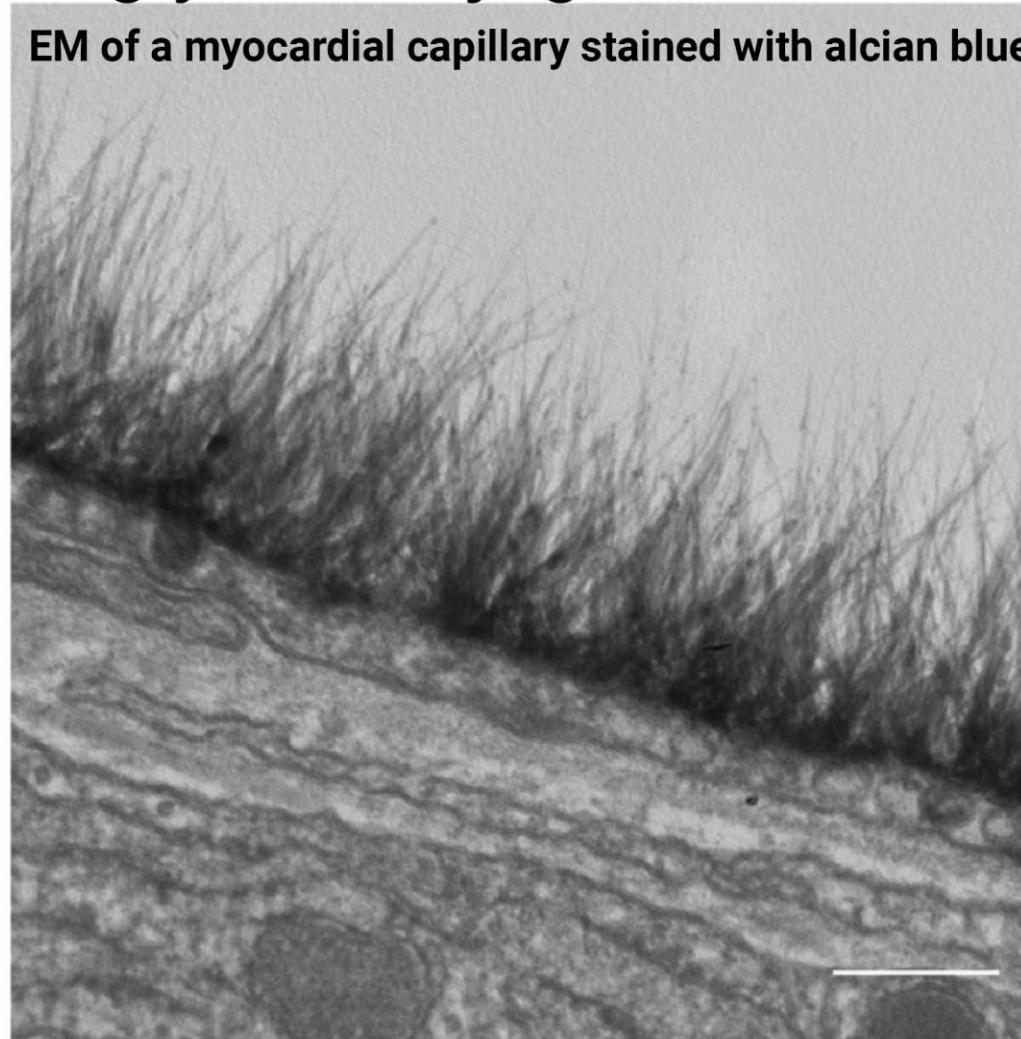




# EVERY CELL HAS A GLYCOCALYX

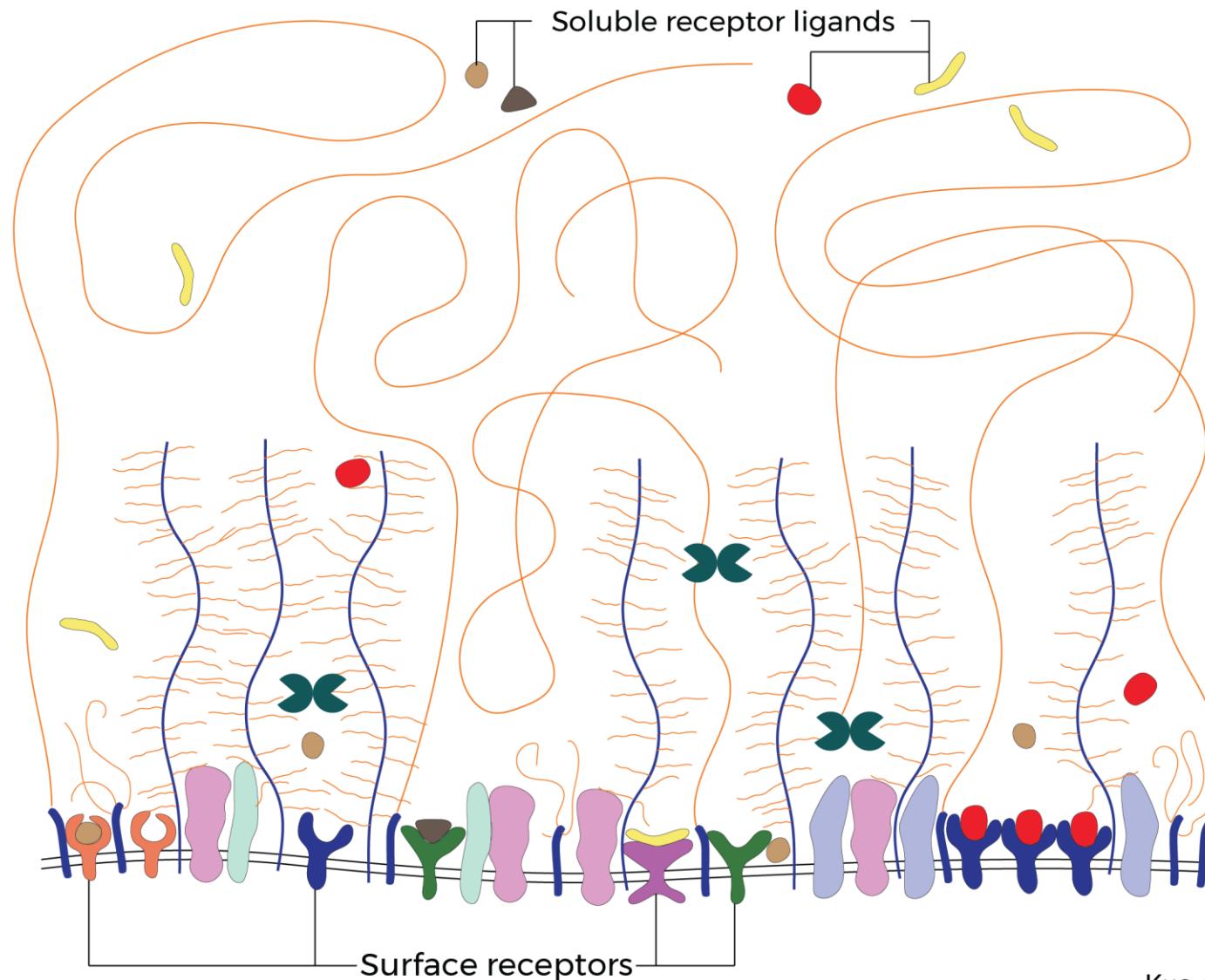
collection of glyco-conjugates at the cell surface

EM of a myocardial capillary stained with alcian blue



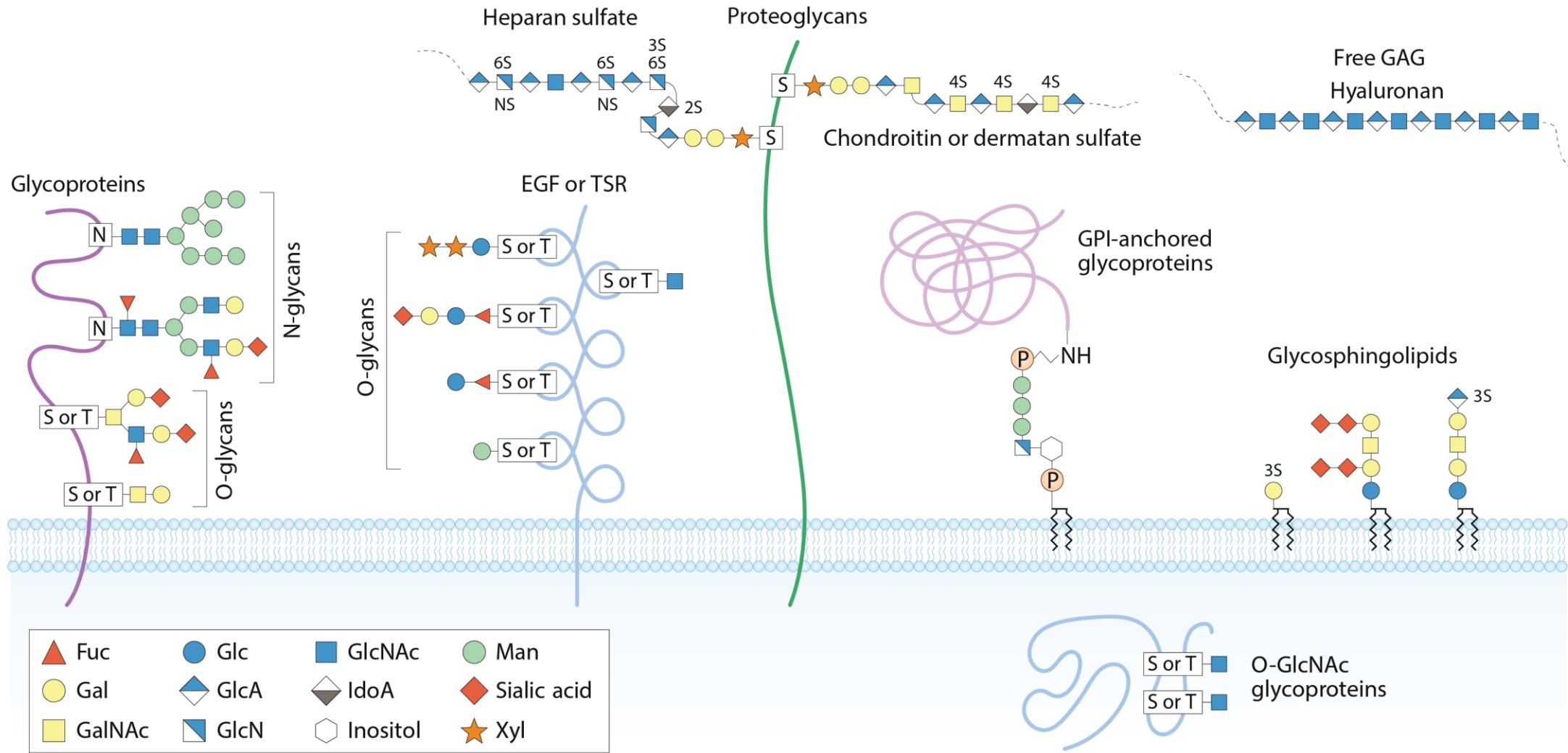


# (GLYCO)BIOLOGY AT THE CELL SURFACE



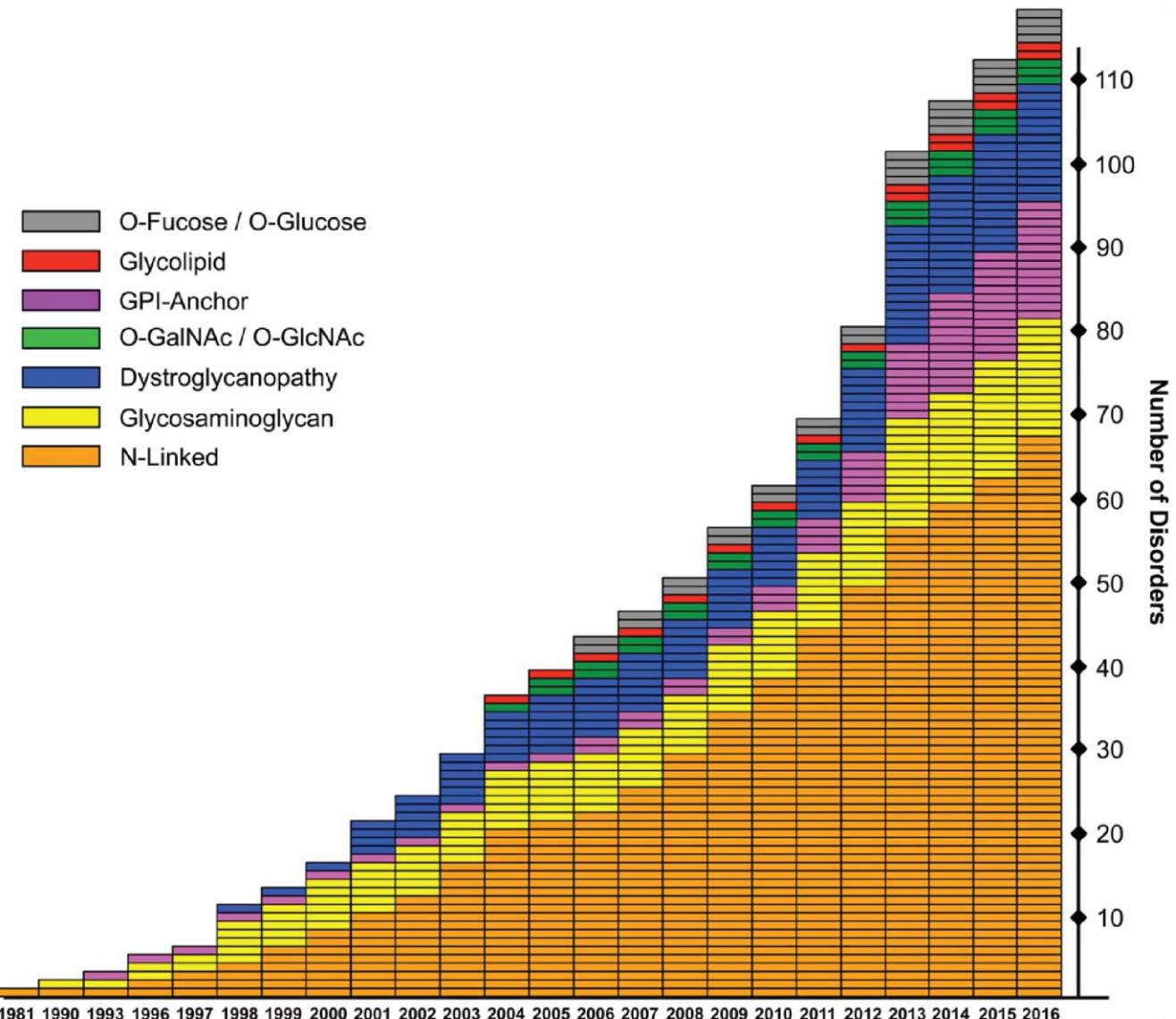
# HUMAN GLYCOSYLATION

## Glycocalyx

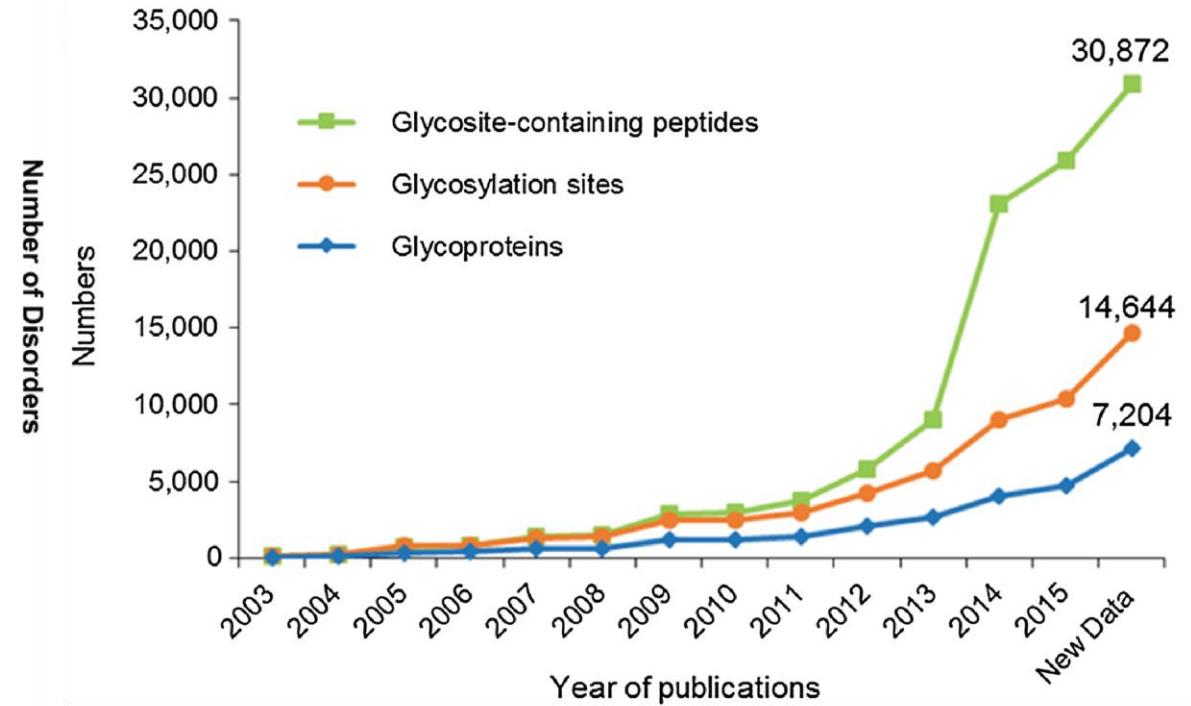




# GLYCOSYLATION IN HEALTH AND DISEASE

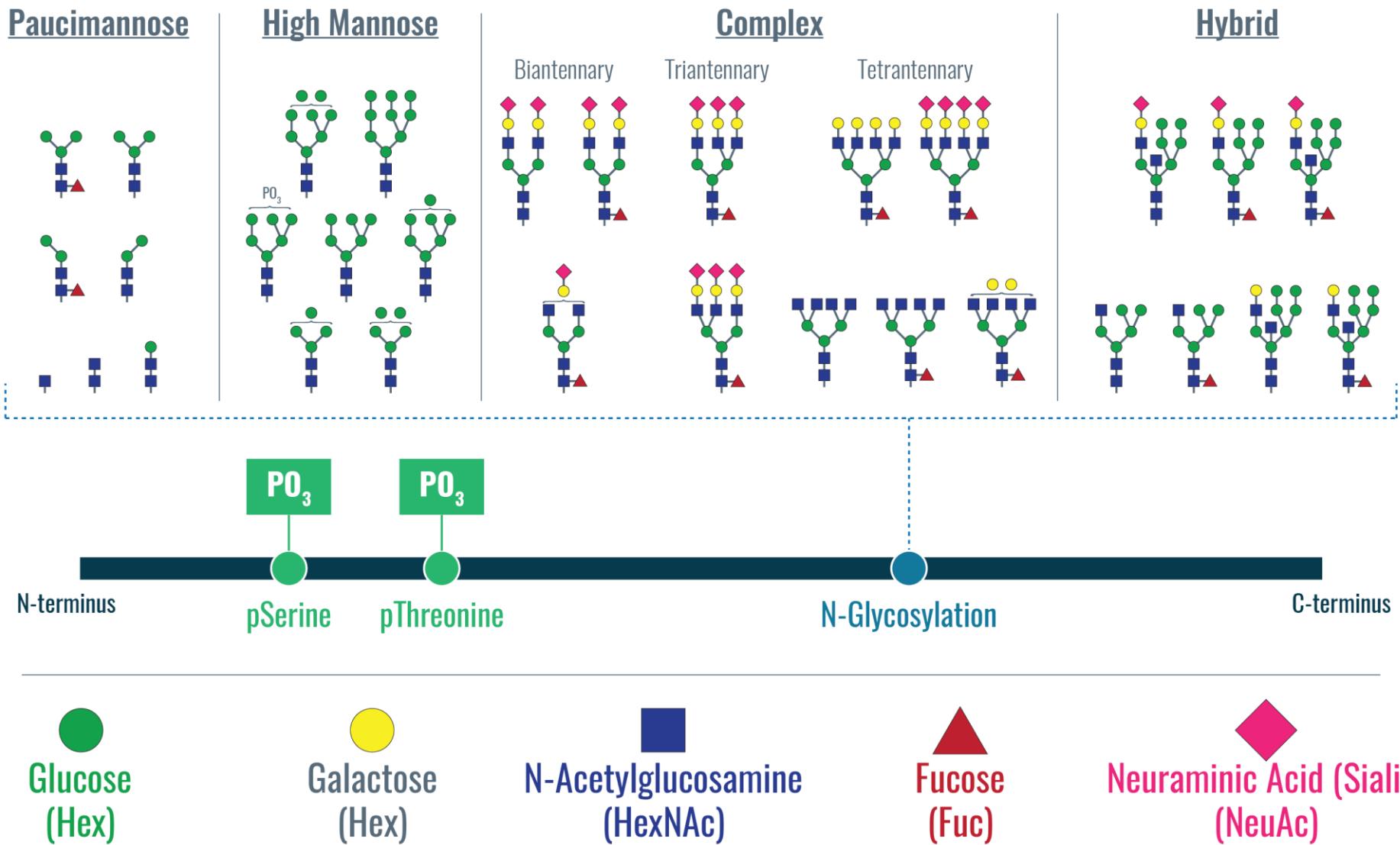


Freeze, et al., Am Journal of Hum. Genetics, 2014, 94: 161-175; updated by Varki 2017



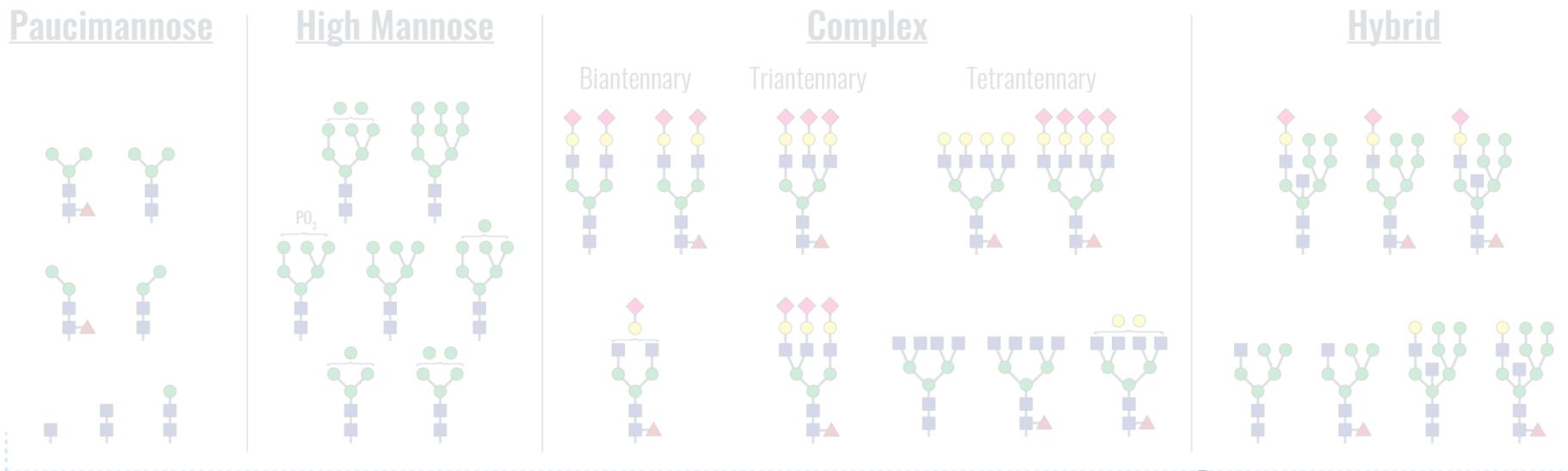
Sun, Zhang, et al., Clinical Proteomics, 2019, 16: 35

# SITE-SPECIFIC MICROHETEROGENEITY





# DE-GLYCOPEPTIDES: EASIER, BUT AT A PRICE



Glucose  
(Hex)

Galactose  
(Hex)

