

**Answering What and Where in complex samples:  
Advances in Imaging mass spectrometry and the  
impact of high resolution mass and ion mobility**

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# Outline

## Part 1:

- Imaging and sample selection
- Sample prep and Ionization

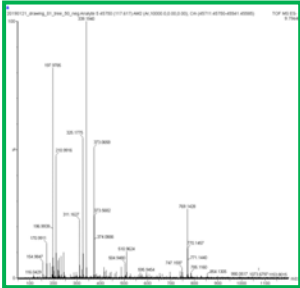
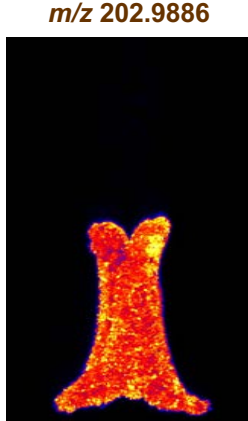
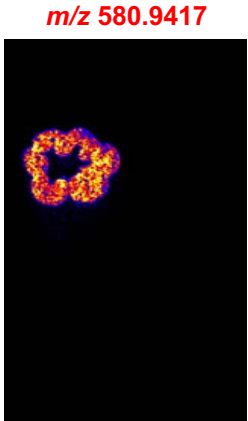
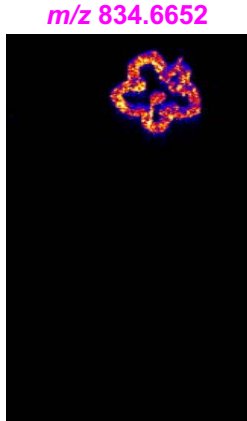
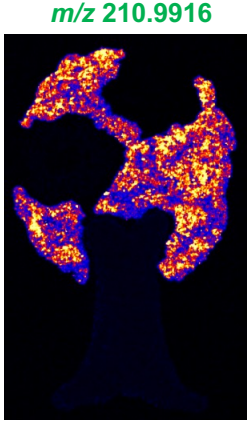
## Part 2:

- Technology for analysis

## Part 3:

- Setup and data flow
- Considerations for quantitation

# MS imaging: an overview



# Mass Spectrometry Imaging application areas

## Forensics

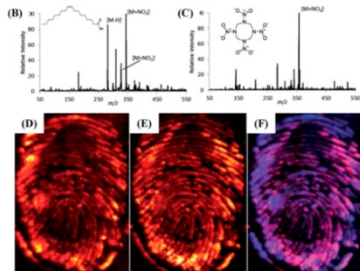
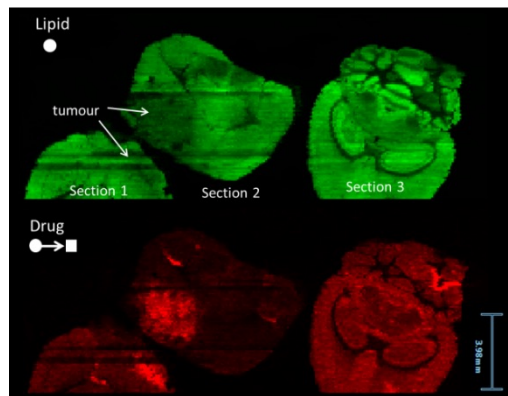
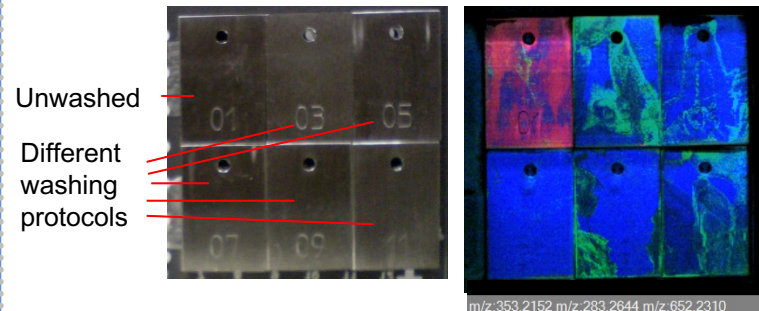


Figure 3. (A) Schematics of the DEFFIMS imaging setup. Spectra in the negative ion mode showing detection of (B) oleic acid and (C) HMX. DEFFIMS images of an HMX laden artificial fingerprint deposited onto forensic lift tape for (D) oleic acid and (E) HMX as well as (F) a colocalization map of both oleic acid (blue) and HMX (red). Adapted with permission from ref. 47. Copyright 2014 Royal Society of Chemistry.

