

# STANFORD UNIVERSITY MASS SPECTROMETRY

VINCENT COATES FOUNDATION MASS SPECTROMETRY LABORATORY



## SUMS In-Gel digest protocol

1. Cut gel pieces in 1mm x 1mm squares.
2. Ensure gel pieces are at neutral pH by adding 50-100  $\mu$ l 100mM Ammonium bicarbonate, let sit for 10 minutes and discard)
3. Add 10  $\mu$ l 50mM dithiothreitol (DTT) and 100  $\mu$ l 50mM Ammonium bicarbonate
4. Incubate at 55 °C for 30 minutes.
5. Discard solution phase, let temperature of eppendorf's adjust to room temperature
6. Add 10  $\mu$ l 100mM acrylamide and 100  $\mu$ l 50mM Ammonium bicarbonate (pH 8-8.2)
7. Incubate at room temperature for 30 minutes.
8. Discard solution phase
9. Wash gel pieces with 100-150  $\mu$ l 50mM Ammonium bicarbonate/ 50% acetonitrile for 10 minutes, vortex every 5 minutes. (Disregard this step for Silver stained samples)
  - a. Depending on intensity of stain, repeat step 9 until the gel pieces are clear.
  - b. If after doing this twice and the stain has not dissipated and 100  $\mu$ l of 50mM Ammonium bicarbonate for 1 min, then discard. Repeat step 6.
10. Discard solution phase and dry samples in speed vac for 5-10 minutes.
11. Add 5pmol promega sequencing grade trypsin in 50mM Ammonium bicarbonate, 0.02% Protease Max, to each sample and spin down.
  - a. Add 200  $\mu$ l of 50 mM Ammonium bicarbonate to 1 Protease max stock vial.
  - b. Add 75  $\mu$ l of the newly diluted Protease Max stock to the Trypsin stock.
    1. **Non-Silver Stained**  
10  $\mu$ l of diluted Trypsin stock will be 5pmol trypsin.  
Add 10  $\mu$ l of diluted Trypsin stock to each eppendorf for in-gel digestion.
    2. **Silver Stained**  
Add 1.6  $\mu$ l of diluted Trypsin to each eppendorf for in-gel digestion
  - c. Add a minimal amount of 0.02% Protease Max diluted in 50mM Ammonium bicarbonate (typically 15-40  $\mu$ l) just to cover gel pieces.
12. Incubate overnight in 37°C oven (or Incubate 1-2 hour at 50° C)
13. Spin Down and pull off all liquid, and transfer to a newly labeled mini-eppendorf (0.7mL).
14. Add 40-50  $\mu$ l 1% formic acid, 66% acetonitrile 33% 100mM Ammonium bicarbonate
15. Incubate for 10 minutes at 37° C.
16. Spin at 10,000G for 2 minutes
17. Extract all solution
18. Speed vac total combined extract to dryness. Save gel pieces & store at -4° C until analysis is confirmed.

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**Supplies:**

- Promega – Trypsin/Lys-C Mix, Mass Spec Grade, 20 µg V5071 (20µg) or V5073 (5x20µg)  
[http://www.promega.com/products/protein-expression-and-mass-spectrometry/peptidases-and-services-for-mass-spectrometry/trypsin\\_lys\\_c-mix\\_-mass-spec-grade/](http://www.promega.com/products/protein-expression-and-mass-spectrometry/peptidases-and-services-for-mass-spectrometry/trypsin_lys_c-mix_-mass-spec-grade/)
- Promega - ProteaseMAX™ Surfactant, Trypsin Enhancer, 1mg, V2071 (1mg) or V2072 (5x1mg)  
[http://www.promega.com/products/protein-expression-and-mass-spectrometry/peptidases-and-services-for-mass-spectrometry/proteasemax-surfactant\\_-trypsin-enhancer/](http://www.promega.com/products/protein-expression-and-mass-spectrometry/peptidases-and-services-for-mass-spectrometry/proteasemax-surfactant_-trypsin-enhancer/)
- 50mM DTT
- 100mM Acrylamide
- 50mM Ammonium Bicarbonate
- 100mM Ammonium Bicarbonate