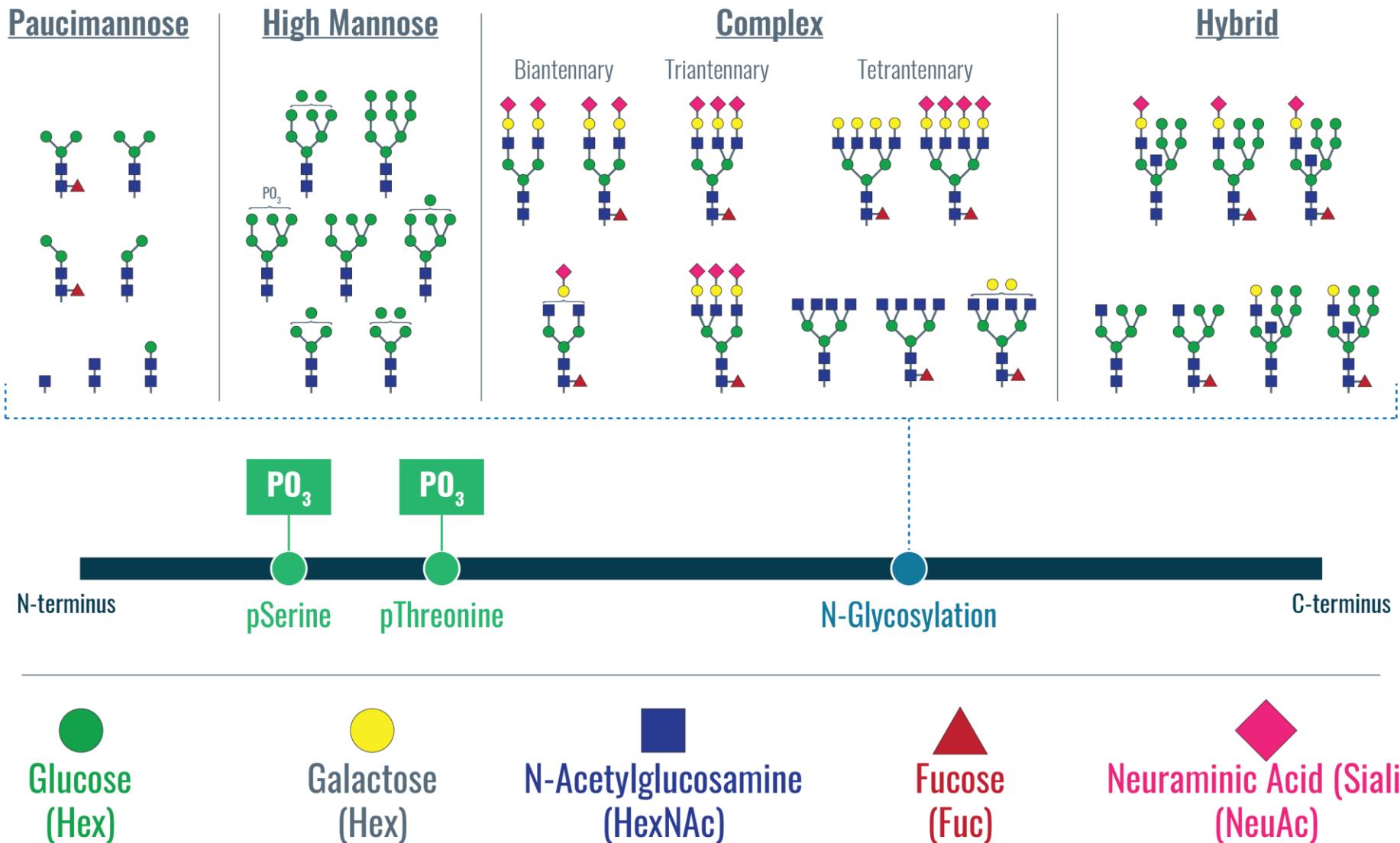
N-Acetylglucosamine  
(HexNAc)

Fucose  
(Fuc)

Neuraminic Acid (Sialic)  
(NeuAc)



# INTACT GLYCOPEPTIDES: MICROHETEROGENEITY



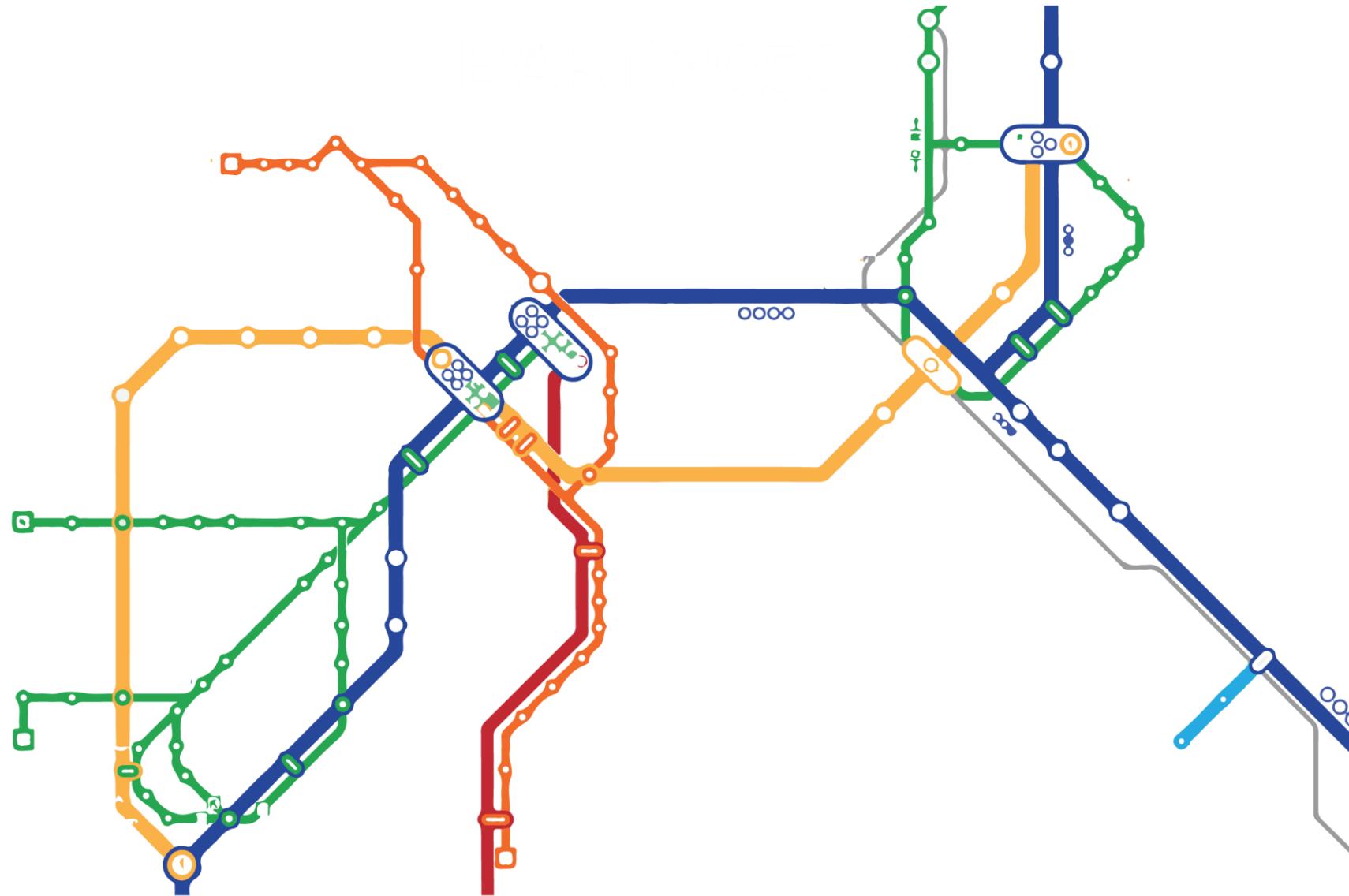


# MAPPING THE GLYCOPROTEOME



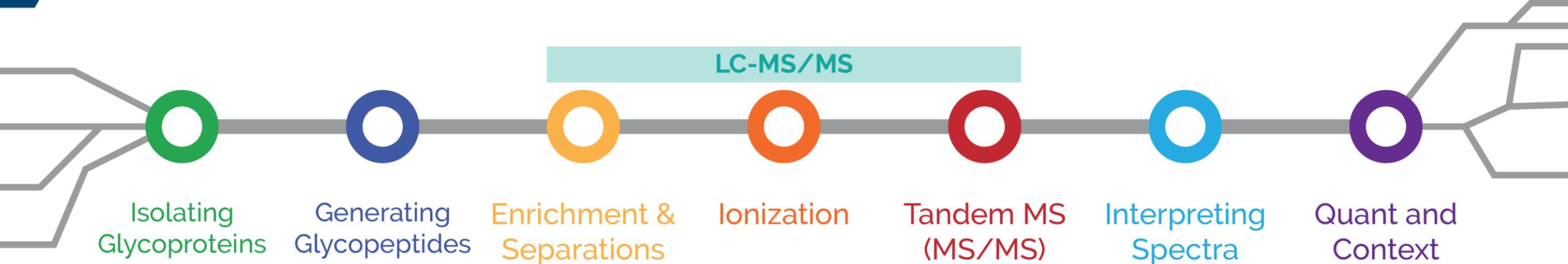


# MAPPING THE GLYCOPROTEOME



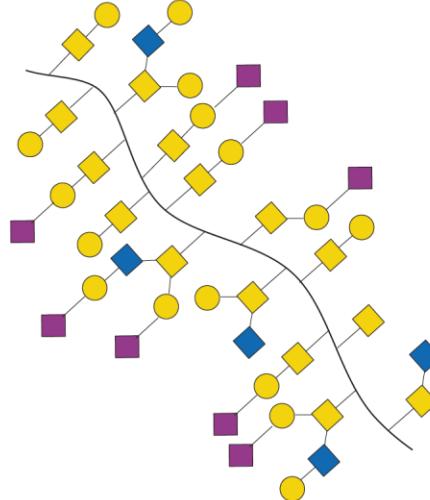


# MAPPING THE GLYCOPROTEOME

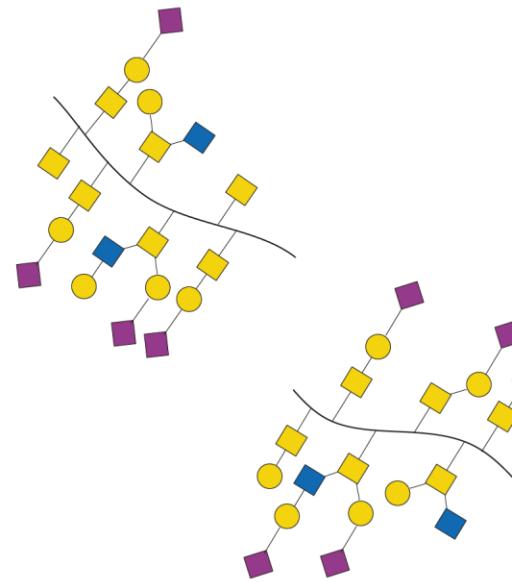




# THE RISE OF O-GLYCOPROTEASES



O-glycoproteases

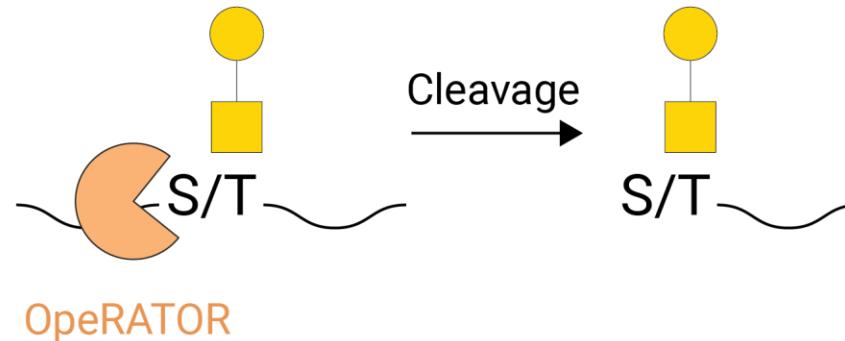


# THE RISE OF O-GLYCOPROTEASES

## OpeRATOR: broadly active on glyco-S and -T

Mapping the O-glycoproteome using site-specific extraction of O-linked glycopeptides (EXoO)

Weiming Yang<sup>ID</sup>, Minghui Ao, Yingwei Hu, Qing Kay Li & Hui Zhang\*



### Deciphering Protein O-Glycosylation: Solid-Phase Chemoenzymatic Cleavage and Enrichment

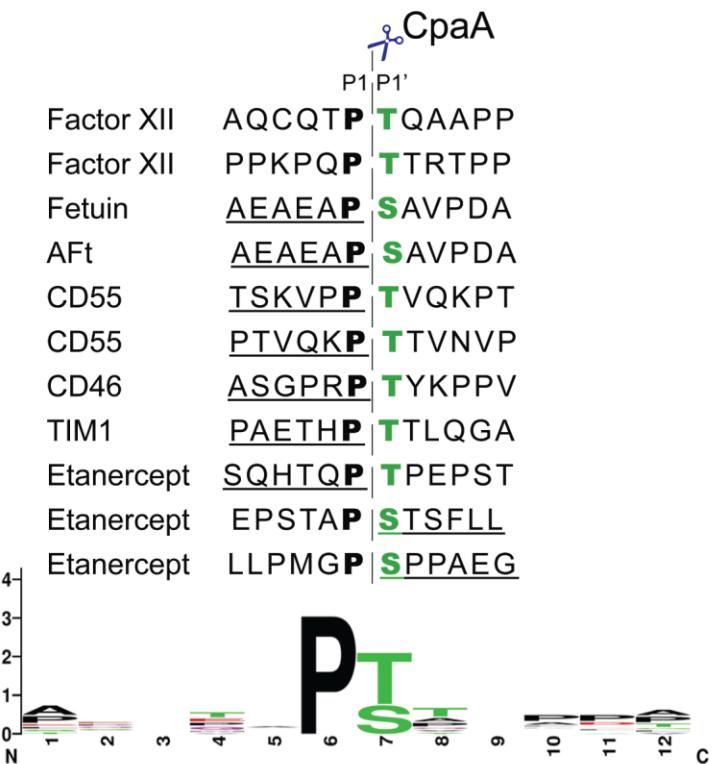
Shuang Yang, <sup>ID</sup> Philip Onigman, Wells W. Wu, Jonathan Sjogren, Helen Nyhlen, Rong-Fong Shen, and John Cipollo



The glycoprotease CpaA secreted by medically relevant *Acinetobacter* species targets multiple O-linked host glycoproteins

M. Florencia Haurat, Nichollas E. Scott, Gisela Di Venanzio, Juvenal Lopez, Benjamin Pluvinage, Alisdair B. Boraston, Michael J. Ferracane, Mario F. Feldman

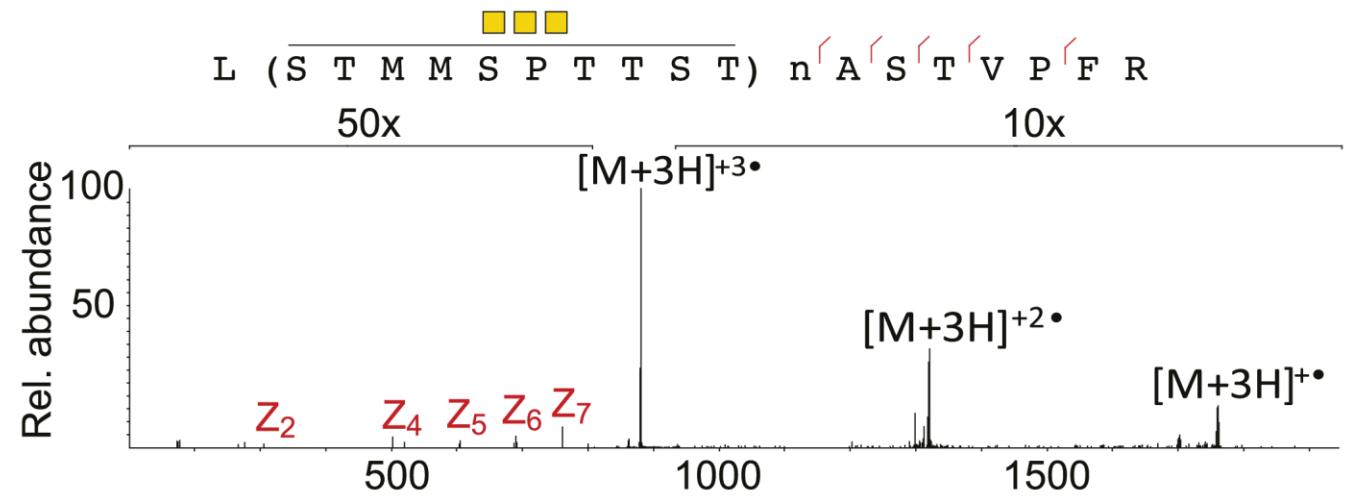
doi: <https://doi.org/10.1101/2020.07.22.216978>



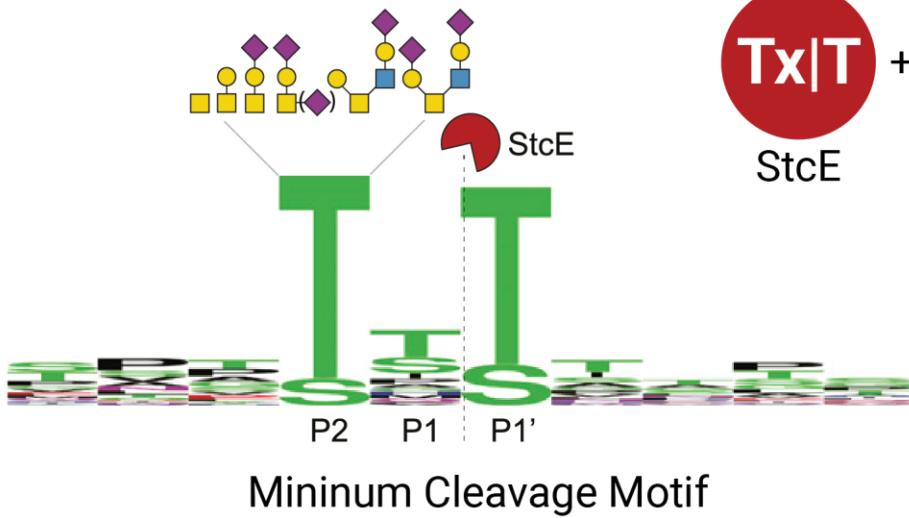


# MS-AMENABLE O-GLYCOPEPTIDES

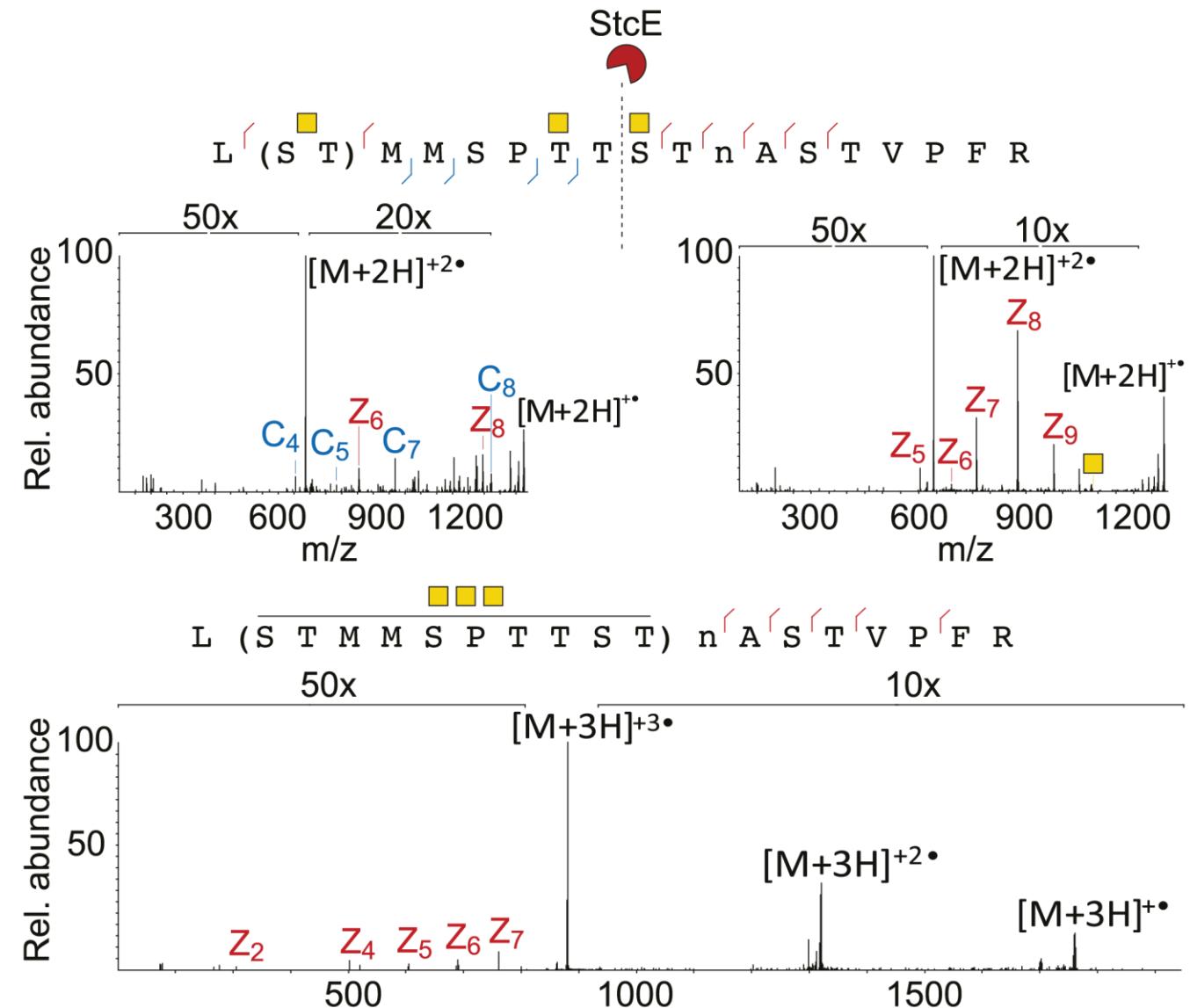
RK|  
Trypsin



# MS-AMENABLE O-GLYCOPEPTIDES



RK|  
Trypsin

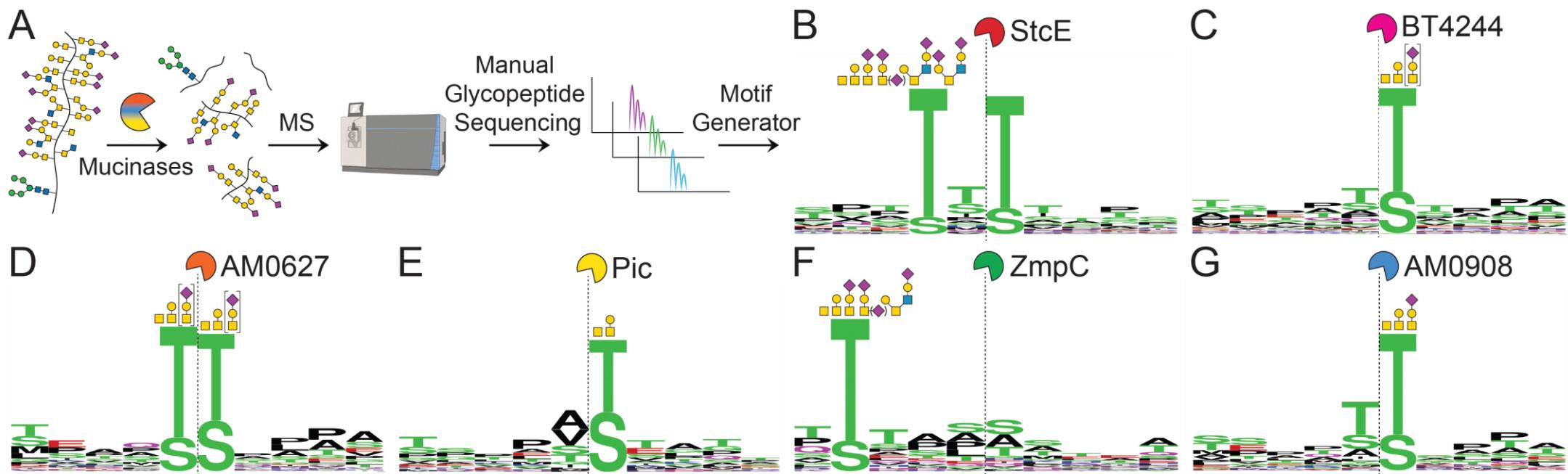


# O-GLYCOPROTEASE SUBSETS: MUCINASES

An enzymatic toolkit for selective proteolysis, detection, and visualization of mucin-domain glycoproteins