## DMPK



## Materials



## Synthetic biology

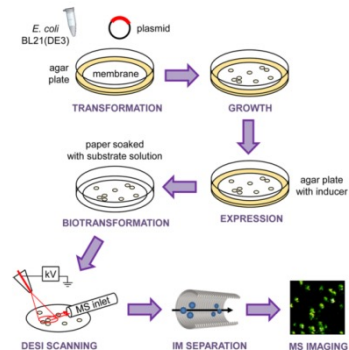
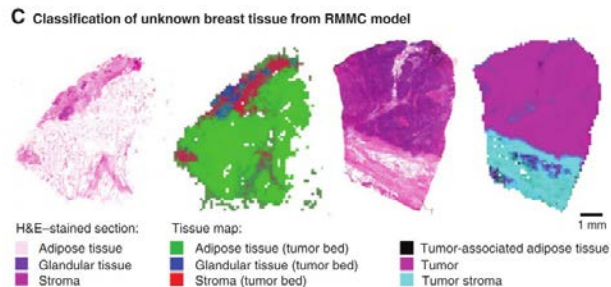
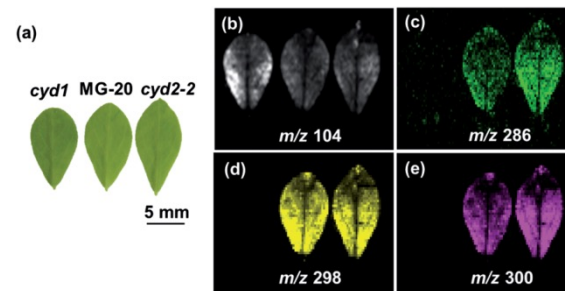


Figure 1. Workflow of DiBT-IMMS imaging of bacterial colonies expressing biocatalysts on agar plates under ambient conditions.

## Histopathology



## Agriculture



## Different MSI techniques/ionisation modes

MALDI – Matrix assisted laser desorption ionisation

DESI – Desorption electrospray ionisation

Nano DESI – Nanospray desorption electrospray ionisation (also known as liquid micro junction, not DESI and not nano!)

LESA – Liquid extraction surface analysis

SIMS – secondary ion mass spectrometry

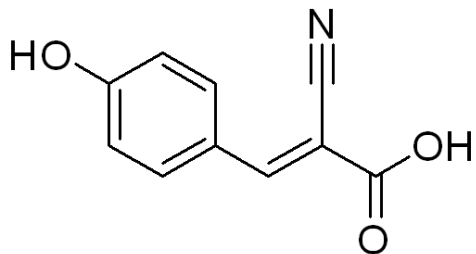
LAESI – Laser ablation electrospray mass spectrometry

LDI – Laser Desorption Ionisation

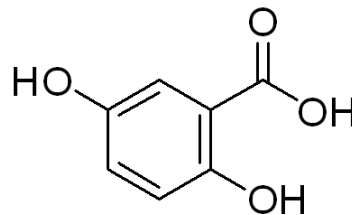
Plus many more – if information can be generated from a discrete position it can be used for imaging

## MALDI – Matrix Assisted Laser Desorption Ionisation

- Phrase first used in 1985<sup>1</sup>.
- An investigation of laser desorption of amino acids found that pure standard of alanine would not ionise but could be ionised in the presence of tryptophan
- An analyte is co-dissolved with an excess of a “matrix” molecule ( typically a small organic acid chromophore such as  $\alpha$ -Cyannohydroxycinnamic Acid (CHCA) or 2,5-Dihydroxybenzoic Acid (DHB)
- “soft” ionisation technique




