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MASS SPECTROMETRY IN METABOLOMICS

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Stanford Mass Spectrometry
Users' Meeting
August 21, 2008



Origin of Metabolomics

Proc. Nat. Acad. Sci. USA

Vol. 68, No. 10, pp. 2374–2376, October 1971

Quantitative Analysis of Urine Vapor and Breath by Gas–Liquid Partition Chromatography

(orthomolecular medicine/vitamins/controlled diet)

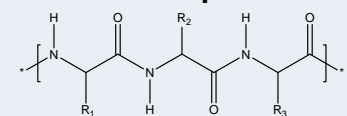
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Contributed by Linus Pauling, July 29, 1971



Metabolomics and Proteomics

	Proteomics	Metabolomics
Quantitation	Under development SILAC, AQUA, iTRAQ Relative quantitation (x fold)	Well established Classical Analytical Chemistry Absolute (nM, pg/mL) or relative (x fold) quantitation
Identification	Well established <i>De novo</i> sequencing  PTMs still a challenge	Under development Huge diversity of structures, NMR often required Moderate success with mass spectral libraries



Types of Experiments in Metabolomics

targeted

non-targeted

quantitative

semi-quantitative

- Number of analyzed metabolites is limited by the number of available **standards**

- **Absolute quantitation** of metabolites (nM, mg/mL)

- Number of analyzed metabolites is limited by the number of available **library spectra**

- **Relative quantitation** of metabolites (fold)

- Number of analyzed metabolites is limited by capacity of **analytical instrumentation**

- **Relative quantitation** of metabolites (fold)



Applications of Metabolomics

- Studies of biochemical pathways
- Observational studies (“hypothesis generating” studies)
 - Phenotyping
 - Search of diagnostic biomarkers of disease
 - Early detection of toxic effects of drug candidates



GC-MS Analysis of Metabolites: Overview

- **50-600 (300) amu mass range**
mono- and disaccharides, amino acids, fatty acids
(mostly primary metabolites)
- **Derivatization required**
- **Metabolite libraries are available** due to instrument-independent and well understood nature of electron ionization that generates extensive fragmentation and information rich spectra
- Advantageous for flux analysis using ^{13}C labeling