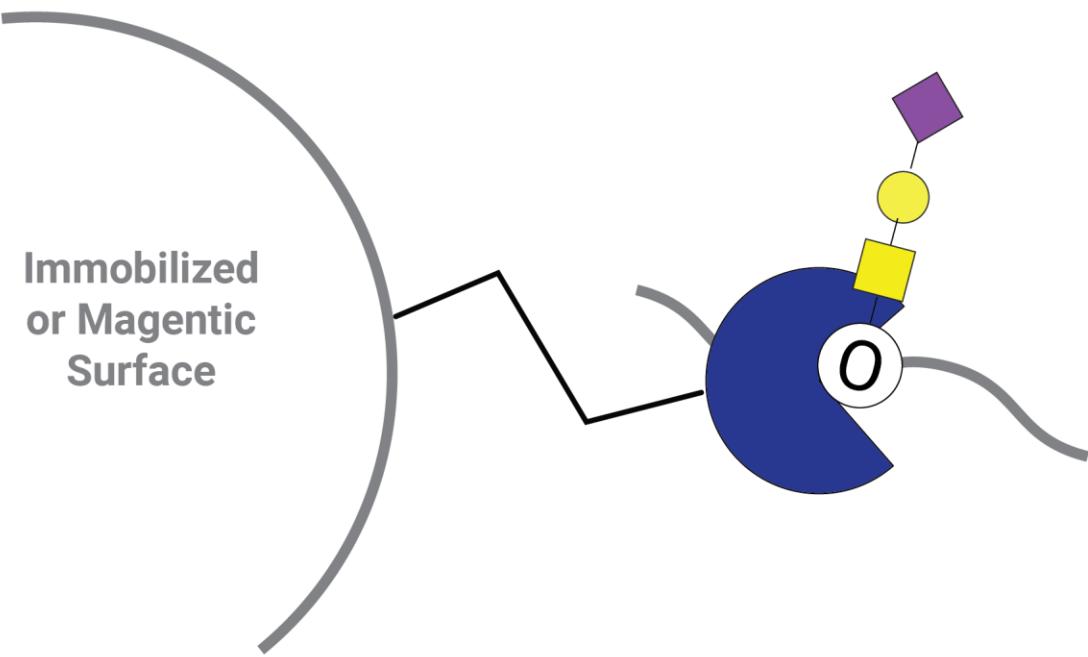
D. Judy Shon<sup>a</sup> , Stacy A. Malaker<sup>a</sup> , Kayvon Pedram<sup>a</sup> , Emily Yang<sup>a</sup>, Venkatesh Krishnan<sup>b</sup>, Oliver Dorigo<sup>b</sup> , and Carolyn R. Bertozzi<sup>a,c,1</sup> 

PNAS September 1, 2020 117 (35) 21299-21307





# O-GLYCOPROTEASE ENRICHMENTS

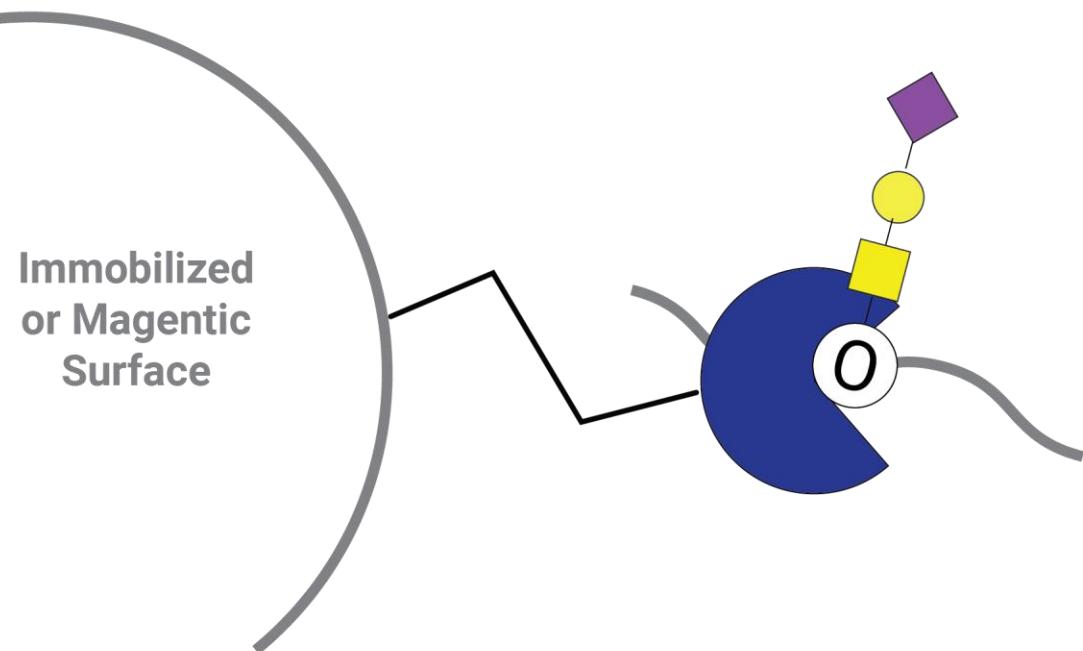




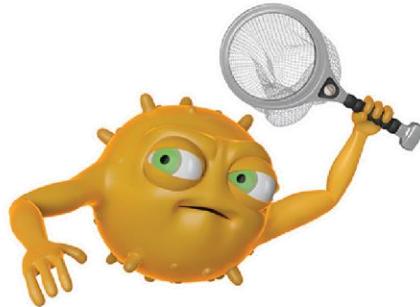
# O-GLYCOPROTEASE ENRICHMENTS



GlycoCatch: Genovis (OpeRATOR)



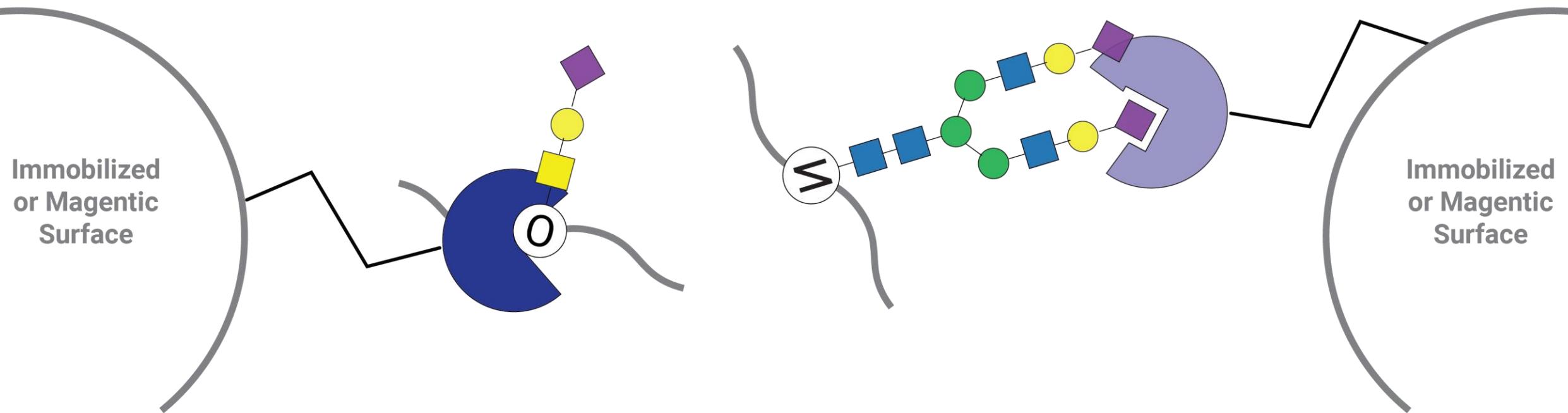
# ENZYMES FOR ENRICHMENTS IN GLYCO



GlycoCatch: Genovis (OpeRATOR)

SiaFind Lectenz Kits (Engineered Sialidases)

 **Lectenz® Bio**  
*Glycoscience Made Simple*



# CHEMICAL BIOLOGY AND GLYCOPROTEOMICS



Isolating  
Glycoproteins



Generating  
Glycopeptides



Enrichment &  
Separations

LC-MS/MS



Ionization



Tandem MS  
(MS/MS)



Interpreting  
Spectra



Quant and  
Context

## CHEMICAL REVIEWS

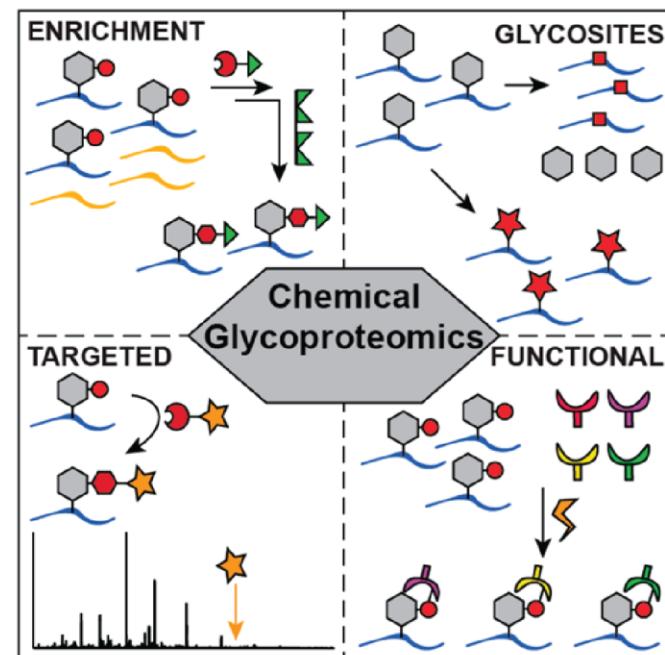
### Chemical Glycoproteomics

Krishnan K. Palaniappan<sup>†</sup> and Carolyn R. Bertozzi<sup>‡,§</sup>

<sup>†</sup>Verily Life Sciences, 269 East Grand Ave., South San Francisco, California 94080, United States

<sup>‡</sup>Department of Chemistry and <sup>§</sup>Howard Hughes Medical Institute, Stanford University, Stanford, California 94305, United States

Review  
[pubs.acs.org/CR](https://pubs.acs.org/CR)



# Bioorthogonal Chemoenzymatic

## Bioorthogonal Metabolic

**MCP** | MOLECULAR & CELLULAR PROTEOMICS

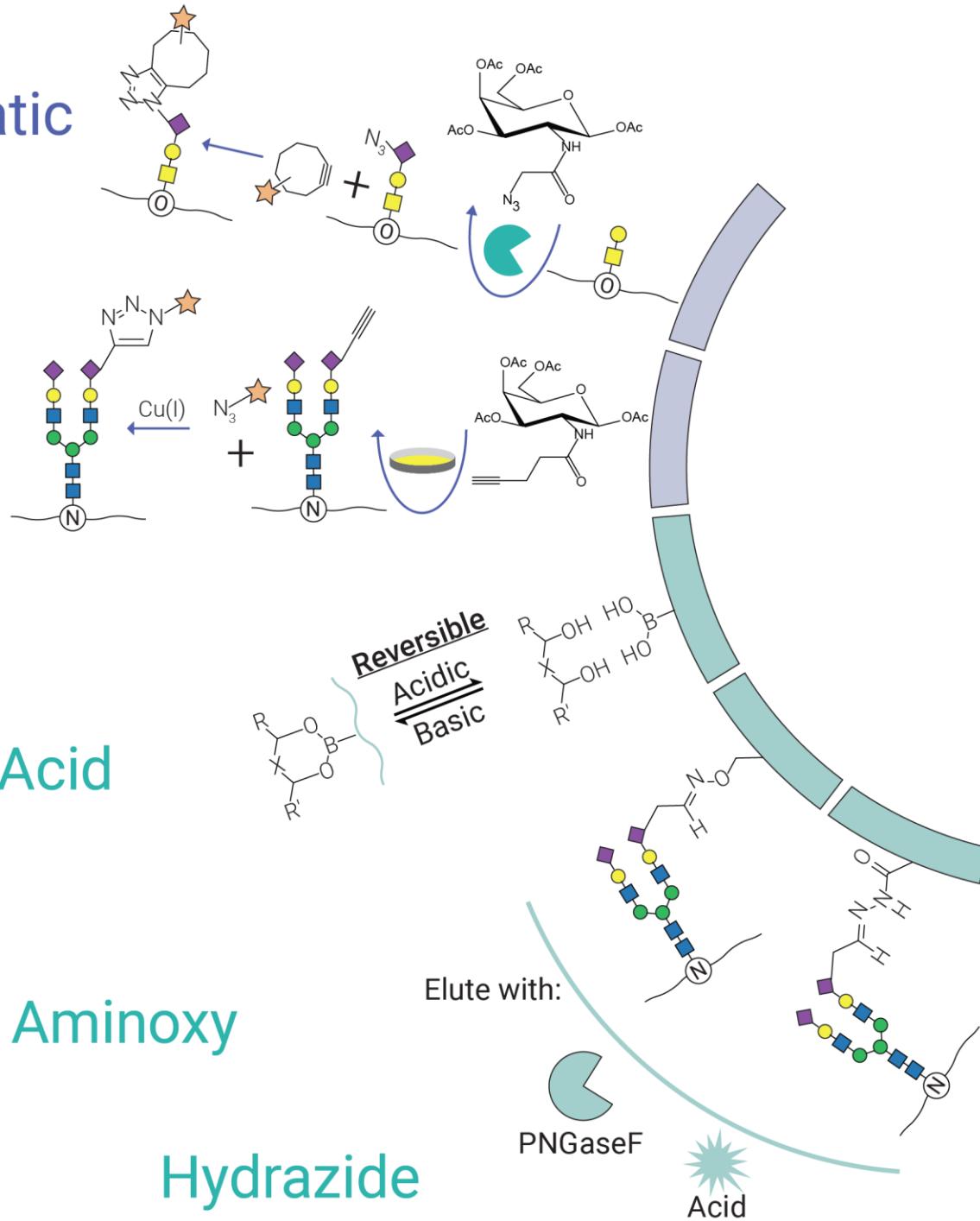
A Pragmatic Guide to  
Enrichment Strategies  
for Mass Spectrometry-  
based Glycoproteomics

Riley, Bertozzi, Pitteri,  
Mol. & Cell. Prot, 2020

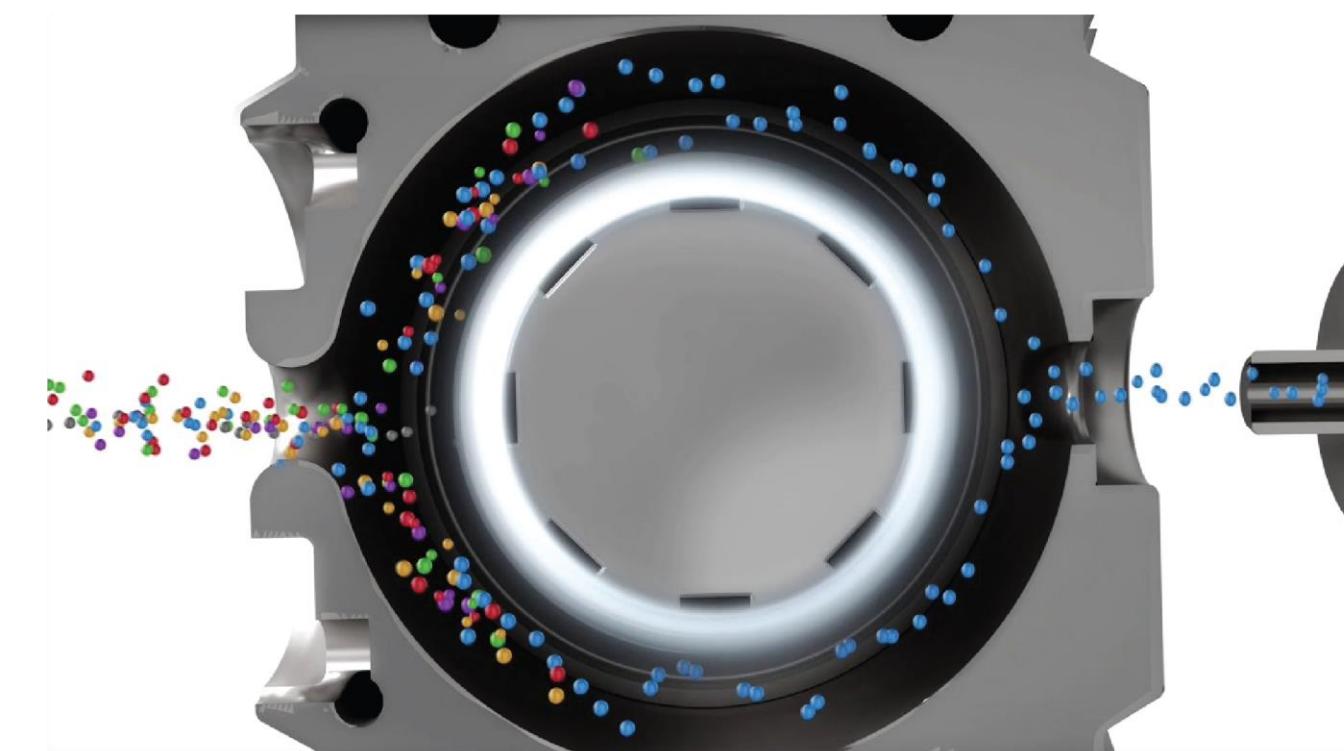
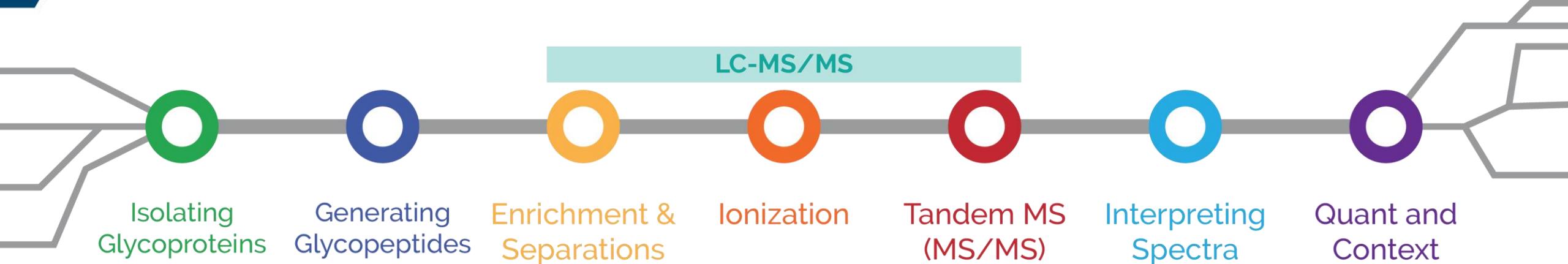
## Boronic Acid

## Aminoxy

## Hydrazide



# GAS-PHASE ENRICHMENT



**Field Asymmetric Ion Mobility Spectrometry (FAIMS)**

**bioRxiv**

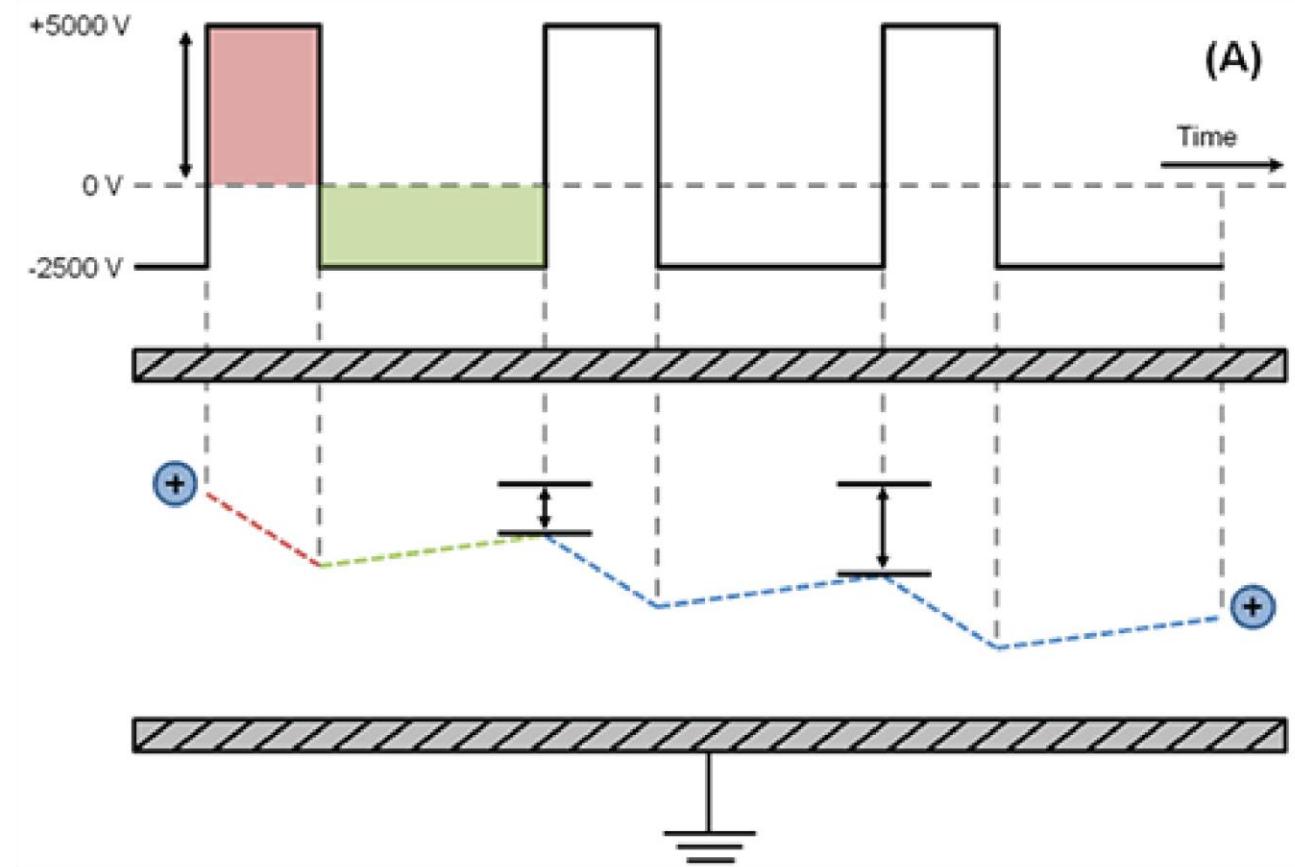
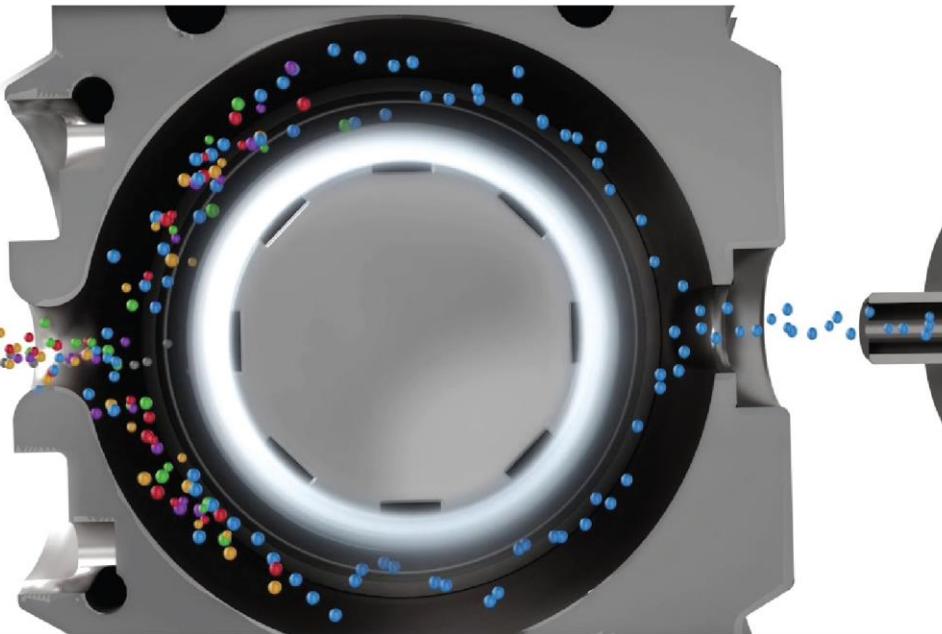
THE PREPRINT SERVER FOR BIOLOGY

What are we missing by using hydrophilic enrichment? Improving bacterial glycoproteome coverage using total proteome and FAIMS analysis.