CHCA







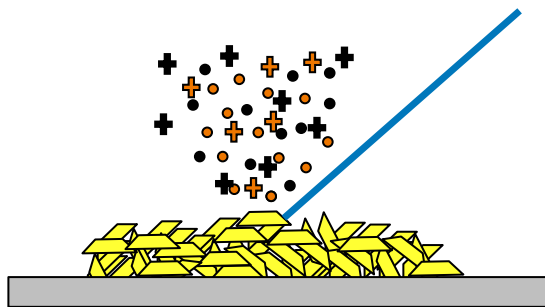
DHB

1. Karas *et al*, *Analytical Chemistry* **1985**, 57, 2935-2939

- Matrix absorbs UV light / energy protecting the analyte – triggers rapid gas phase transition

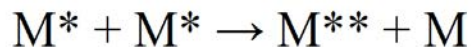
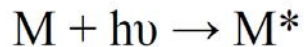
 Co-crystallised analyte / matrix

-  Neutral analyte molecule
-  Neutral matrix molecule
-  Charged analyte molecule
-  Charged matrix molecule

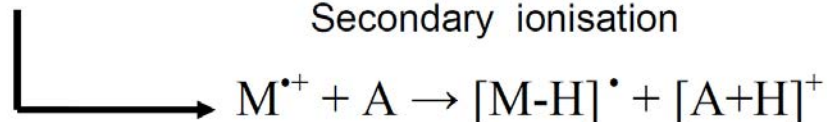


Energy pooling through singlet singlet annihilation<sup>2</sup>

Primary ionisation



Secondary ionisation



2. Ludemann *et al*, *Rapid Communications in Mass Spectrometry* **2002**, 16, 1287-1294

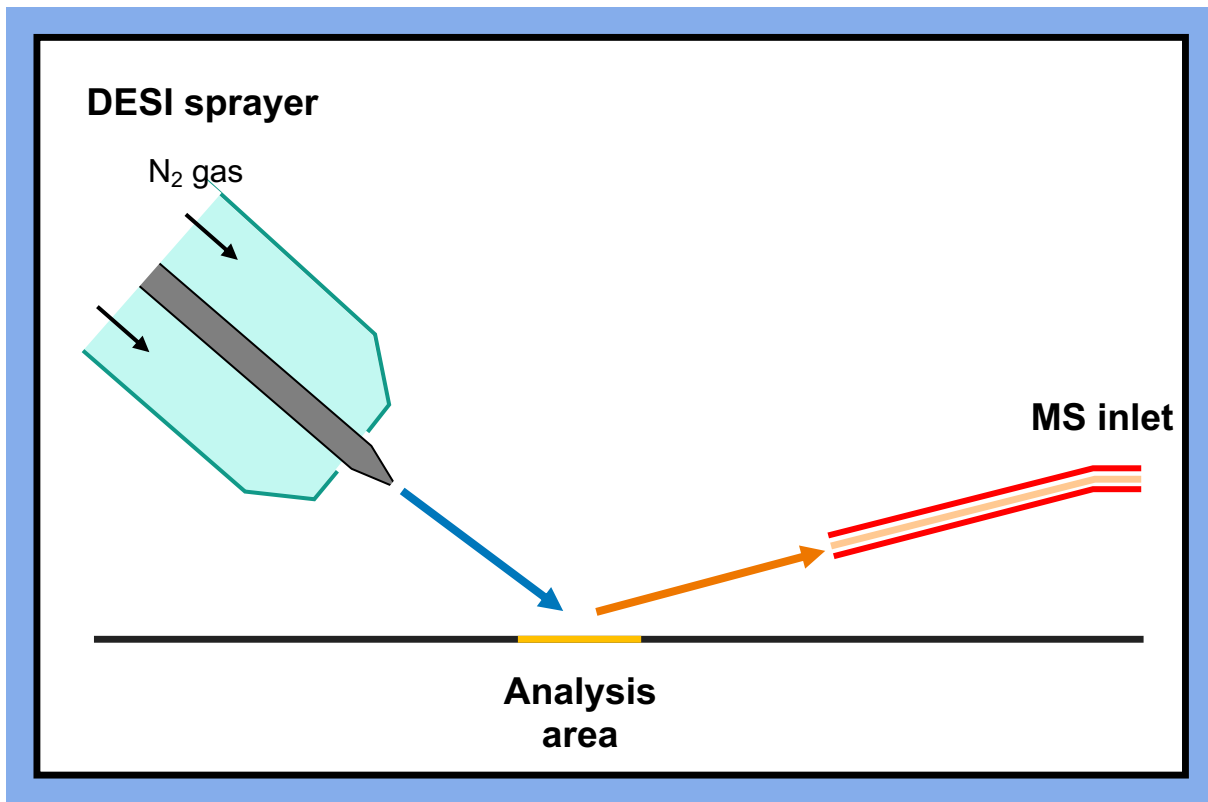