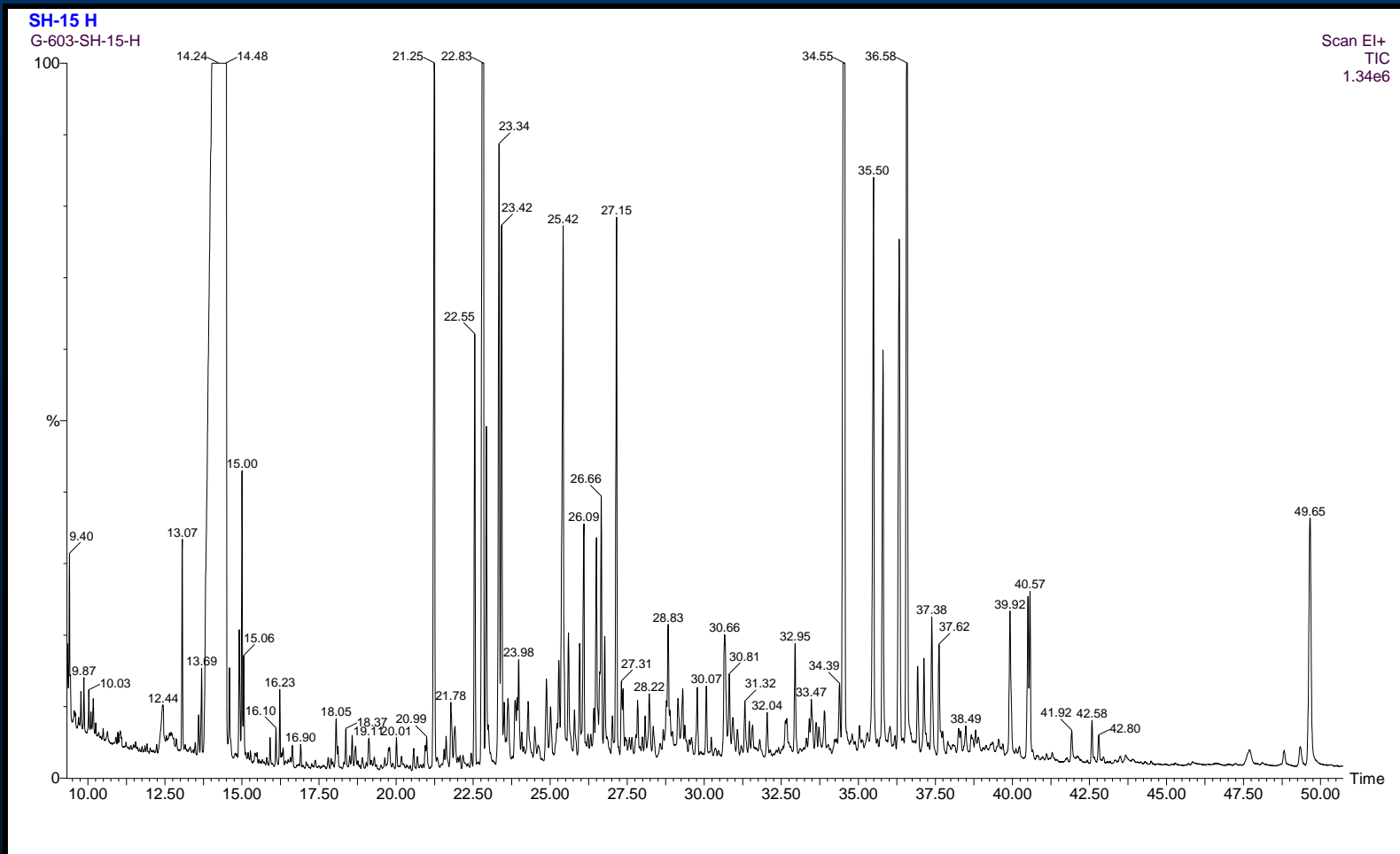


GC-MS Analysis of Metabolites: Workflow

- Sample preparation:
 - depletion of abundant metabolites (urine: urease treatment)
 - extraction, homogenization, or lyophilization
 - oximation (sugars), and silylation
- GC-MS analysis
 - disposable glass liners are preferred to eliminate carry-over
 - retention index (RI) standards can be used to aid identification
- Deconvolution of mass spectra using libraries
 - AMDIS (freeware from NIST)



GC-MS Profile of Urine





LC-MS Analysis of Metabolites: Overview

- 100-2000 amu mass range
sugars, amino acids, peptides, lipids, secondary plant metabolites
- No derivatization required
- Low efficiency of LC
especially for polar compounds
- Metabolite mass spectral libraries are missing
instrument-dependent nature of collision induced dissociation,
insufficient fragmentation
- Ultra-high resolution MS (FT ICR, Orbitrap, TOF) may aid
identification