Ameera Raudah Ahmad Izaham<sup>1</sup> Ching-Seng Ang<sup>2</sup>, Shuai Nie<sup>2</sup>, Lauren E. Bird<sup>1</sup>, Nicholas A. Williamson<sup>2</sup> and Nichollas E. Scott<sup>1#</sup>

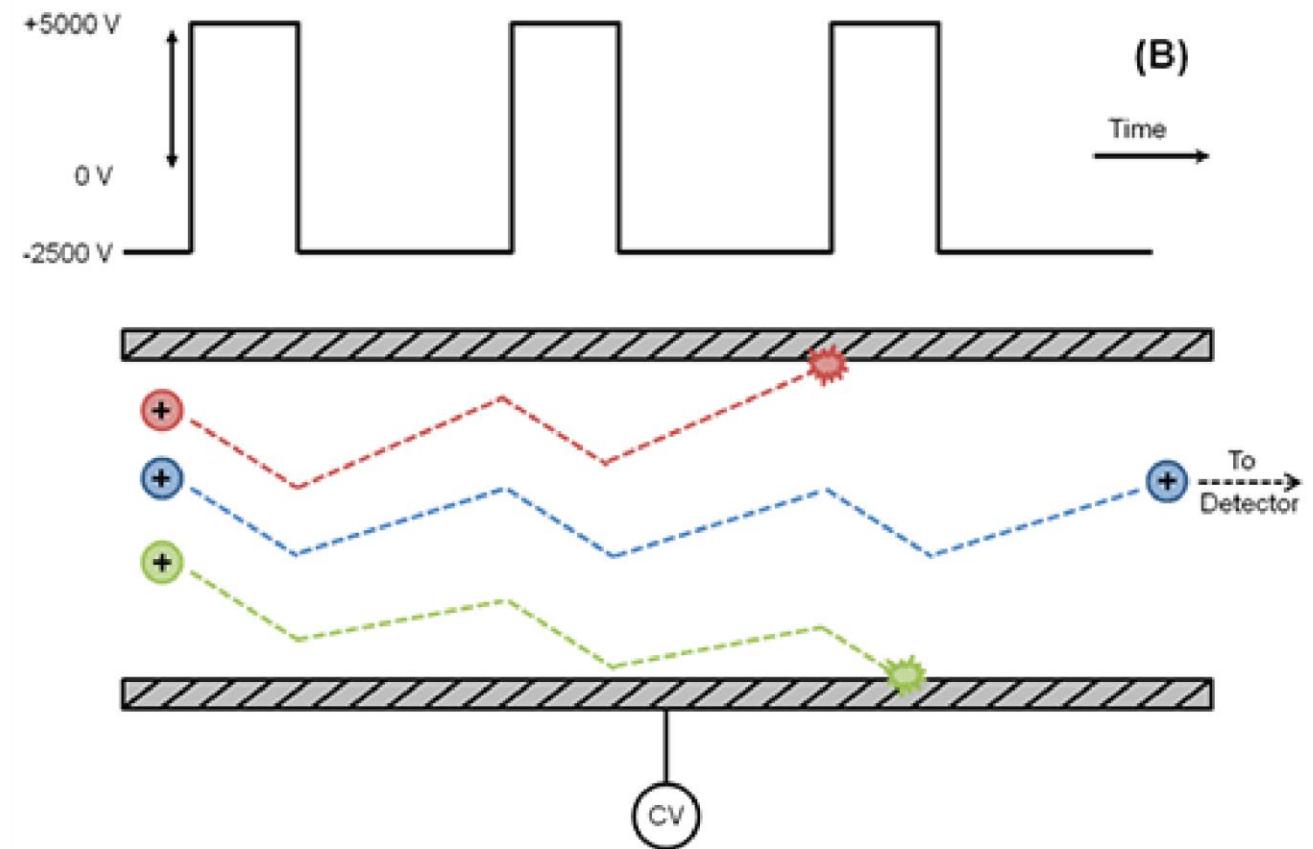
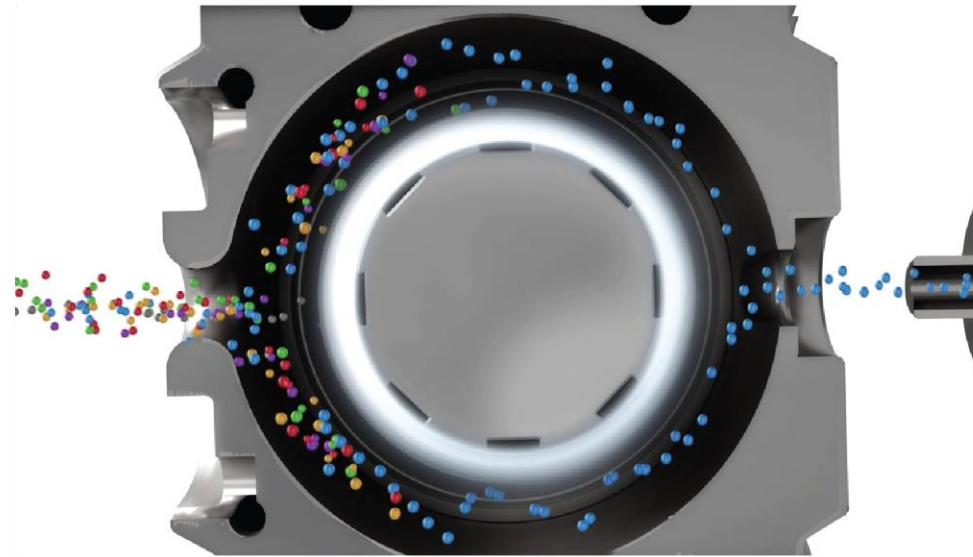


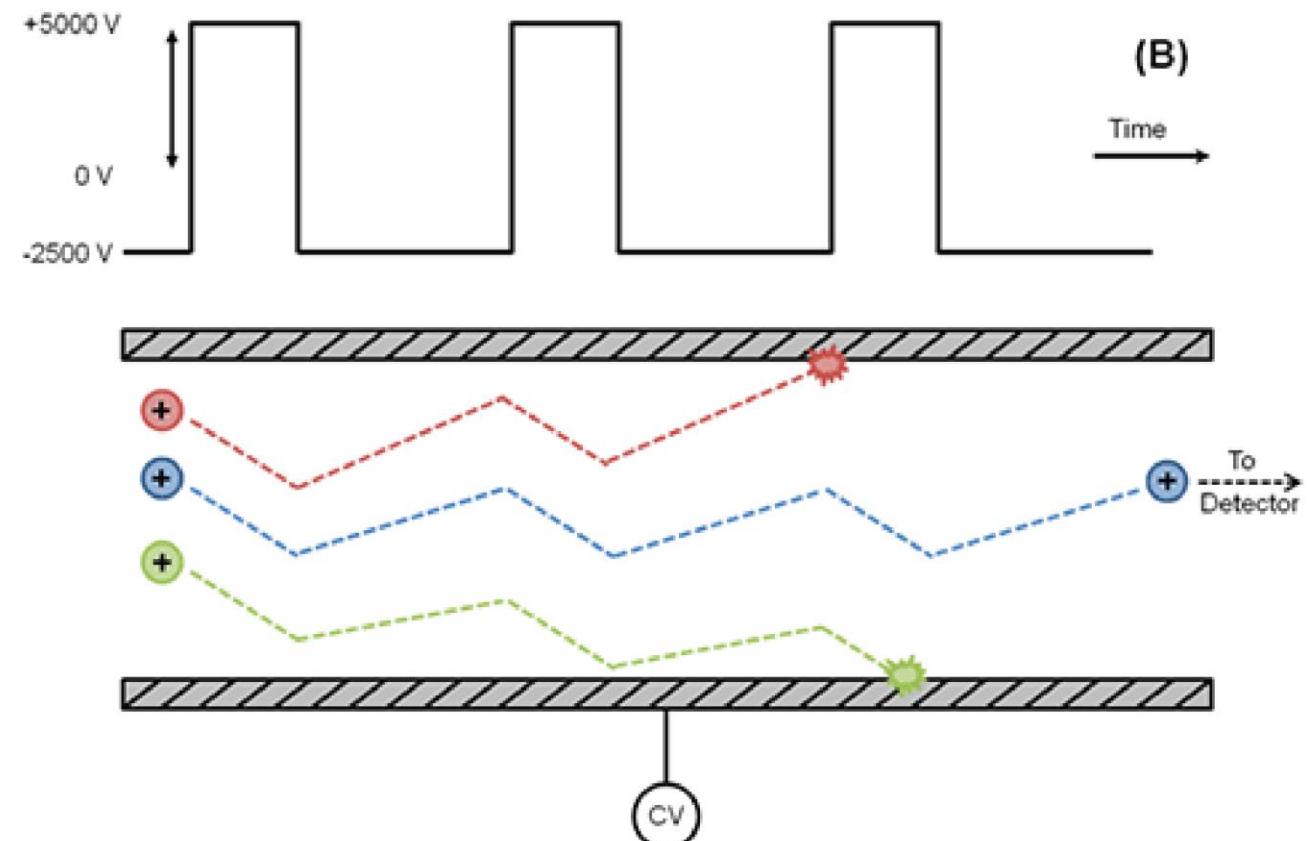
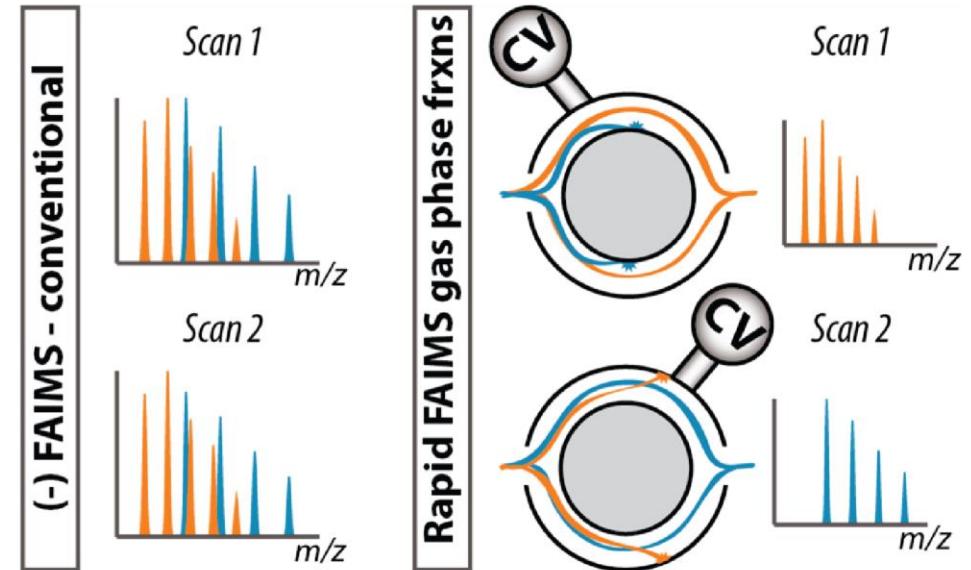
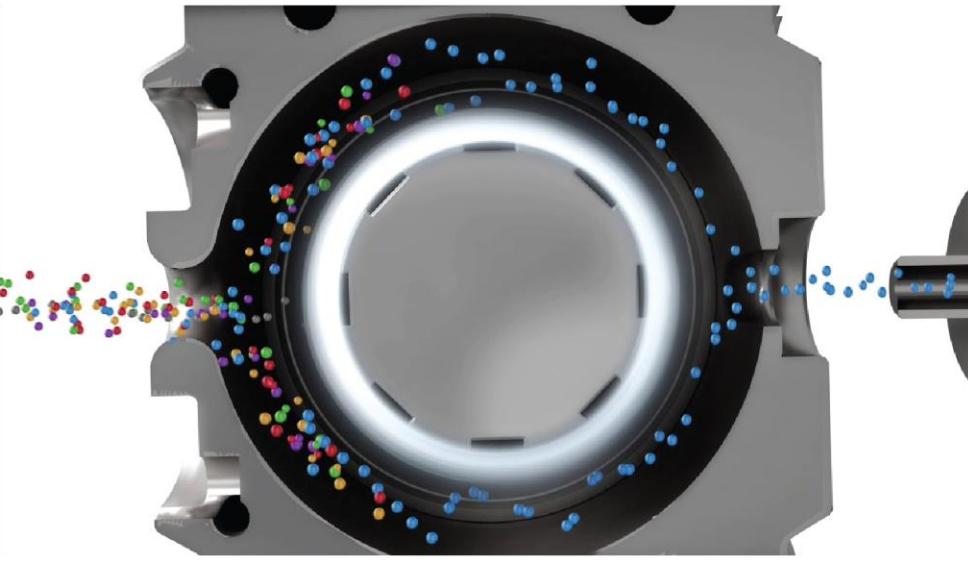
# FAIMS





# FAIMS







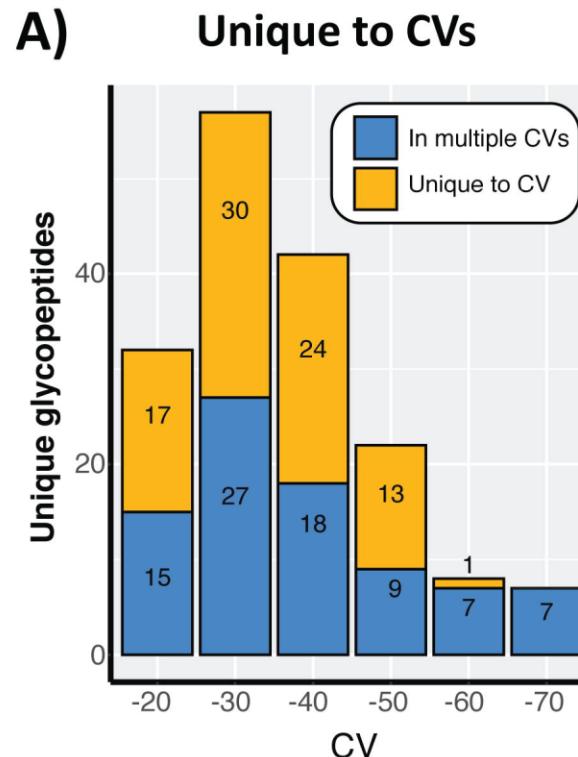
# FAIMS FOR GLYCOPEPTIDES

**bioRxiv**  
THE PREPRINT SERVER FOR BIOLOGY

What are we missing by using hydrophilic enrichment? Improving bacterial glycoproteome coverage using total proteome and FAIMS analysis.

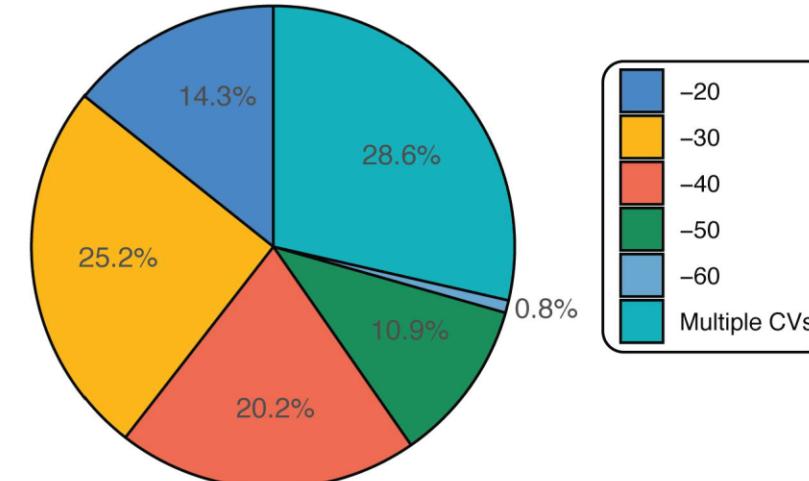
Ameera Raudah Ahmad Izaham<sup>1</sup> Ching-Seng Ang<sup>2</sup>, Shuai Nie<sup>2</sup>, Lauren E. Bird<sup>1</sup>, Nicholas A. Williamson<sup>2</sup> and Nichollas E. Scott<sup>1#</sup>

**A)**

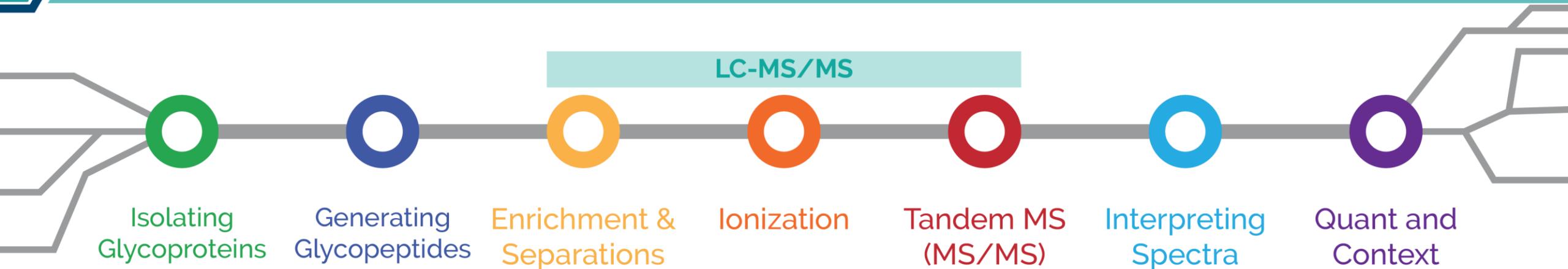


**B)**

**Unique glycopeptides observed across all CVs**

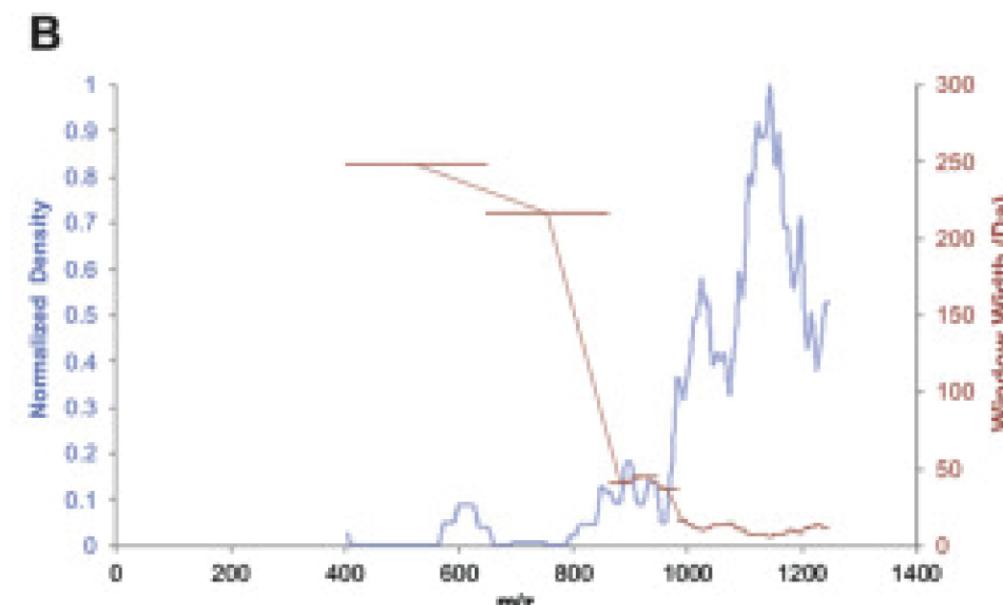
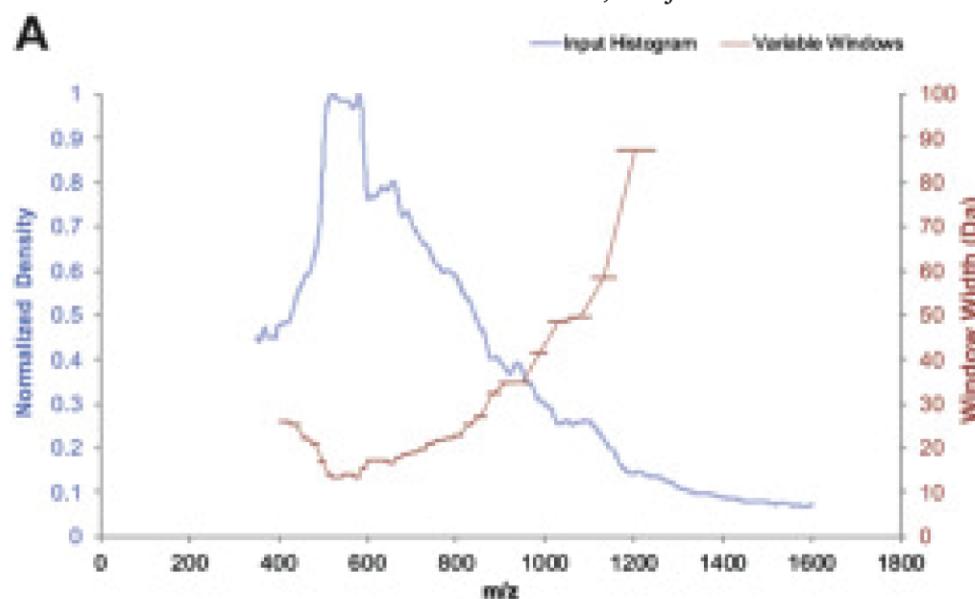


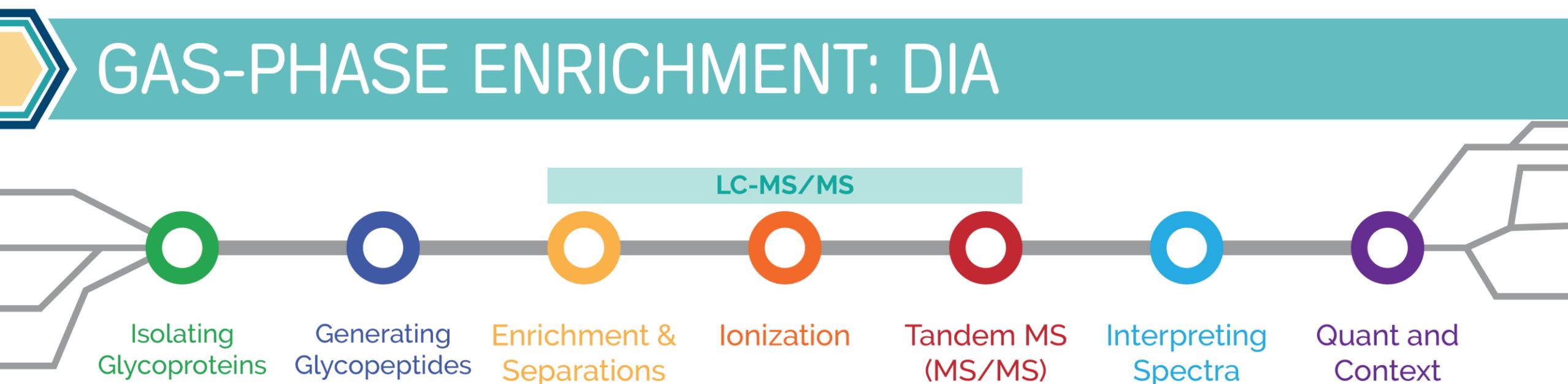
# GAS-PHASE ENRICHMENT: DIA



Glycopeptide variable window SWATH for improved data independent acquisition glycoprotein analysis