Waters™

*Making DESI More Accessible*



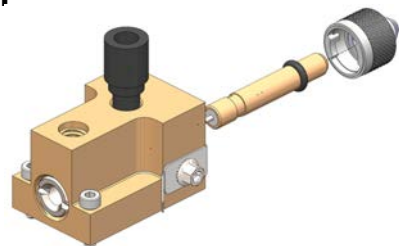
## DESI imaging: principles



- Solvent directed at surface
- $N_2$  gas focuses solvent
- Surface impact of solvent conducts **micro-extraction**
- **Ambient** method
- Little to **no sample preparation**
- ***In-situ*** examination
- **Rapid** analysis

## High- Performance Sprayer

- Improved spray focus delivering greater sensitivity and spatial resolution (25 $\mu$ m)
- Consumable cartridge with fixed emitter gives 'plug and play' ease of use

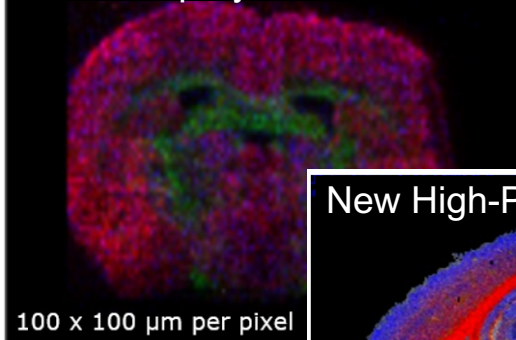


## Heated Transfer Line

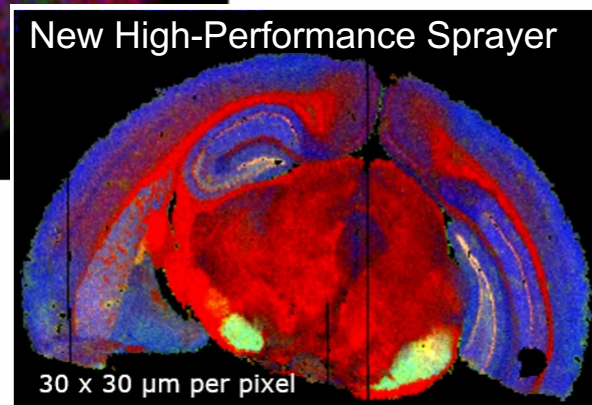
- Enhances the sensitivity performance
- Enhances the molecular range of the technique