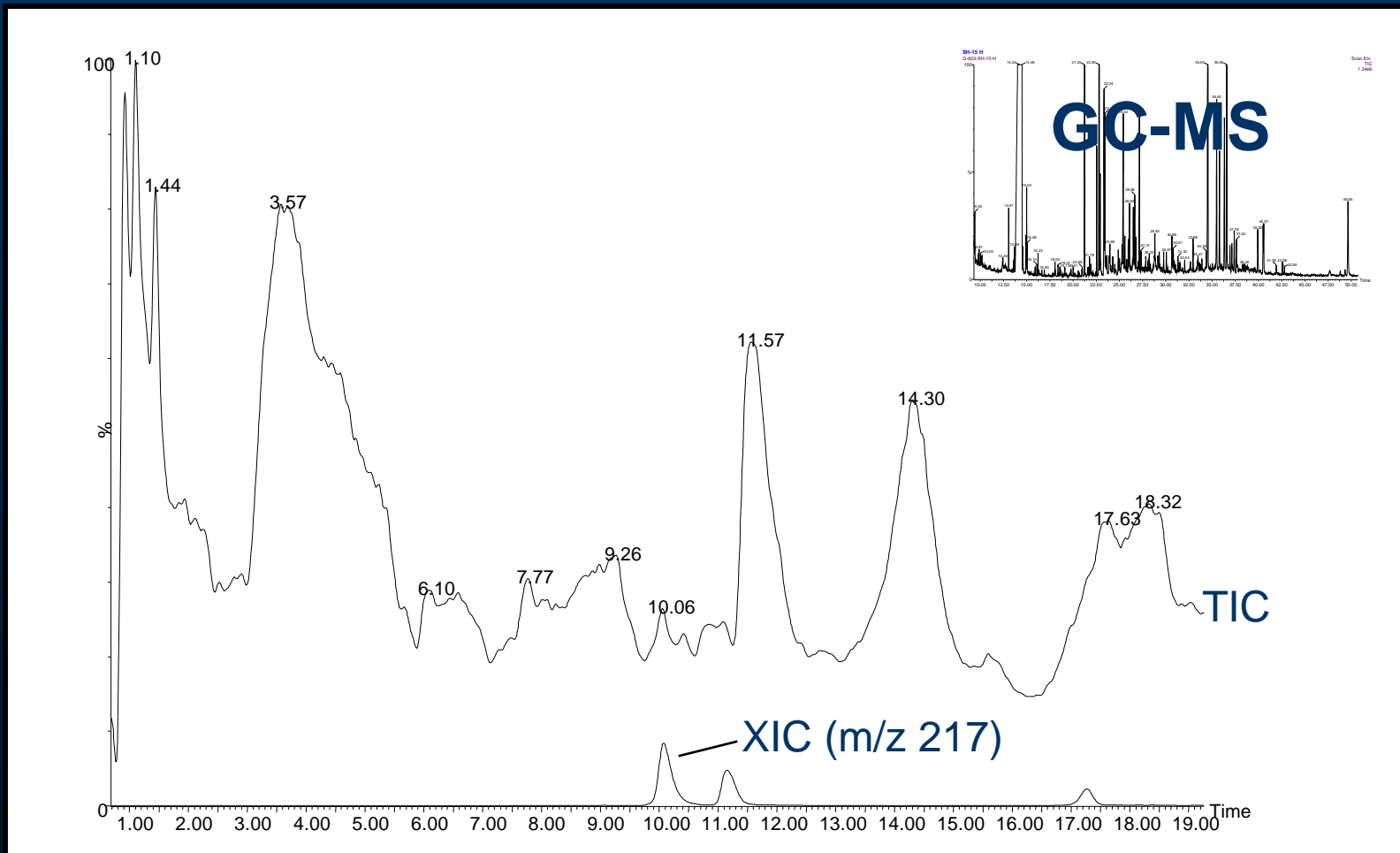


LC-MS Analysis of Metabolites: Workflow

- Sample preparation:
 - depletion of abundant metabolites (urine: urease treatment)
 - extraction or lyophilization; automation with online extraction
- LC-MS analysis
 - combination of ionization modes is preferred (ESI, APCI, +, -)
 - reverse phase LC for non-polar metabolites and hydrophilic interaction chromatography (HILIC) for polar metabolites
- Detection of spectral “features” (molecular ions) using metabolomics software
 - freeware XCMS and MZmine
- Identification based on retention time, accurate mass, and fragmentation (libraries)