Chun Zhou<sup>a</sup>, Benjamin L. Schulz<sup>a,b,c,\*\*</sup>





ARTICLES

<https://doi.org/10.1038/s41592-019-0504-x>

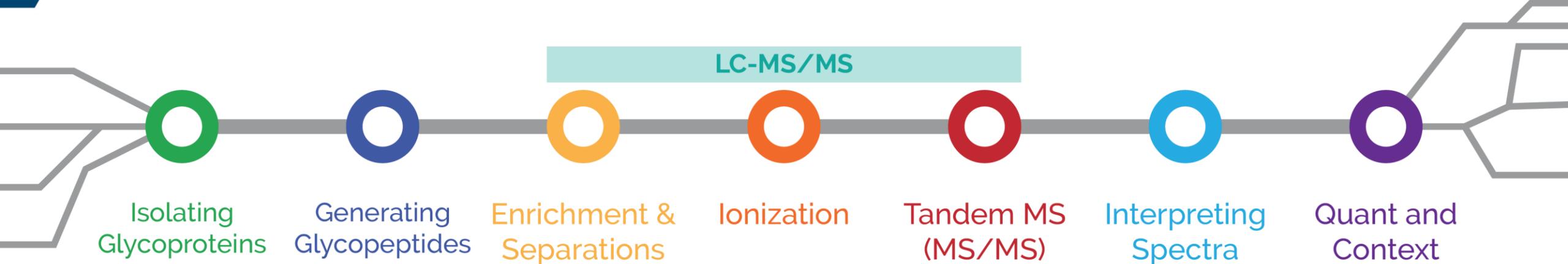
nature|methods

# Glyco-DIA: a method for quantitative O-glycoproteomics with in silico-boosted glycopeptide libraries

Zilu Ye<sup>1</sup>, Yang Mao<sup>1,2</sup>, Henrik Clausen<sup>1</sup> and Sergey Y. Vakhrushev<sup>1\*</sup>

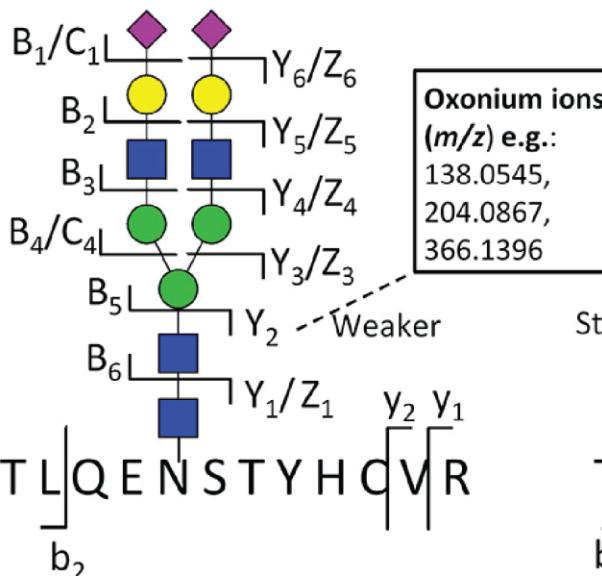
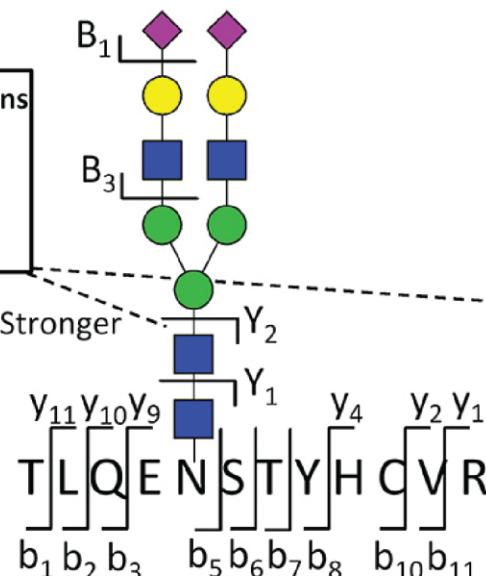
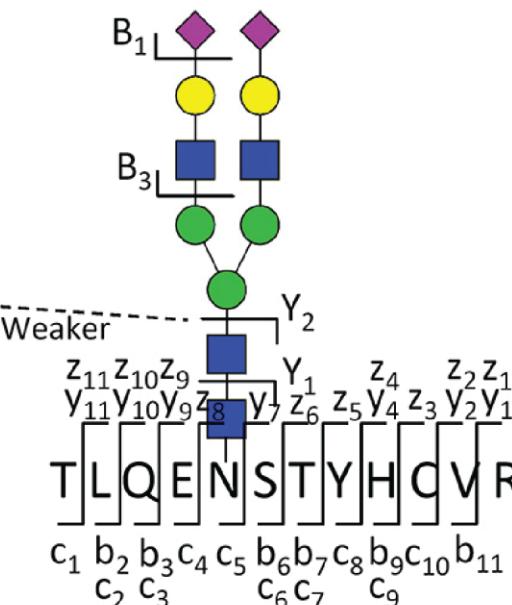
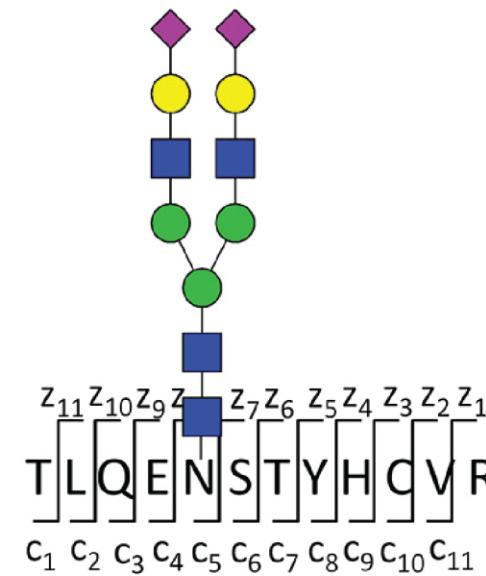


# GLYCOPEPTIDE TANDEM MS





# GLYCOPEPTIDE FRAGMENTATION

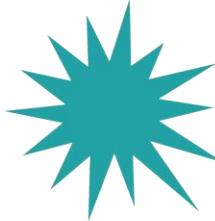
Fragmen-tation type	Resonance activation CID	Beam-type CID / HCD	Electron transfer / higher-energy dissociation (EThcD)	Conventional ETD / ECD
Typical instrument platform	Ion trap (linear, 3D)	Q-TOFs, Orbitraps	Orbitrap Fusion/Lumos	Ion traps, Orbitraps, FT-ICR
Typical bond cleavages and fragment formation	 <p>Oxonium ions (<math>m/z</math>) e.g.: 138.0545, 204.0867, 366.1396</p> <p>Weaker</p> <p>T L Q E N S T Y H C V R</p> <p>b<sub>2</sub></p>	 <p>Stronger</p> <p>T L Q E N S T Y H C V R</p> <p>b<sub>1</sub> b<sub>2</sub> b<sub>3</sub> b<sub>5</sub> b<sub>6</sub> b<sub>7</sub> b<sub>8</sub> b<sub>10</sub> b<sub>11</sub></p>	 <p>Weaker</p> <p>T L Q E N S T Y H C V R</p> <p>c<sub>1</sub> b<sub>2</sub> b<sub>3</sub> c<sub>4</sub> c<sub>5</sub> b<sub>6</sub> b<sub>7</sub> c<sub>8</sub> b<sub>9</sub> c<sub>10</sub> b<sub>11</sub></p>	 <p>Z<sub>11</sub> Z<sub>10</sub> Z<sub>9</sub> Z<sub>8</sub> Z<sub>7</sub> Z<sub>6</sub> Z<sub>5</sub> Z<sub>4</sub> Z<sub>3</sub> Z<sub>2</sub> Z<sub>1</sub></p> <p>T L Q E N S T Y H C V R</p> <p>c<sub>1</sub> c<sub>2</sub> c<sub>3</sub> c<sub>4</sub> c<sub>5</sub> c<sub>6</sub> c<sub>7</sub> c<sub>8</sub> c<sub>9</sub> c<sub>10</sub> c<sub>11</sub></p>
Glycopeptide information	<ul style="list-style-type: none"> <li>▪ Glycan identification*</li> <li>▪ Peptide mass identification</li> </ul>	<ul style="list-style-type: none"> <li>▪ Peptide identification**</li> <li>▪ Partial glycan identification</li> </ul>	<ul style="list-style-type: none"> <li>▪ Peptide identification**</li> <li>▪ Site identification</li> <li>▪ Partial glycan identification</li> </ul>	<ul style="list-style-type: none"> <li>▪ Peptide identification</li> <li>▪ Site identification</li> <li>▪ Glycan mass identification</li> </ul>



# GLYCOPEPTIDE FRAGMENTATION

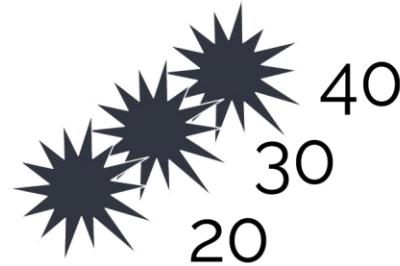
Fragmentation methods affect instrumentation decisions

HCD



Orbitrap Exploris 480 (Thermo)

sceHCD



timsTOF Pro (Bruker)

ETD



Orbitrap Eclipse (Thermo)

EThcD



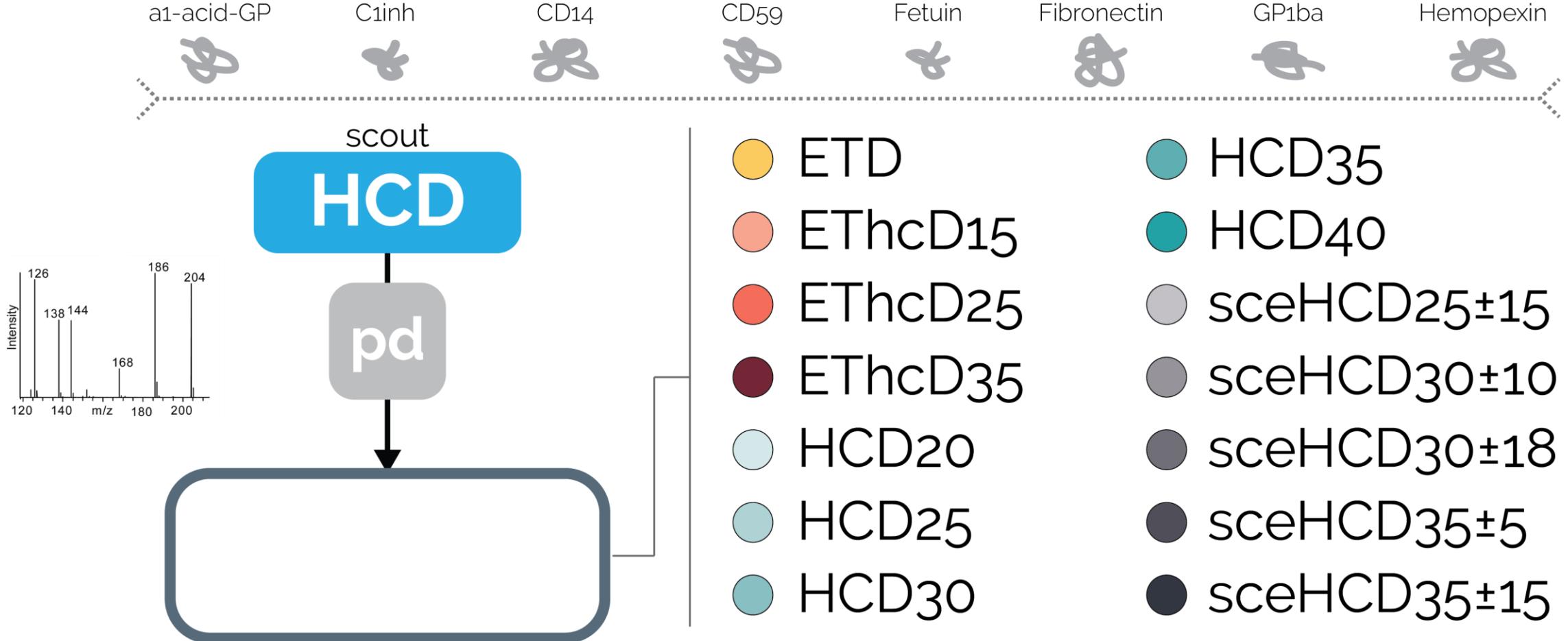
solariX XR (Bruker)



Synapt G2-Si (Waters)



# GLYCOPEPTIDE FRAGMENTATION

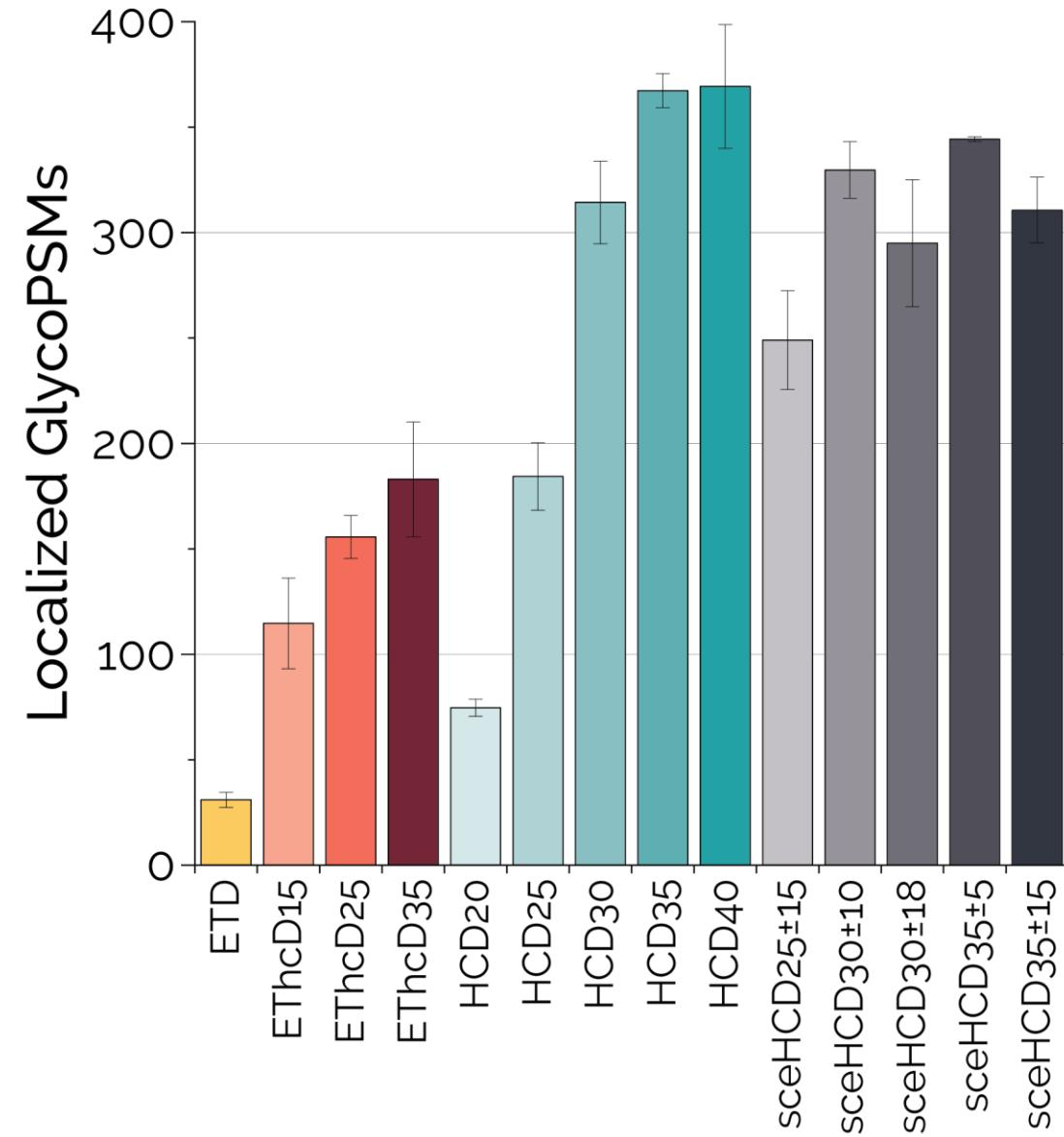




# FRAGMENTATION SCORECARD

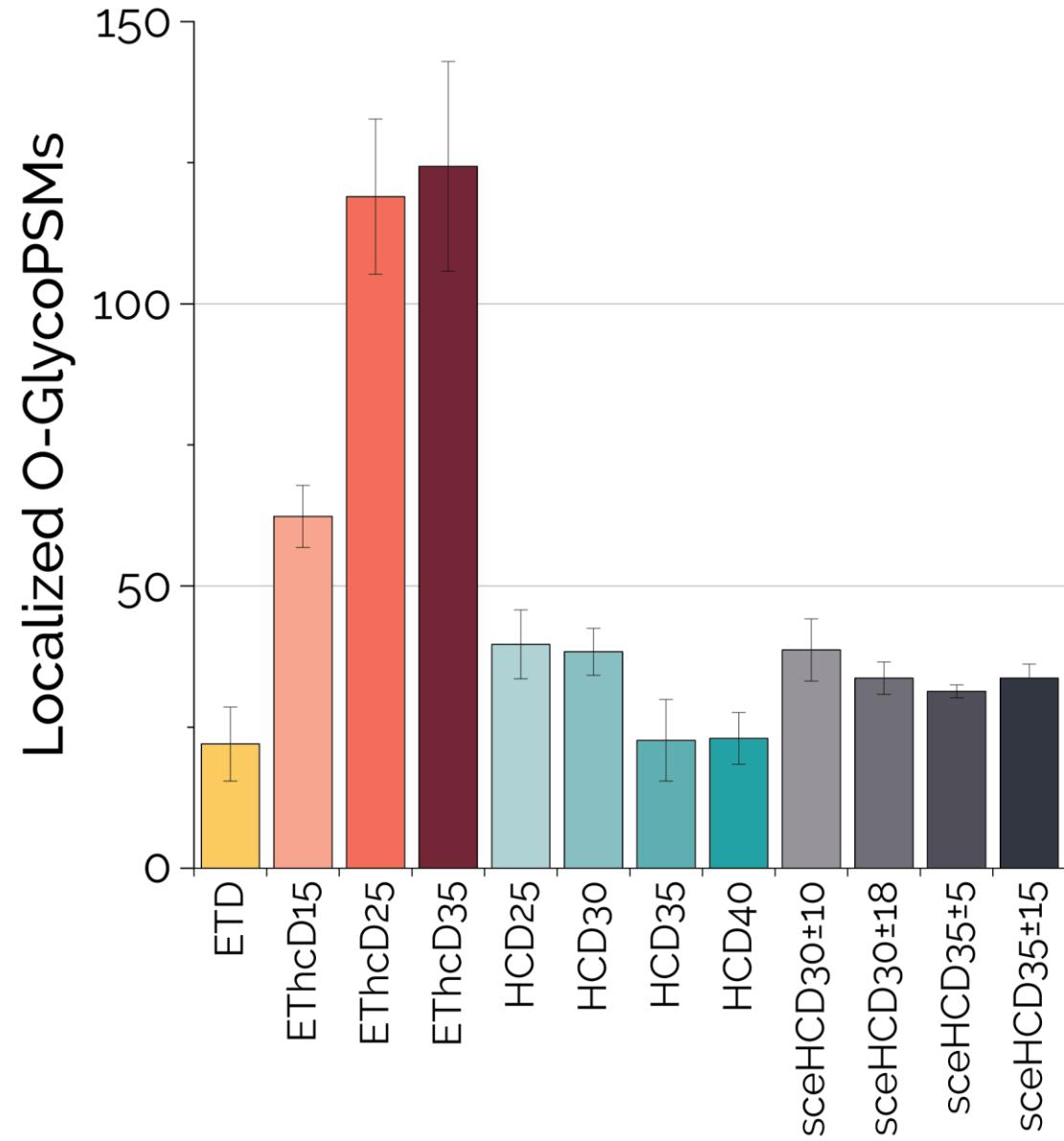


# N-GLYCOPEPTIDE FRAGMENTATION





# O-GLYCOPEPTIDE FRAGMENTATION





# FRAGMENTATION SCORECARD

Recommended  
 Good

	ETD	EThcD15	EThcD25	EThcD35	HCD20	HCD25	HCD30	HCD35	HCD40	sceHCD25 <sup>±</sup> 15	sceHCD30 <sup>±</sup> 10	sceHCD30 <sup>±</sup> 18	sceHCD35 <sup>±</sup> 5	sceHCD35 <sup>±</sup> 15
Acquisition Speed														
Peptide Fragmentation														
Glycan Fragmentation														
Localization														
Use for N-glycopeptides														
Use for O-glycopeptides														

pubs.acs.org/jpr

Article

## Optimal Dissociation Methods Differ for *N*- and *O*-Glycopeptides

Nicholas M. Riley, Stacy A. Malaker, Marc D. Driessens, and Carolyn R. Bertozzi\*



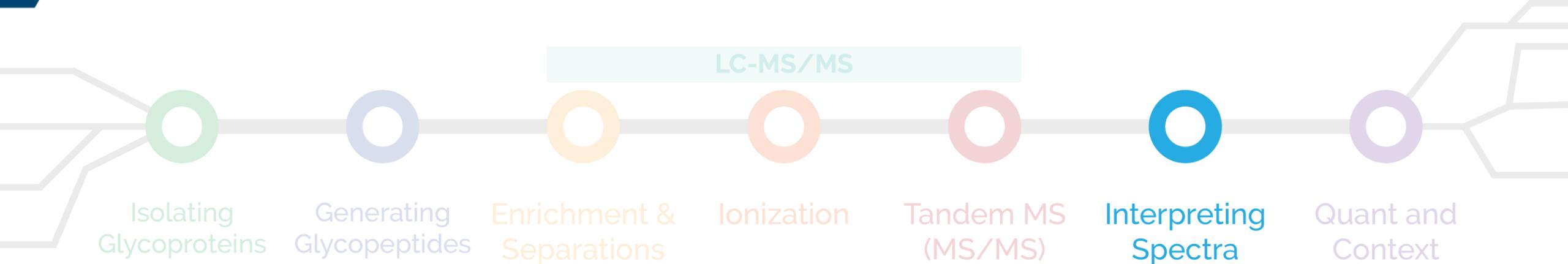
Cite This: <https://dx.doi.org/10.1021/acs.jproteome.0c00218>



Read Online



# GLYCOINFORMATICS



Available through Stanford (Protein Metrics)

GUI with viewing options

Flexible with fragmentation methods

Compatible with N- and O-glycopeptides



Free (Academic software)

GUI with limited (but still usable) options

Designed for sceHCD (more coming soon)