Current Sprayer



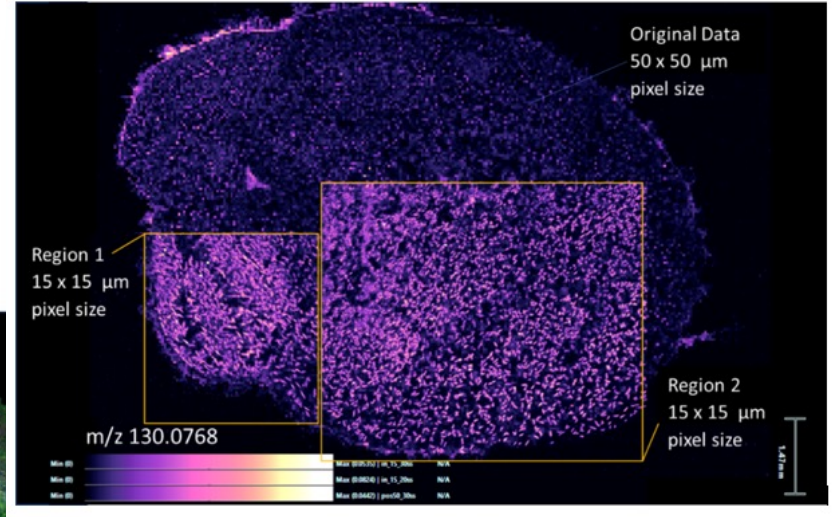
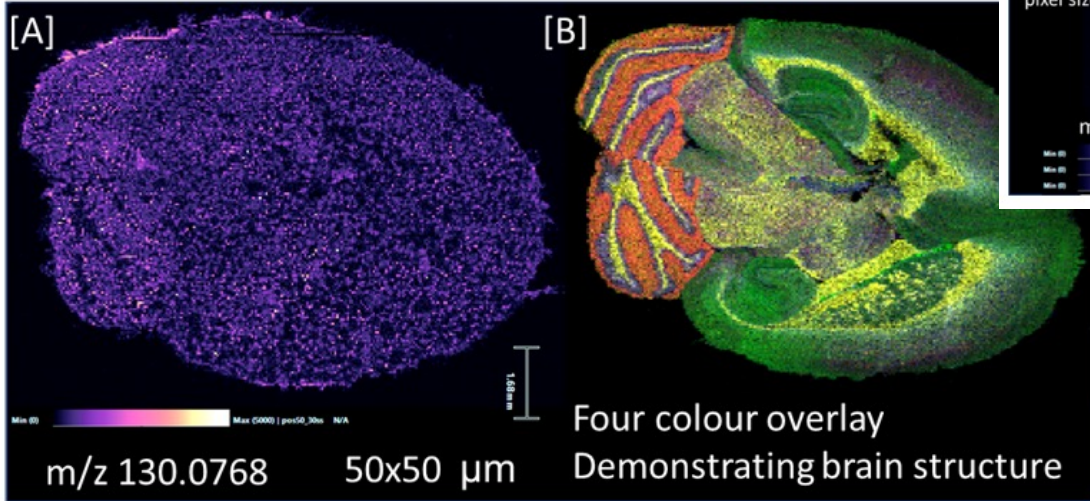
New High-Performance Sprayer



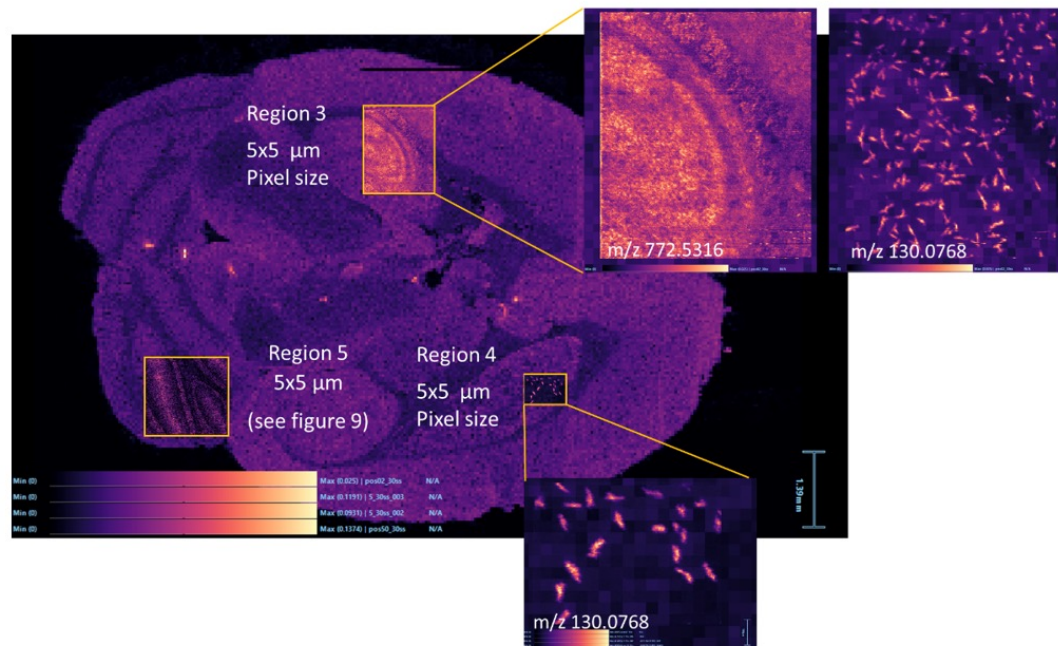
**Both improvements together can give >100x sensitivity increase**  
Enable rapid DESI imaging | Improved usability | Improved robustness