HILIC-MS Profile of Urine

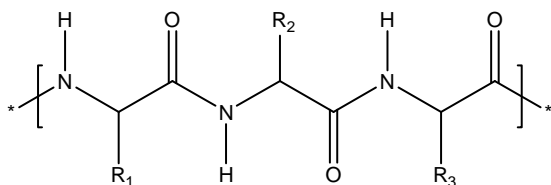




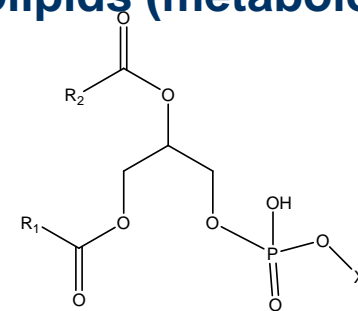
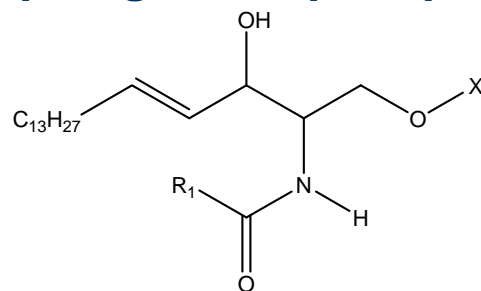
Lipid Analysis by ESI-MS

- Many classes of lipids are too heavy and non-volatile for GC separation (> 500 amu)
- Surface activity of polar lipids is advantageous for electrospray ionization (ESI)
- Some lipids have simple structure based on glycerol or sphingoid base scaffold

Peptides (proteomics)