Generally only works for N-glycopeptides

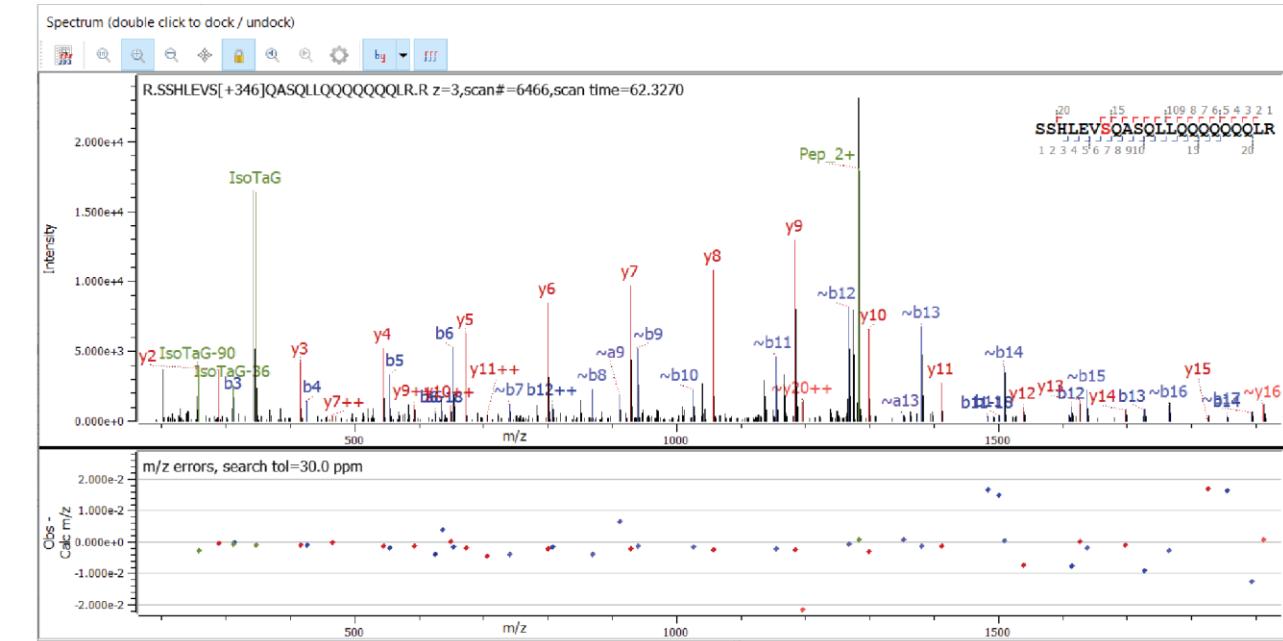
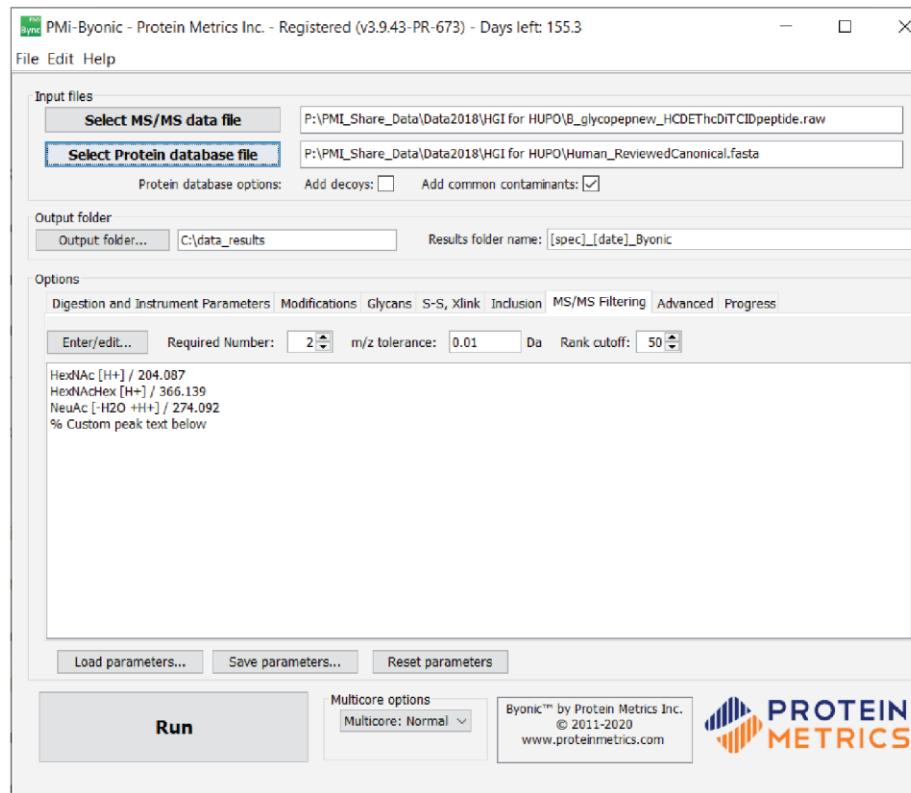


# RECENT BYONIC UPGRADES



## Peak Filtering, Peak Annotation, and Wildcard Search for Glycoproteomics

Abhishek Roushan, Gary M. Wilson, Doron Kletter, K. Ilker Sen, Wilfred Tang,  
Yong J. Kil, Eric Carlson, Marshall Bern\*





# VARIABLE MOD SEARCHING

Current Canon:

Peptide-centric  
DB generation



↓  
different theoretical spectra  
for each PTM considered

O-glycopeptide from CD43 (leukosialin)

T G S L E P S S G A S G P Q V S S V K

7 potential O-glycosites,  
allow up to 3 to be modified:

# VARIABLE MOD SEARCHING

Current Canon:

Peptide-centric  
DB generation

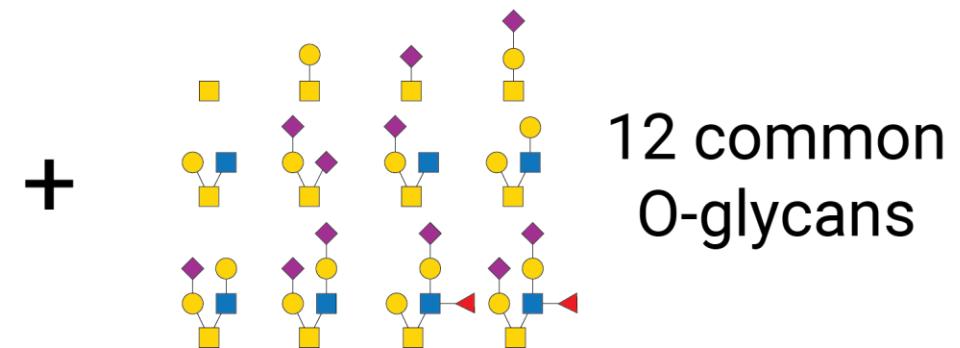


↓  
different theoretical spectra  
for each PTM considered

O-glycopeptide from CD43 (leukosialin)

T G S L E P S S G A S G P Q V S S V K

7 potential O-glycosites,  
allow up to 3 to be modified:



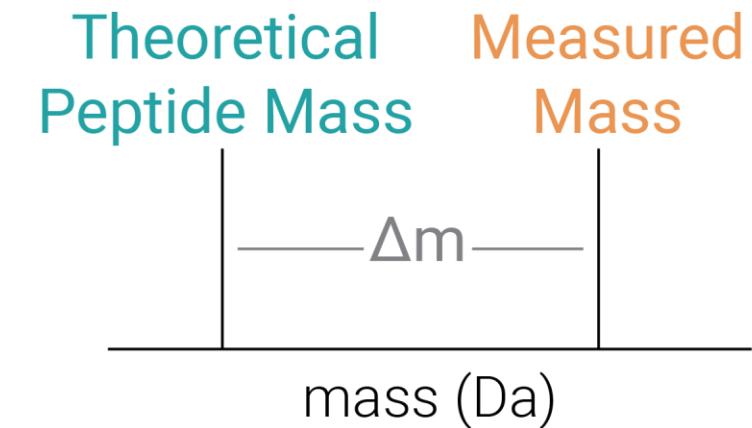
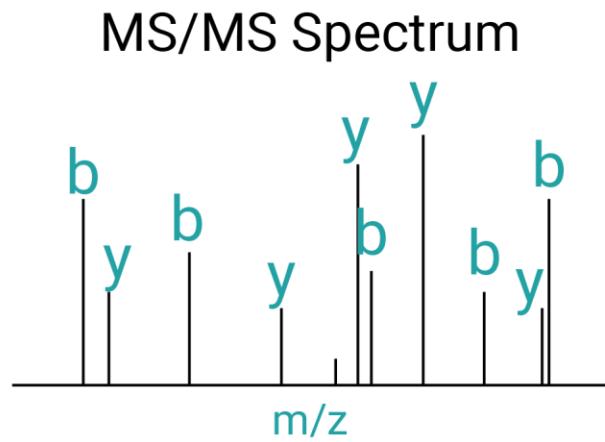
>60,000  
combinations to consider

...for one peptide



# OPEN MOD SEARCHES

Identify peptide candidates without pre-determined modifications



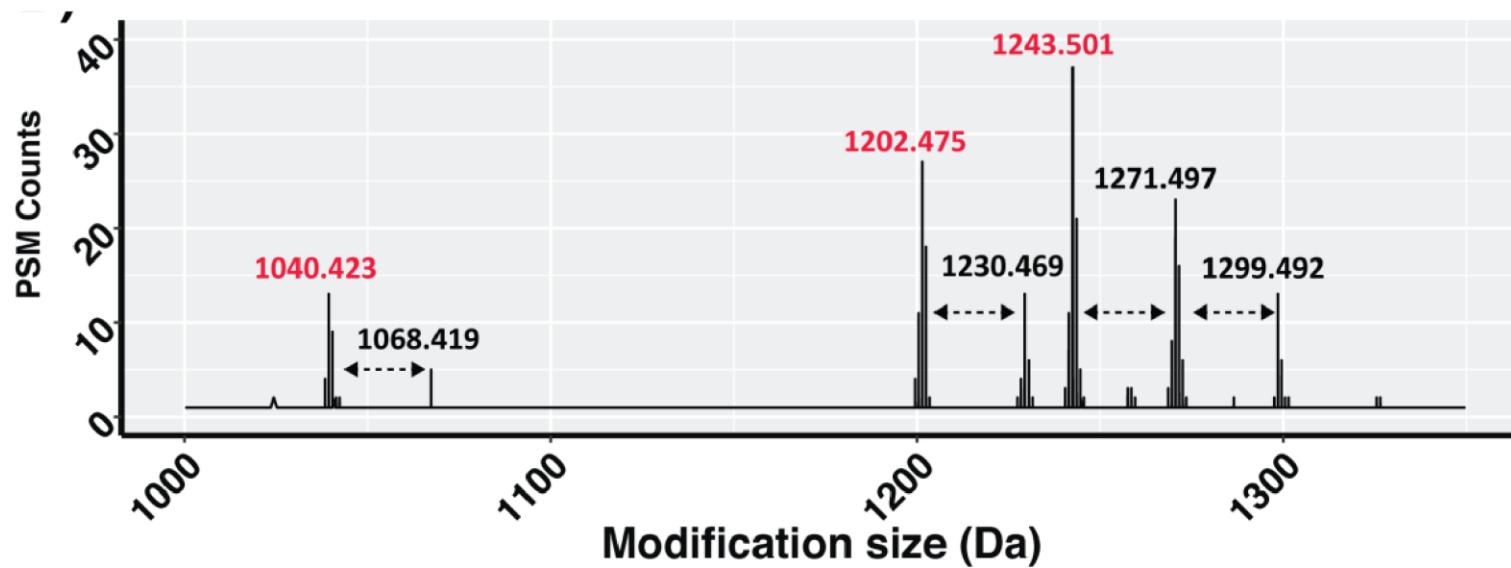


# OPEN MOD SEARCHES WITH BYONIC

**MCP** | MOLECULAR  
& CELLULAR  
PROTEOMICS

Open database searching enables the identification and comparison of bacterial glycoproteomes without defining glycan compositions prior to searching

Ameera Raudah Ahmad Izaham<sup>1</sup> and Nichollas E. Scott<sup>1\*</sup>



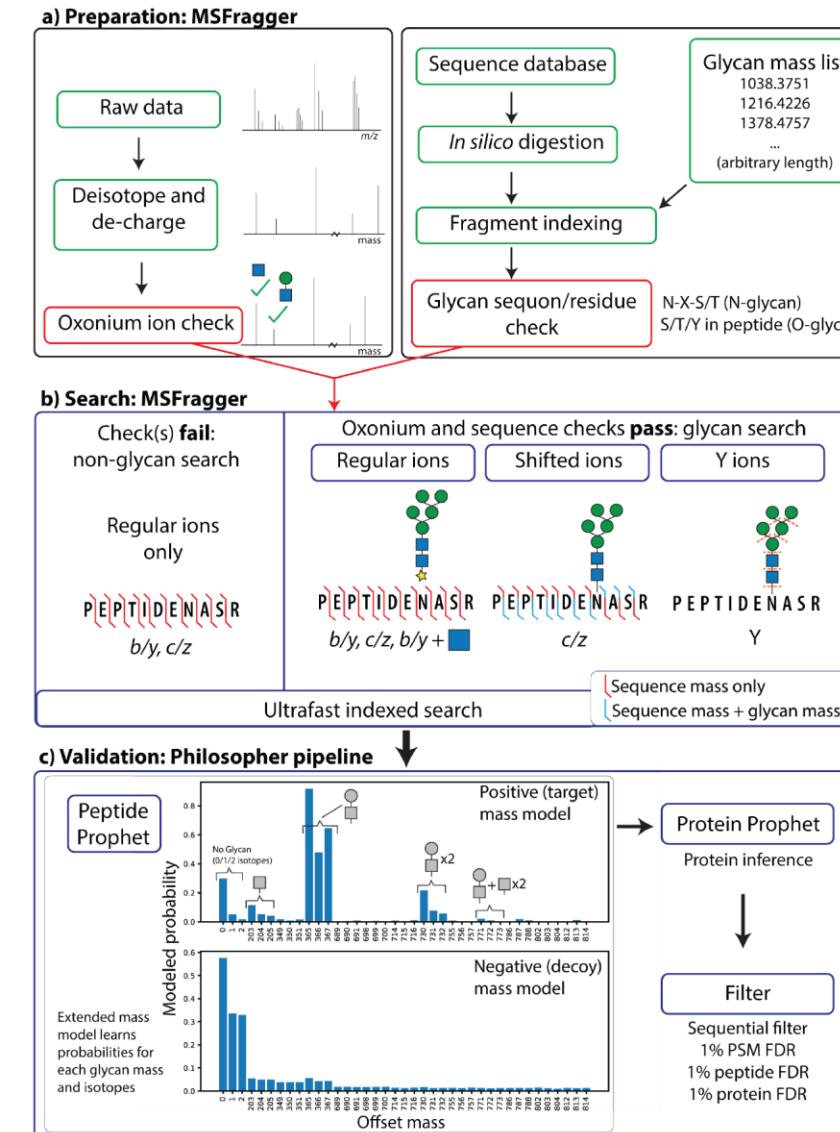
# OPEN MOD SEARCHES WITH OTHER PLATFORMS



THE PREPRINT SERVER FOR BIOLOGY

## Fast and Comprehensive N- and O-glycoproteomics analysis with MSFragger-Glyco

Daniel A. Polasky<sup>1</sup>, Fengchao Yu<sup>1</sup>, Guo Ci Teo<sup>1</sup>, Alexey I. Nesvizhskii<sup>\*1,2</sup>



# OPEN MOD SEARCHES WITH OTHER PLATFORMS



THE PREPRINT SERVER FOR BIOLOGY

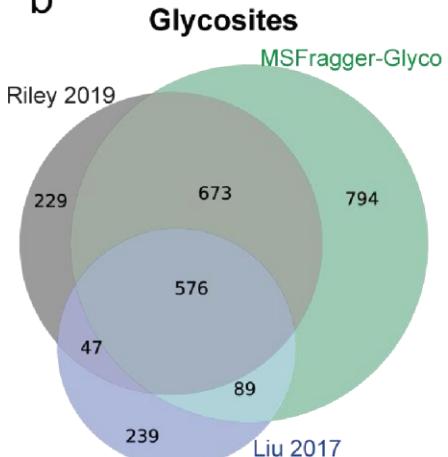
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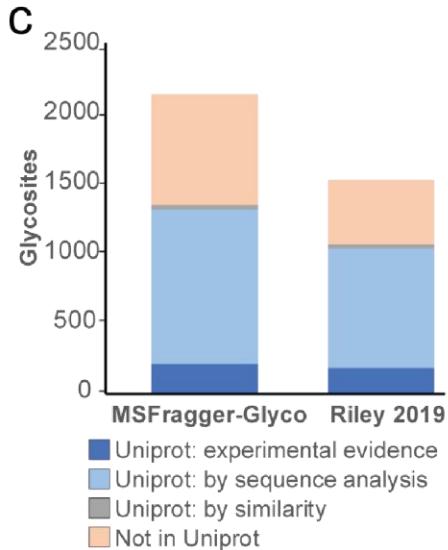
a

Search	Compositions searched	GlycoPSMs	Unique glycopeptides	Unique glycoproteins	Unique glycosites
MSFagger	182	44,187	2,822	1,070	2,133
Riley 2019	182	24,099	1,803	771	1,545

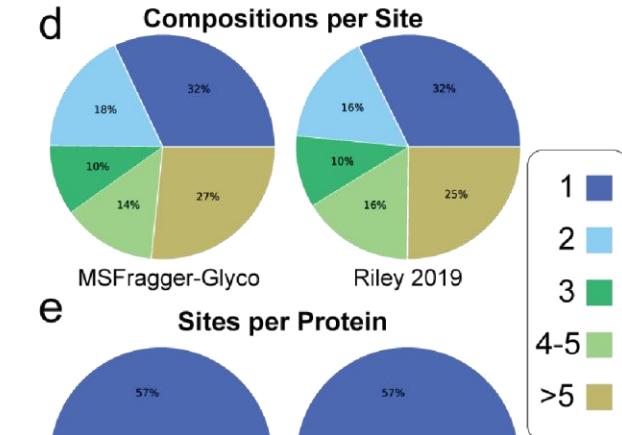
b



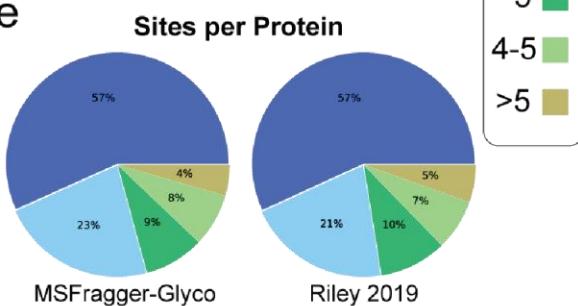
c



d Compositions per Site



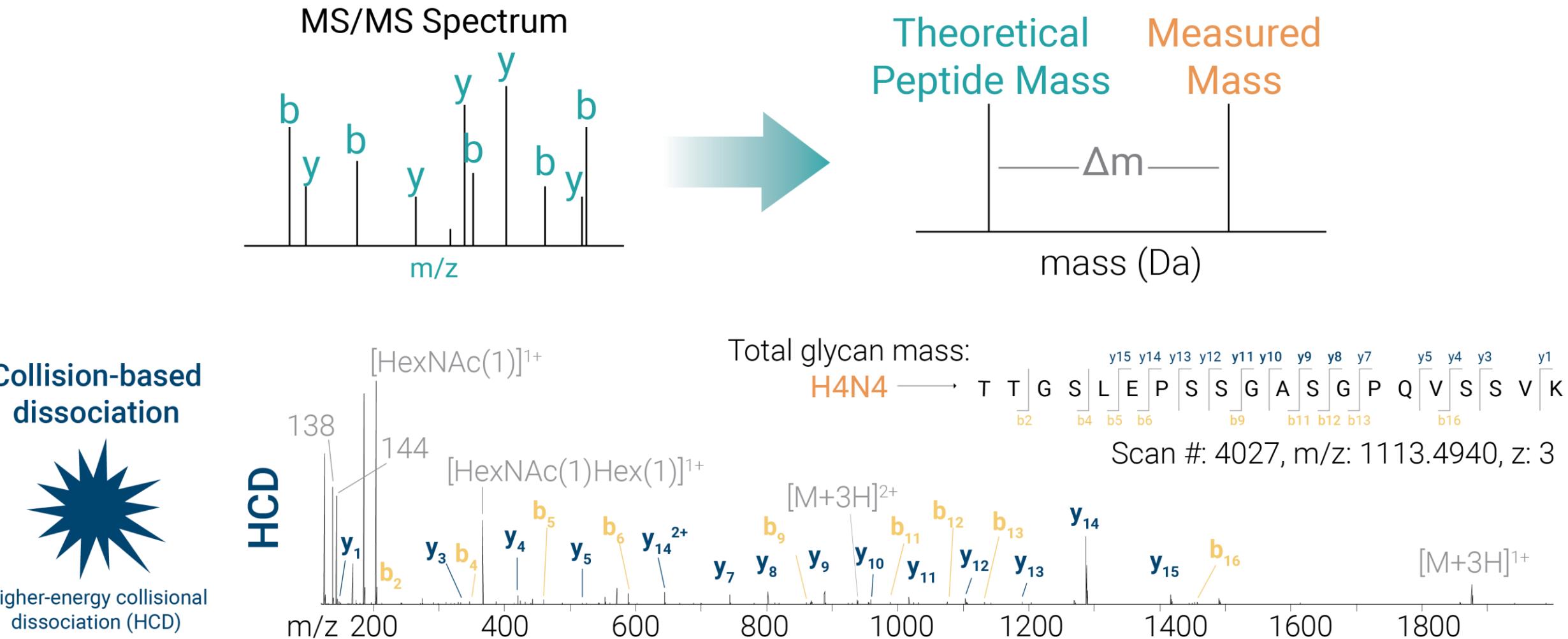
e



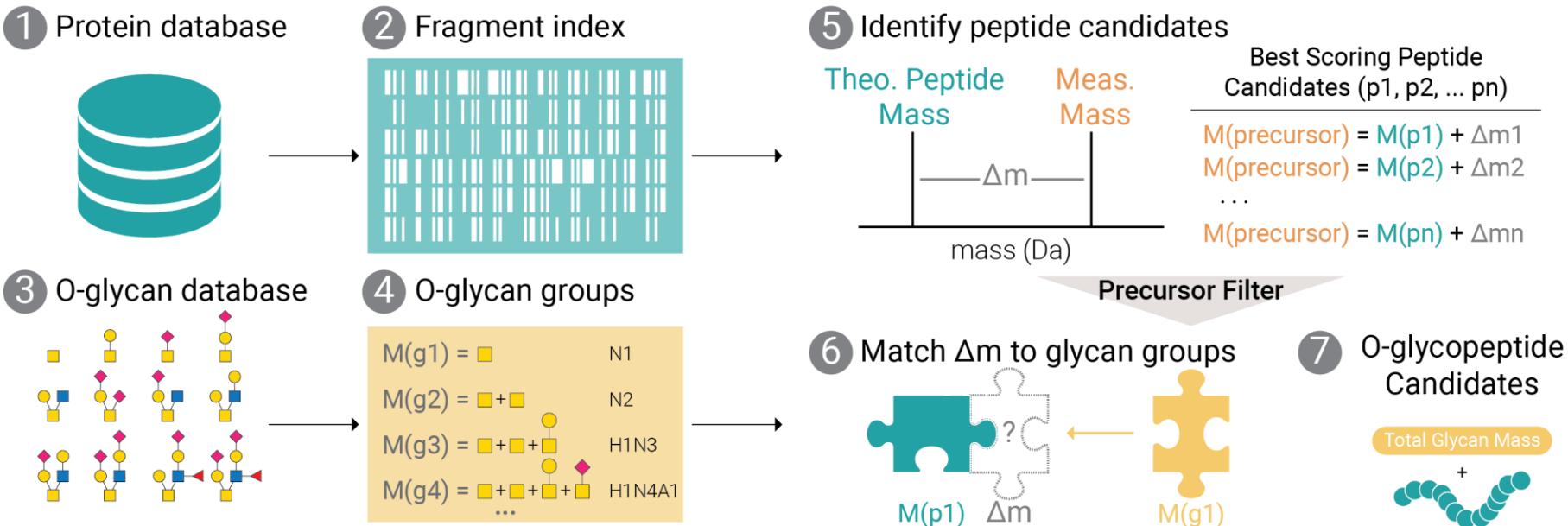
- 1
- 2
- 3
- 4-5
- >5

# O-PAIR SEARCH FOR O-GLYCOPEPTIDES

Identify peptide candidates without pre-determined modifications



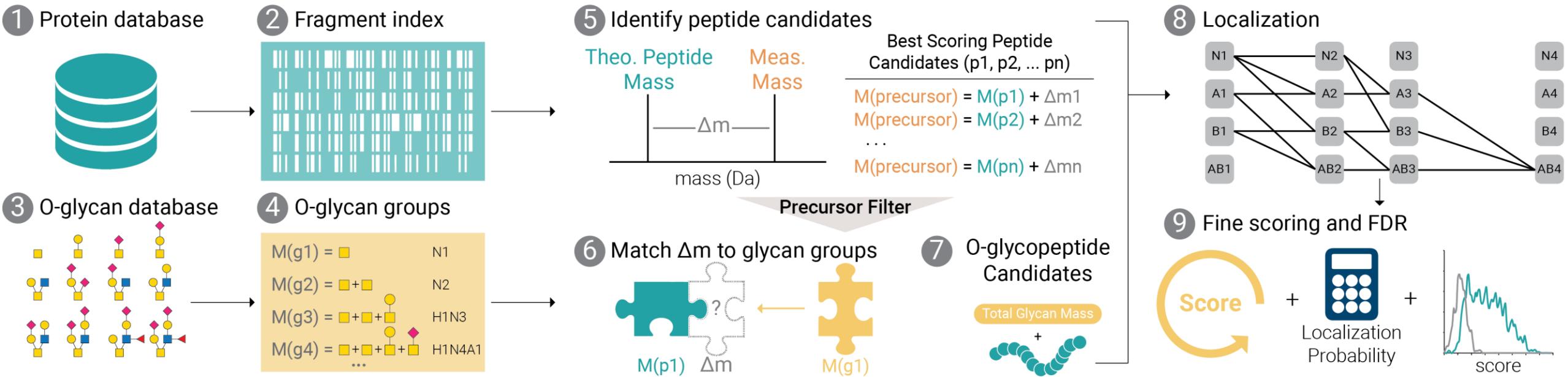
# O-PAIR SEARCH FOR O-GLYCOPEPTIDES



# O-PAIR SEARCH FOR O-GLYCOPEPTIDES

## O-Pair Search

Combining fragment-indexed Open Search with glycan group delta masses and graph theory localization