# Application of the new technologies

The ability to re-analyse the same sample at different spatial resolutions allows the DESI user to obtain the required information without large time burdens.

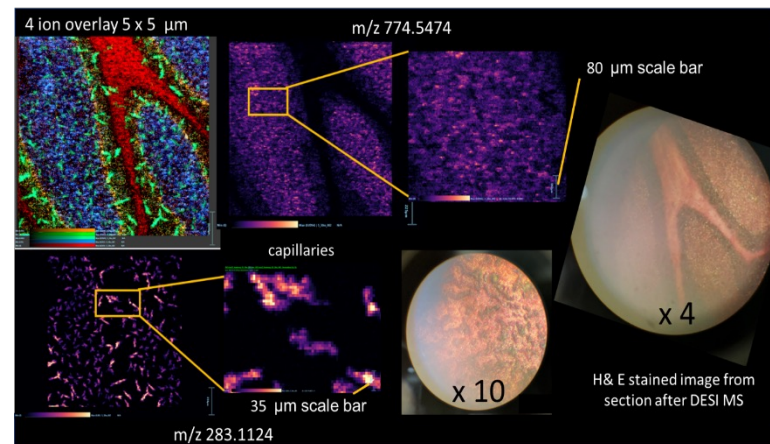


# Application of the new technologies



Two areas of the tissue analysed at **5x5  $\mu\text{m}$**

A staining procedure can enable accurate co-registration between the known structures of the tissue and MS data



Showing features which are consistent in size to **single cells within the tissue**

The background features a complex network of interconnected nodes and lines, resembling a molecular structure or a data network. The nodes are represented by small circles in various shades of blue and grey, connected by thin, light blue lines. The overall aesthetic is clean and technical, with a gradient background transitioning from light blue on the left to a darker blue on the right.

# Sample Preparation

# Imaging MS Workflow

1

## SECTIONING



Cut tissue into sections (10-15µm thick) using a cryostat

2

## SAMPLE PREPARATION



MALDI: Matrix application  
DESI: No Sample Prep

3

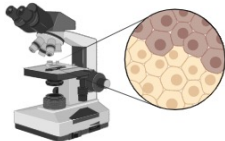
## EXPERIMENT SET-UP



Co-registration & definition of acquisition parameters

6

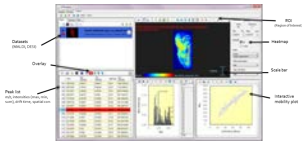
## TISSUE STAINING



Optional staining

5

## DATA PROCESSING & VISUALIZATION



Data processing and visualization in HDI

4

## MS IMAGING ACQUISITION



Data acquisition



# Sample Preparation Considerations

## *Sample and source dependant*

- non-imaging applications can be as simple as a 1  $\mu$ L spot of sample
- E.g. surface analysis

*Some samples for DESI do not require preparation but can be analyzed directly in their native state.*

- E.g. Pharmaceutical tablets, Tape used for skin analysis or plant surfaces.

*Some samples may require cleaning or treatment prior to analysis*

- E.g. Leaves may require their waxy layer removed using solvent
- E.g. Tissue will require digestion if looking for peptides

*Many samples will benefit from cryosectioning prior to analysis*

- To flatten the surface for optimal imaging
- To cut through a samples and allow imaging of the inside cross-section.