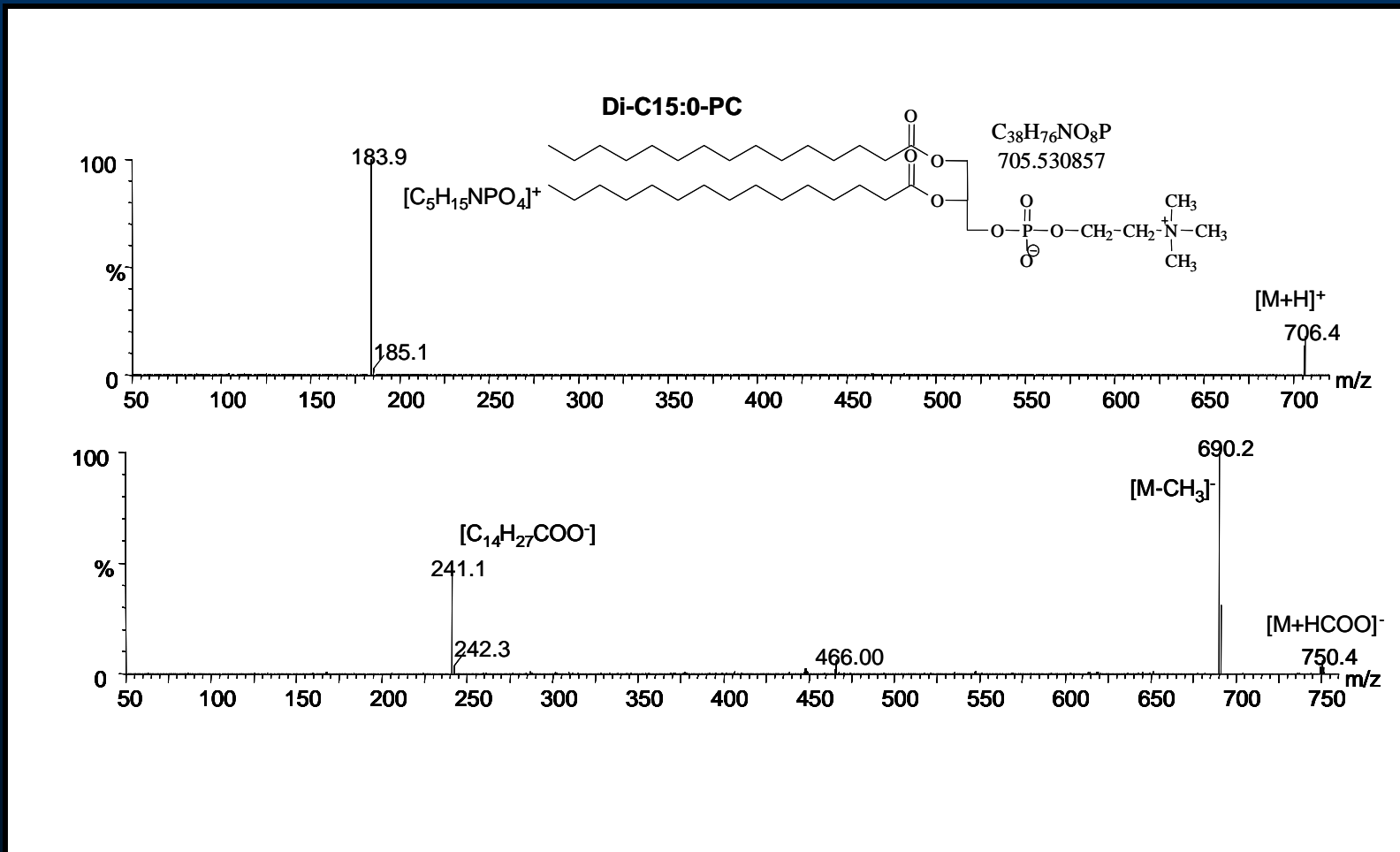


Sphingo- and phospholipids (metabolomics)





Identification of Phospholipids Using ESI-MS





Summary

- Three metabolomics platforms can be provided at SUMS in the future:
 1. Profiling of phospholipids and sphingolipids using nanoESI-MS-QTOF
 2. Profiling of primary metabolites including ^{13}C flux analysis using GC-MS
 3. Profiling of polar (HILIC) and non-polar (C18, C4) metabolites using LC-ESI-MS-QTOF
- #1 and #2 may be automated in the future to provide simple sample drop-in service



References

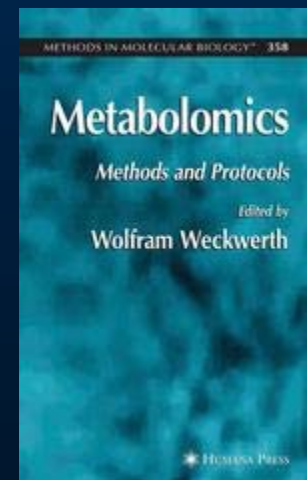
Overview:

<http://fiehnlab.ucdavis.edu/>

<http://masspec.scripps.edu/index.php>

Metabolomics: Methods and Protocols

<http://www.springerprotocols.com/BookToc/doi/10.1007/978-1-59745-244-1>





Metabolomics Webinar

Metabolomics: Analytics, Tools, and Applications (Free!)

Broadcast Date Thursday, August 28, 2008

Time 1:00 - 2:00 pm EDT

Key learning points for this webinar:

- Introduction to and definition of metabolomics
- Examples of metabolomes and where they fit into the larger biological picture
- Analytical techniques for uncovering and exploiting metabolomics—capabilities and limitations
- Implications of metabolomics for biomedicine, drug discovery, and development
- Metabolomics case studies in diabetes and prostate cancer
- Because the metabolome is reflected in small molecules typically generated in low abundance and with a wide dynamic range, analytical science has been scrambling to devise methods for meeting the metabolomic challenge. Two techniques in particular—gas chromatography and high-performance liquid chromatography—in tandem with mass spectrometry have become the de facto platforms for analyzing the metabolome.
- Your hosts for this webinar, **Steven Fischer, Ph.D.** (Agilent Technologies), **Chris Beecher, Ph.D.** (University of Michigan), and **Christopher Newgard, Ph.D.** (Duke University), bring years of combined experience surrounding metabolomics. They will present specific examples of how metabolomics can be applied to translational medicine, diabetes, and prostate cancer. A live Q&A session will follow the presentations, offering you a chance to pose questions to our expert panelists.



Acknowledgements

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