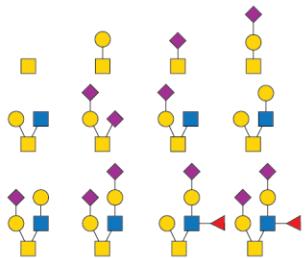


More information:

bioRxiv Pre-Print: <https://www.biorxiv.org/content/10.1101/2020.05.18.102327v1>: provisionally accepted at Nature Methods

# O-PAIR SEARCH PERFORMANCE

12 O-glycan DB



Mucins

MUC16

CD43

Podocalyxin  
PSGL-1

Riley et al., JPR, 19 (8):  
3286–3301 (2020)

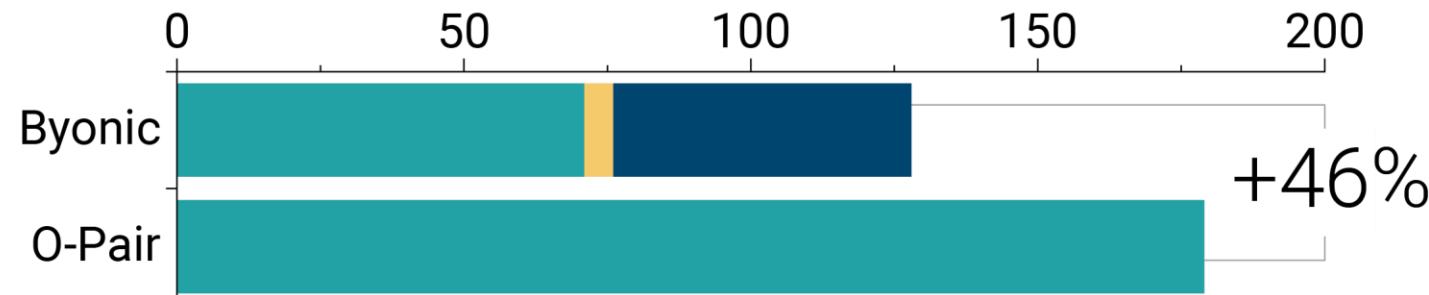
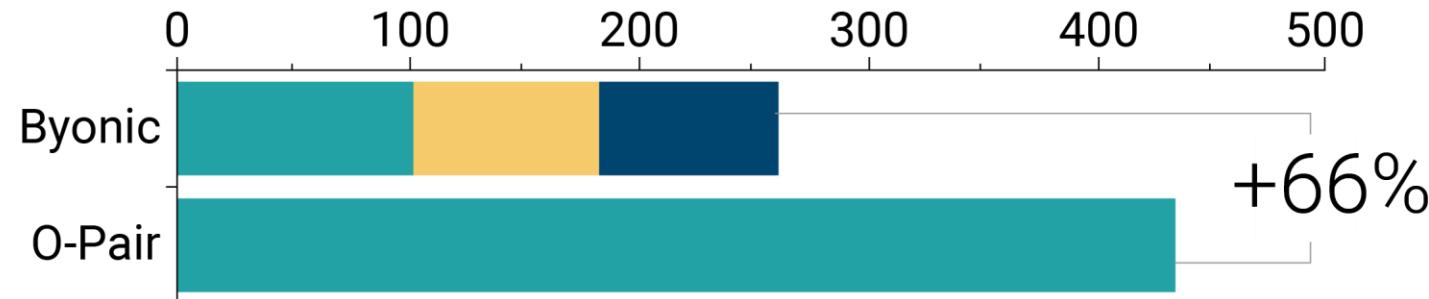
█ HCD-EThcD pair

█ HCD alone

█ EThcD alone

Glycans per peptide	Byonic Time (min)	O-Pair Time (min)	Speed Improvement	Localized Glycosites (Byonic / O-Pair)
2glycans	16.26	0.36	<b>45.24</b>	<span style="color: #FFC107;">33</span> / <span style="color: #00AEEF;">76</span>
3glycans	862.76	0.40	<b>2,161.09</b>	<span style="color: #FFC107;">41</span> / <span style="color: #00AEEF;">120</span>
4glycans	DNF	0.52	NA	<span style="color: #FFC107;">NA</span> / <span style="color: #00AEEF;">134</span>

GlycoPSMs



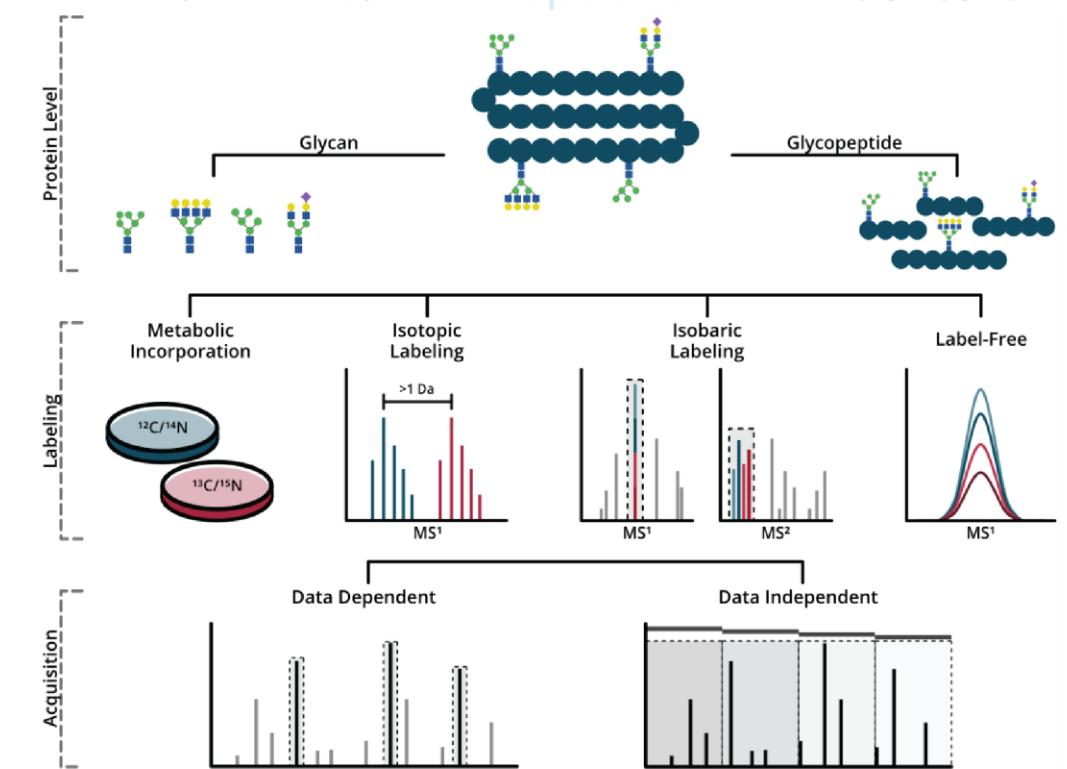
# GLYCOPEPTIDE QUANT



**MCP** | MOLECULAR & CELLULAR PROTEOMICS

## Recent Advances in Analytical Approaches for Glycan and Glycopeptide Quantitation

Daniel G. Delafield<sup>1</sup> and Lingjun Li<sup>1,2,\*</sup>



# GLYCOPEPTIDE QUANT: NOT STRAIGHTFORWARD



Isolating  
Glycoproteins



Generating  
Glycopeptides



Enrichment &  
Separations

LC-MS/MS



Ionization



Tandem MS  
(MS/MS)



Interpreting  
Spectra

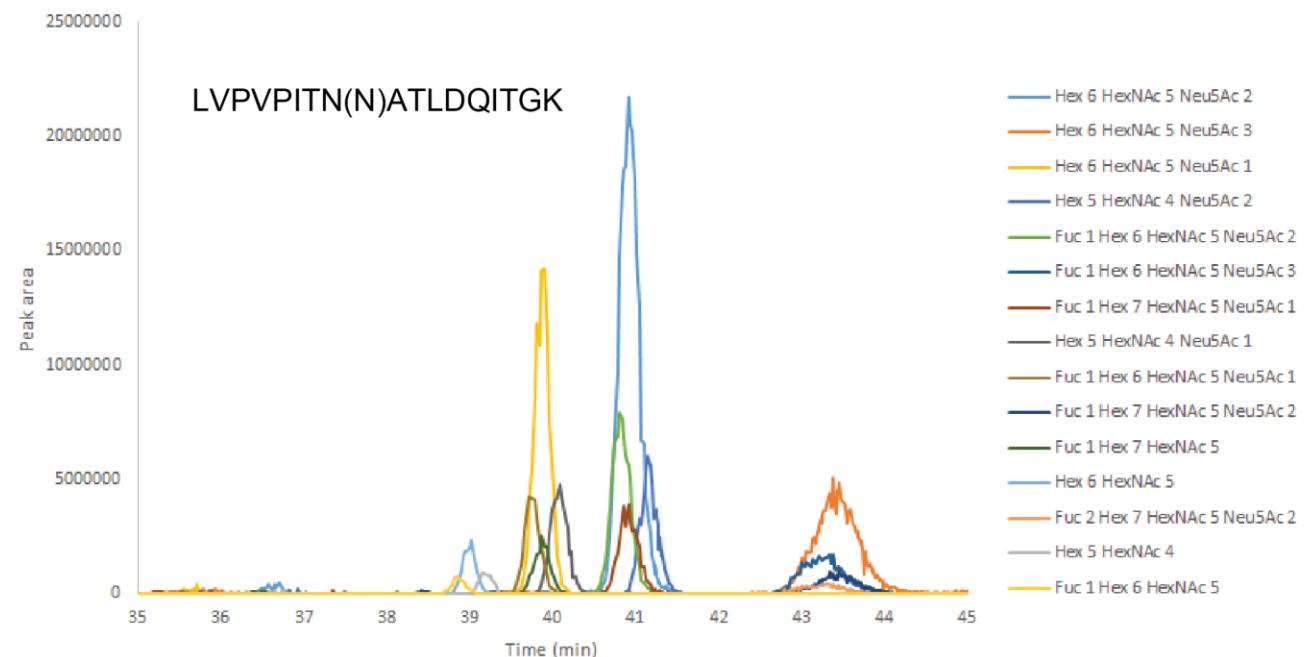


Quant and  
Context



Calculating glycoprotein similarities from mass spectrometric data

William Hackett<sup>1</sup> and Joseph Zaia<sup>1,2</sup>



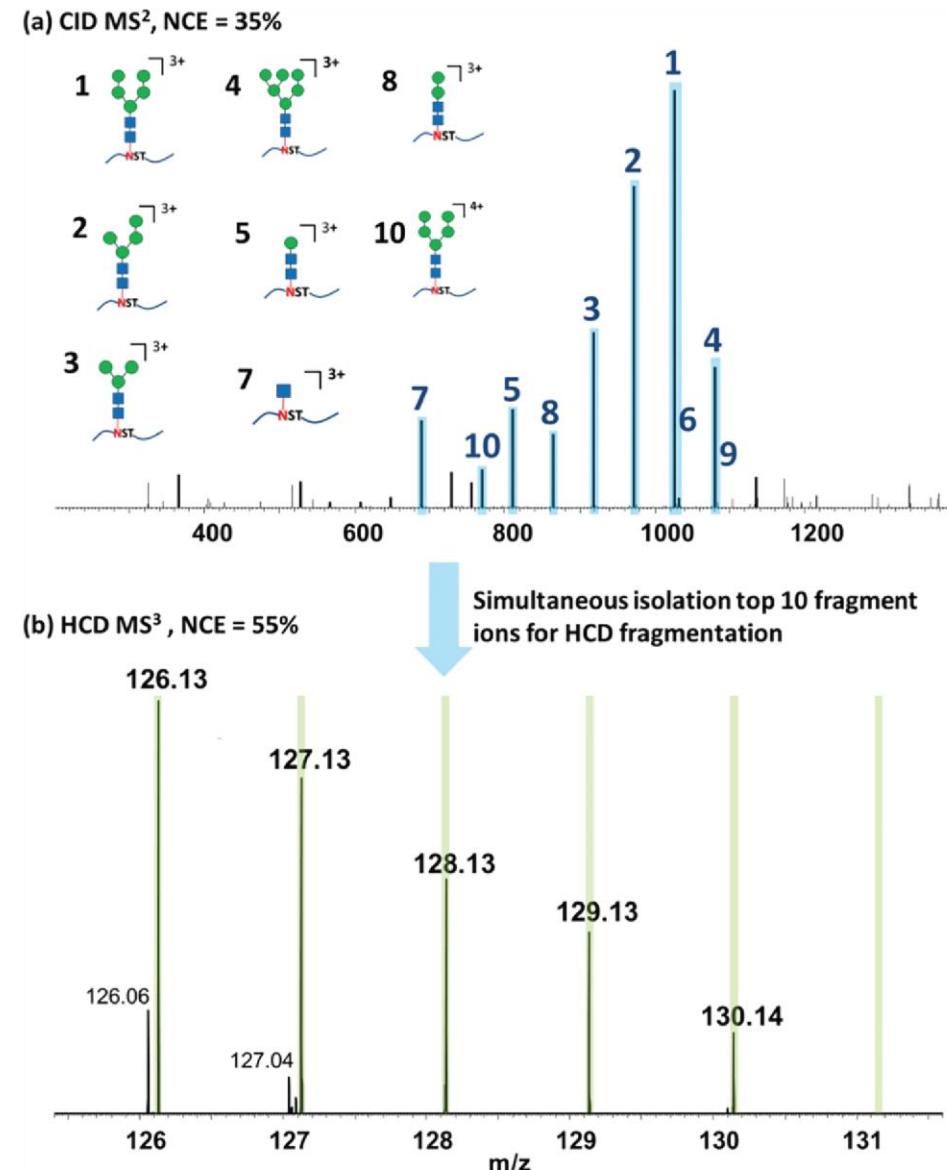
# ISOBARIC LABELING ALSO HAS CHALLENGES

## Multiplexed Comparative Analysis of Intact Glycopeptides Using Electron-Transfer Dissociation and Synchronous Precursor Selection Based Triple-Stage Mass Spectrometry

Hongbin Zhu,\* Chen Qiu, Connie M. Gryniewicz-Ruzicka, David A. Keire, and Hongping Ye

Cite This: *Anal. Chem.* 2020, 92, 7547–7555

Read Online

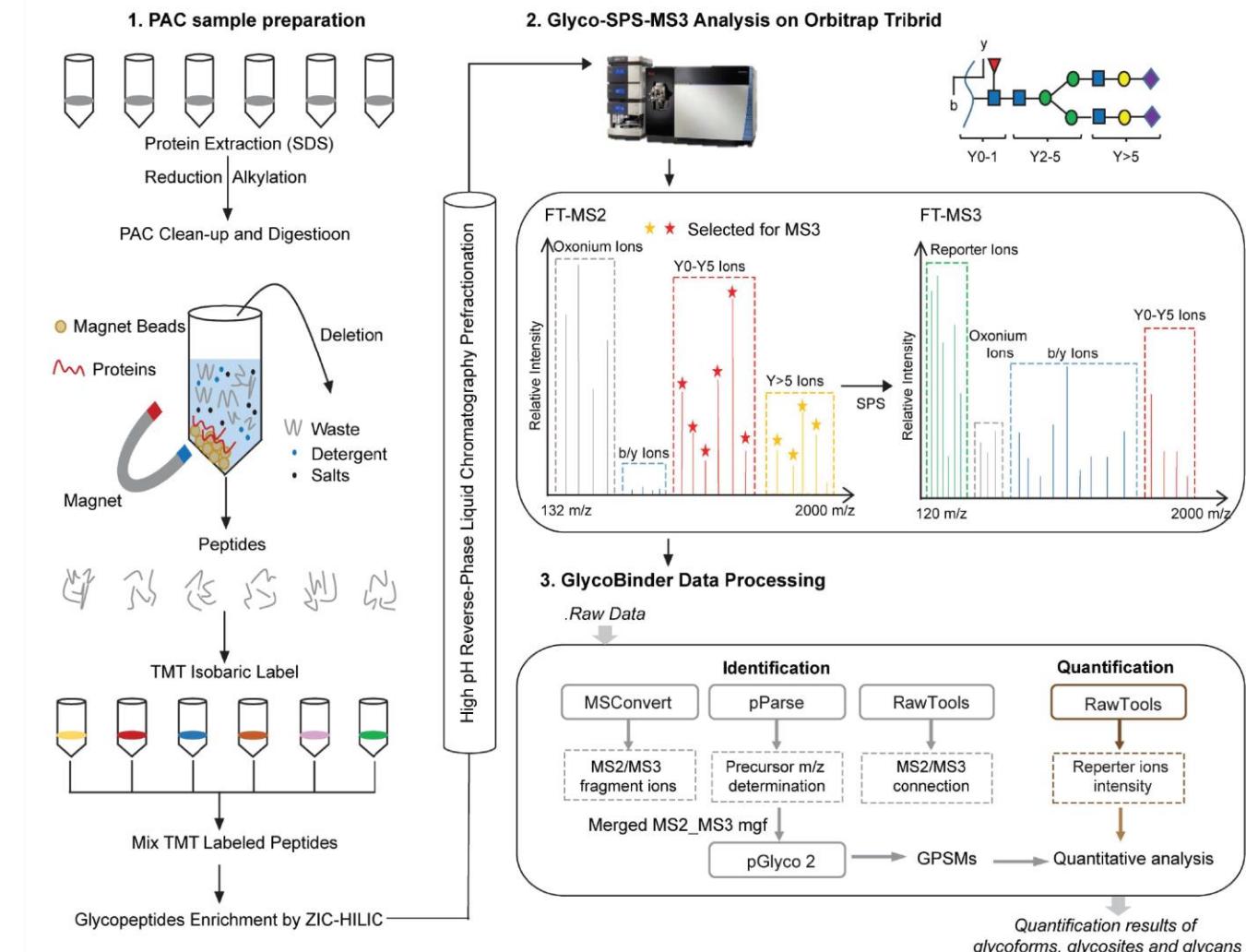


# ISOBARIC LABELING ALSO HAS CHALLENGES



## SugarQuant: a streamlined pipeline for multiplexed quantitative site-specific N-glycoproteomics

Pan Fang<sup>1\*</sup>, Yanlong Ji<sup>1, 2, §\*</sup>, Ivan Silbern<sup>1, 3</sup>, Carmen Doebele<sup>2</sup>, Momchil Ninov<sup>1</sup>, Christof Lenz<sup>1, 3</sup>, Thomas Oellerich<sup>2, 4, 5</sup>, Kuan-Ting Pan<sup>1, §, ¶</sup>, Henning Urlaub<sup>1, 3, ¶</sup>



# HOW TO REPRESENT DIMENSIONALITY?

## Quantitative Longitudinal Inventory of the *N*-Glycoproteome of Human Milk from a Single Donor Reveals the Highly Variable Repertoire and Dynamic Site-Specific Changes

Jing Zhu,<sup>1</sup> Yu-Hsien Lin,<sup>1</sup> Kelly A. Dingess, Marko Mank, Bernd Stahl, and Albert J. R. Heck\*

Cite This: *J. Proteome Res.* 2020, 19, 1941–1952

Read Online



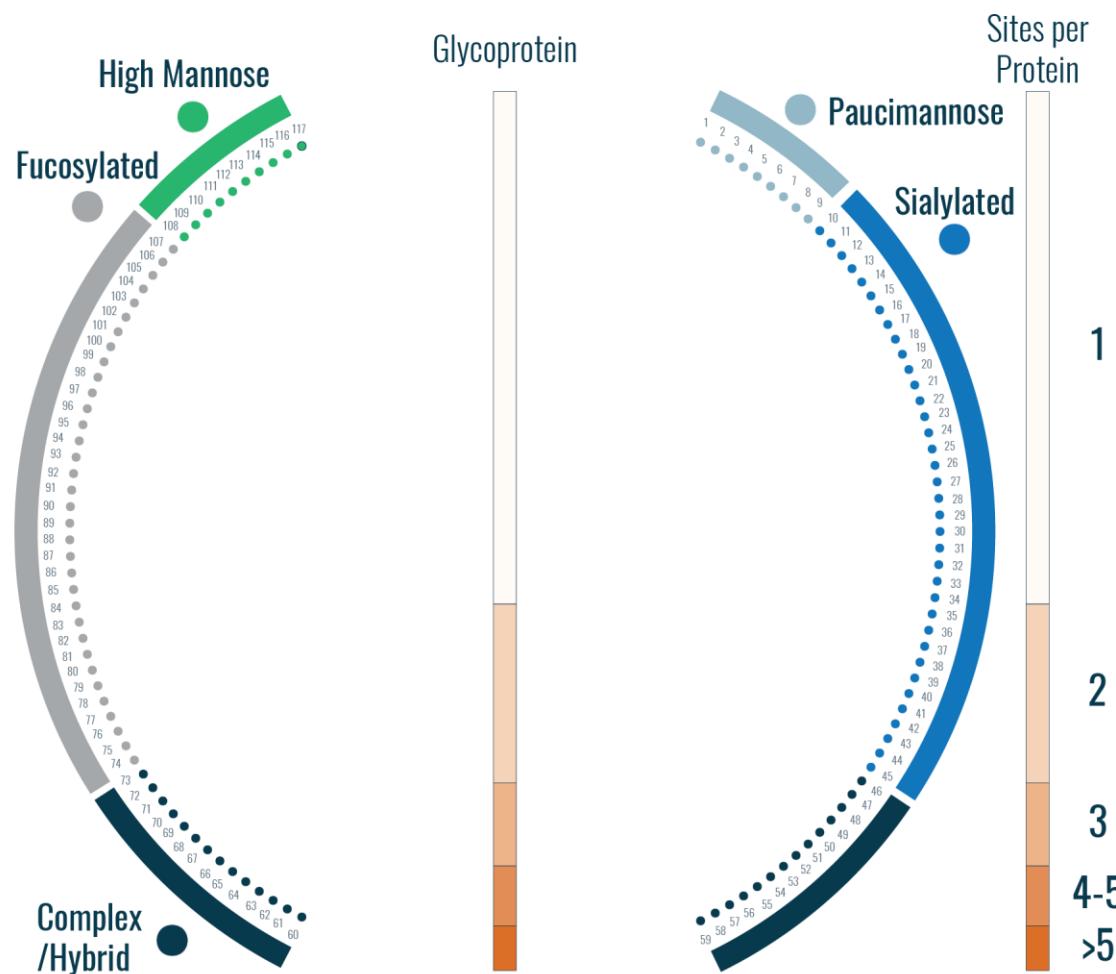


# HOW TO REPRESENT DIMENSIONALITY?

Capturing site-specific heterogeneity with large-scale N-glycoproteome analysis

Nicholas M. Riley<sup>1,2,5</sup>, Alexander S. Hebert<sup>1</sup>, Michael S. Westphall<sup>1</sup> & Joshua J. Coon<sup>1,2,3,4</sup>

## Glycoprotein-Glycan Networks



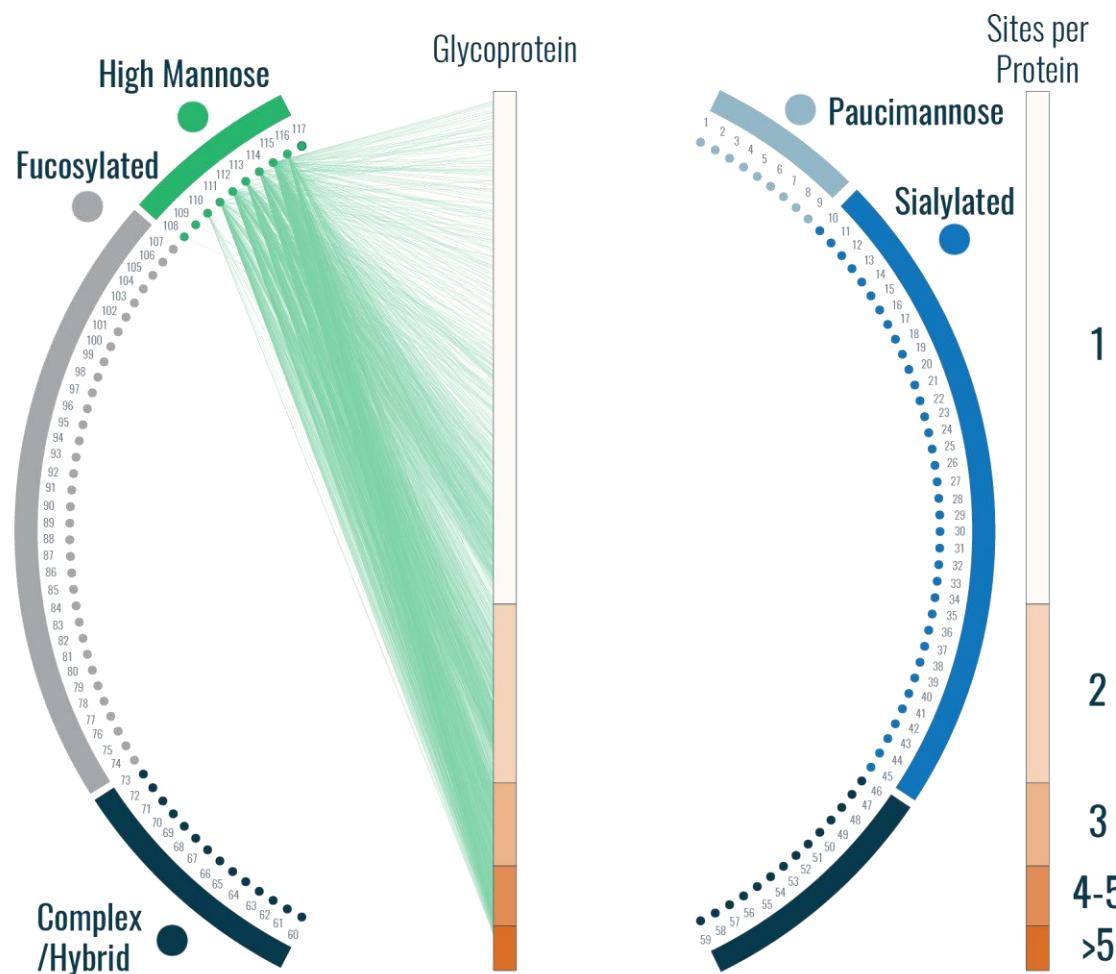


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Glycoprotein-  
Glycan Networks



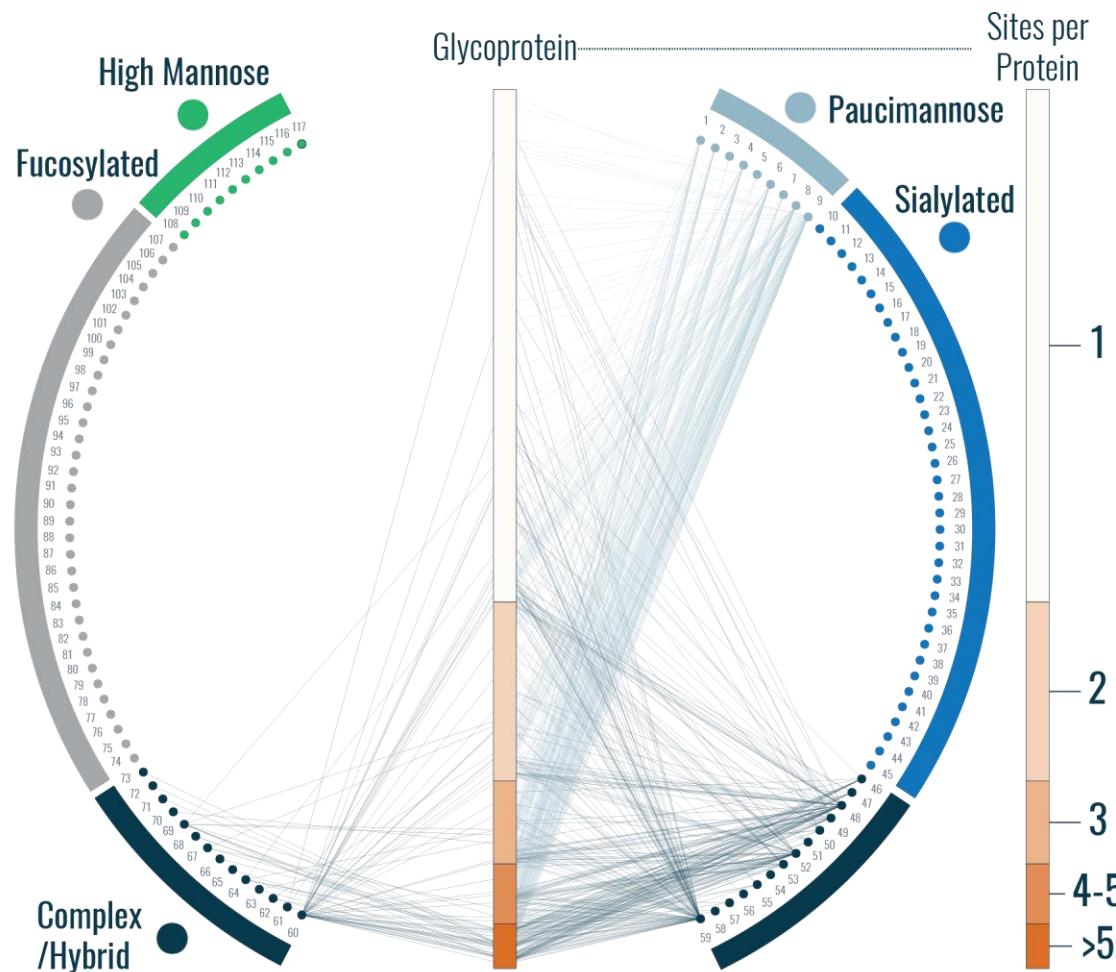


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Glycoprotein-  
Glycan Networks



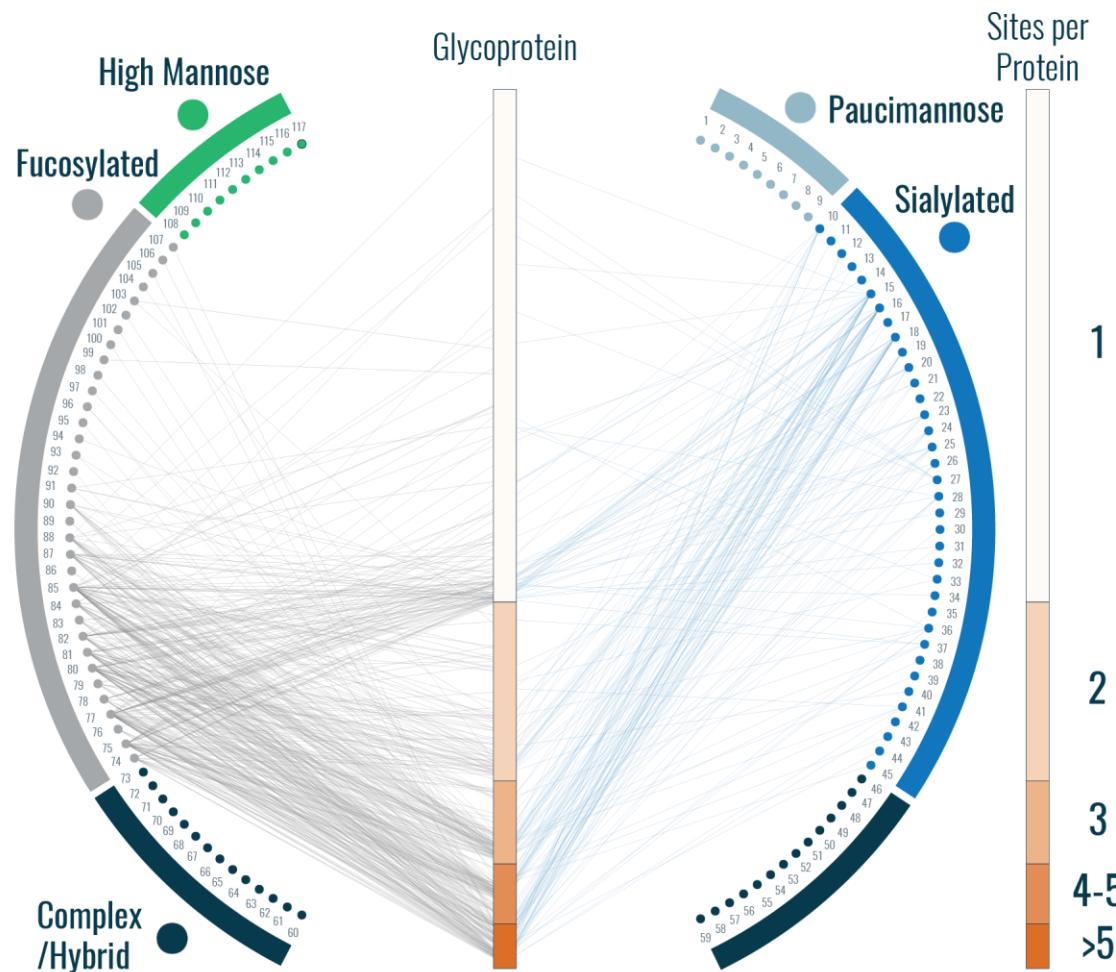


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Glycoprotein-  
Glycan Networks



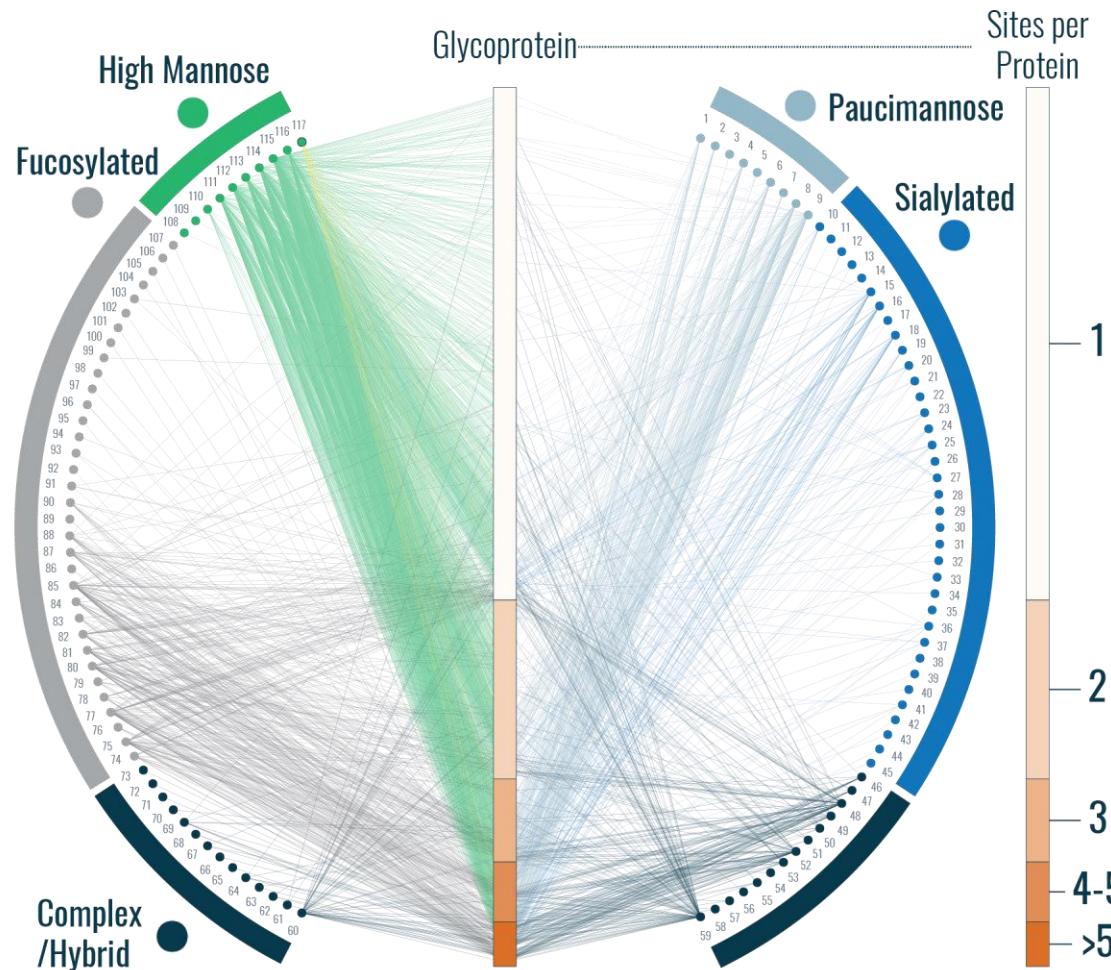


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Capturing site-specific heterogeneity with large-scale N-glycoproteome analysis

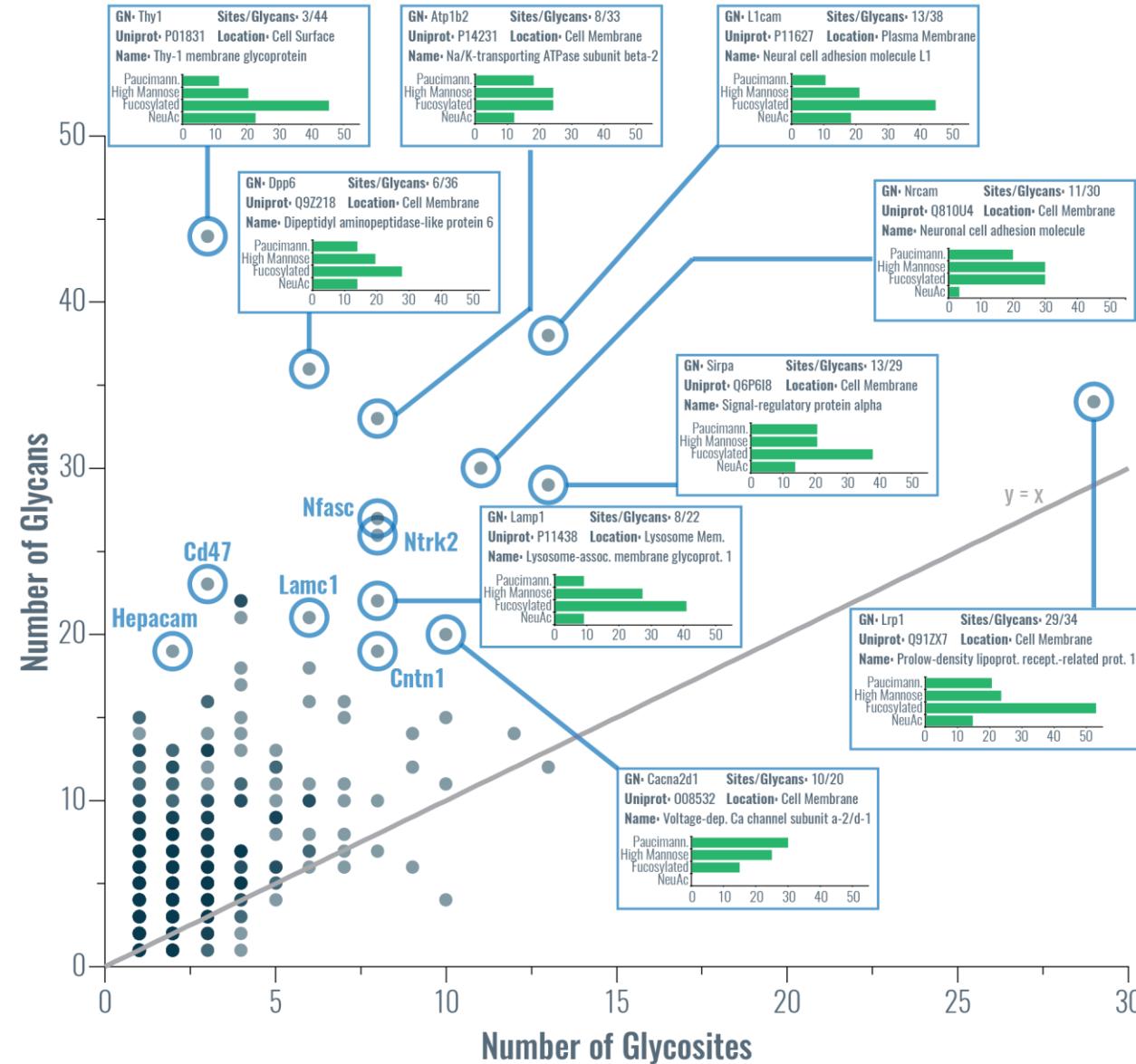
Nicholas M. Riley<sup>1,2,5</sup>, Alexander S. Hebert<sup>1</sup>, Michael S. Westphall<sup>1</sup> & Joshua J. Coon<sup>1,2,3,4</sup>

Glycoprotein-  
Glycan Networks





# HOW TO REPRESENT DIMENSIONALITY?



# HOW TO REPRESENT DIMENSIONALITY?

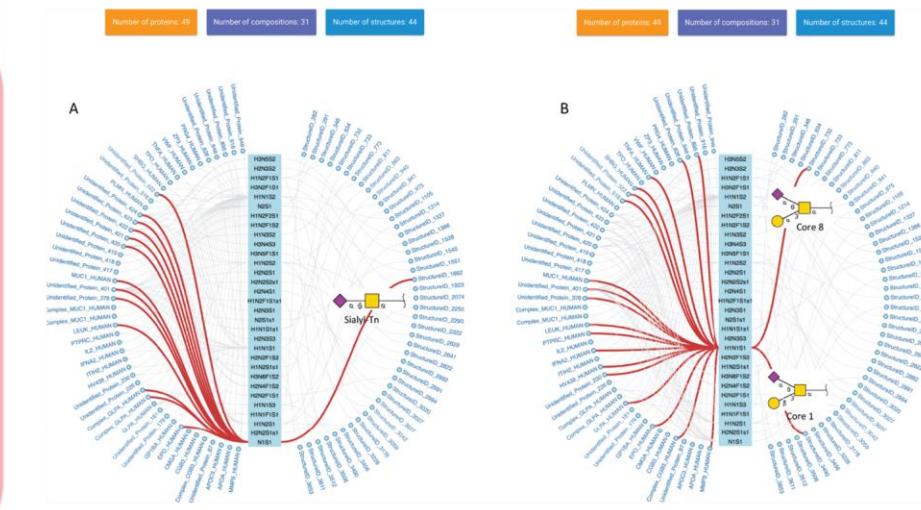
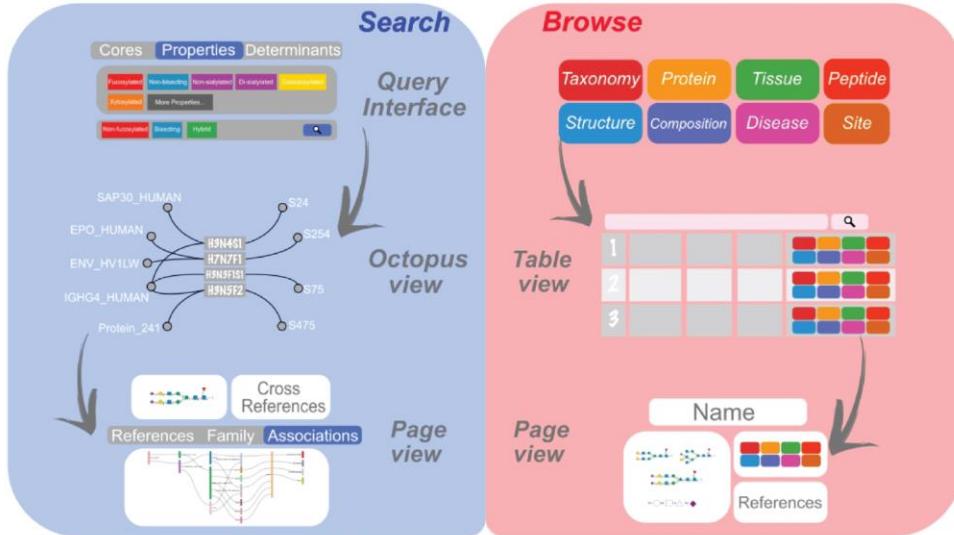
Journal of  
**proteome**  
research

Cite This: *J. Proteome Res.* 2019, 18, 664–677

Article

pubs.acs.org/jpr

Inside GlyConnect





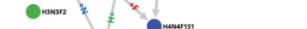
# HOW TO REPRESENT DIMENSIONALITY?

## Examining and Fine-tuning the Selection of Glycan Compositions with GlyConnect Compozitor

Thibault Robin<sup>1,2,3,4</sup>, Julien Mariethoz<sup>1,2</sup> , and Frédérique Lisacek<sup>1,2,5,\*</sup>

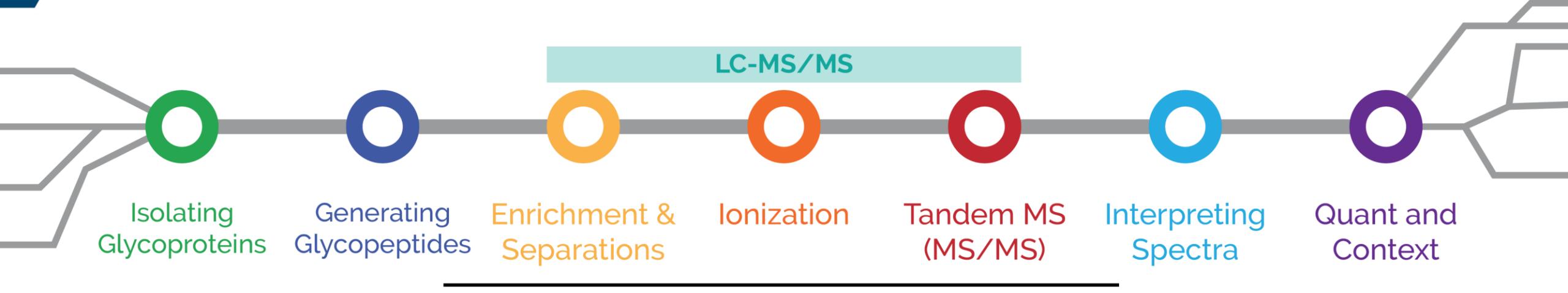
### GlyConnect Compozitor

- 12 (A) Homo sapiens | Integrin alpha-5 | N-Linked | Asn-182,Asn-297,Asn-518,Asn-672,Asn-771,Asn-868
- 8 (B) Homo sapiens | Integrin alpha-5/beta-1 | N-Linked | Asn-97,Undefined
- 31 (C) Homo sapiens | Integrin beta-1 | N-Linked | Asn-50,Asn-212,Asn-406,Asn-481,Asn-520,Asn-669
- 4 (AB)
- 1 (AC)
- 7 (BC)
- 4 (ABC)





# CONCERTED EFFORT FOR INFORMATICS



Letter to Glyco-Forum

## The GlySpace Alliance: toward a collaborative global glycoinformatics community

Kiyoko F Aoki-Kinoshita<sup>1</sup>, Frederique Lisacek<sup>2</sup>, Raja Mazumder<sup>3</sup>, William S York<sup>4</sup> and Nicolle H Packer<sup>5</sup>

<sup>1</sup>Glycan & Life Science Integration Center (GaLSIC), Faculty of Science and Engineering, Soka University, 1-236 Tangi-machi, Hachioji, Tokyo, Japan, 192-8577, <sup>2</sup>Proteome Informatics Group, SIB Swiss Institute of Bioinformatics, Computer Science Department, University of Geneva, route de Drize 7, CH - 1227 Geneva Switzerland, and also Section of Biology, University of Geneva, Geneva, Switzerland, <sup>3</sup>Department of Biochemistry & Molecular Medicine, and Department of Medicine, School of Medicine and Health Sciences, George Washington University, Ross Hall, 2300 Eye St., NW, Washington, DC 20037, USA, <sup>4</sup>Complex Carbohydrate Research Center, University of Georgia, 315 Riverbend Road, Athens, GA 30602, USA, and <sup>5</sup>Department of Molecular Sciences, Faculty of Science & Engineering, Rm 307, Building E8C, Macquarie University, Sydney, NSW 2109, Australia

<sup>1</sup>To whom correspondence should be addressed: Tel/Fax: +81-42-691-4116; e-mail: kkiyoko@soka.ac.jp

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SUMS



Wed-Thurs October 28-29, 2020  
SUMS Research Application Symposium