*MALDI requires a flat surface for good results and will require the application of a reagent to the surface of the sample.*



# Setting Up

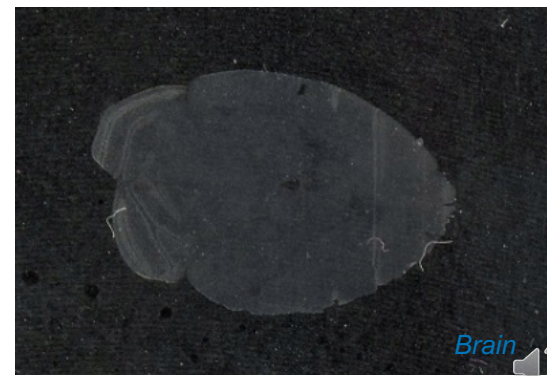
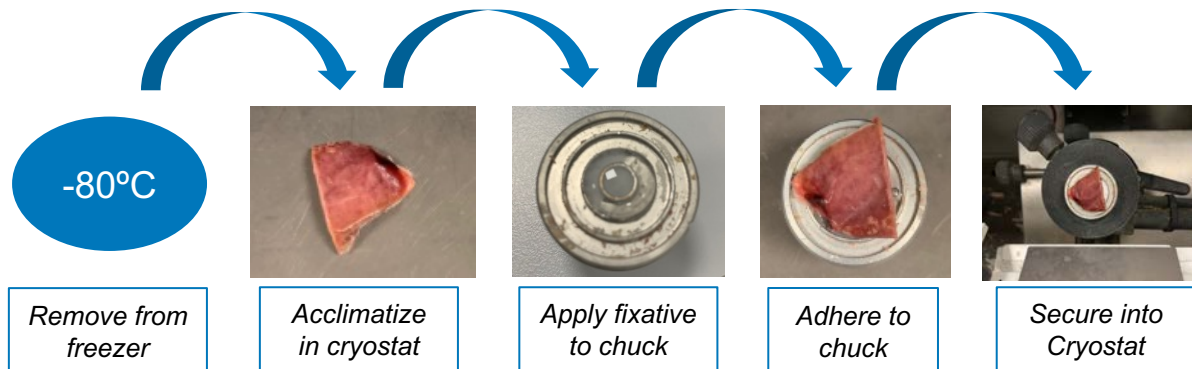
Depending upon sample type, steps for sectioning will vary

Typically, applications will be tissue based

Tissue must be frozen prior to sectioning

Remove the tissue from the freezer and adhere in a suitable orientation to the chuck.

The tissue must then be left in the cryostat for a minimum of 30 minutes to acclimatize to the same temperature as sectioning will occur





# Thickness

Tissue section thickness will depend upon the imaging technique;

MALDI section thickness **is critical**.

MALDI you **must** section at 12  $\mu\text{m}$  or less.

- if sections are thicker than this: salts within the samples will interfere with data quality.
- The salts from the tissue are preferentially drawn into the matrix and can negatively affect crystallisation and ionisation.

DESI section thickness is not critical

Typically for DESI in house we section at 16 to 20  $\mu\text{m}$

- this generates robust easily handled sections.
- these sections give good signal strength by DESI
- this thickness allows for multiple passes without loss of tissue regions.





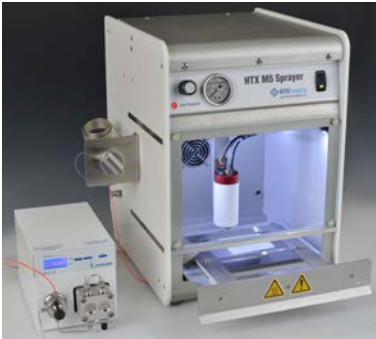
# Matrix applicators



Labcyte Portrait  
spotter



Sunchrom  
SunCollect sprayer



HTX M5 Sprayer



HTX M3+ Sprayer



HTX sublimator



## Options Pros & Cons

### Spotted matrix

- Highest sensitivity but very limited resolution
- Typically, spots are 100  $\mu\text{m}+$
- Can be used for enzyme application

### Sprayed matrix

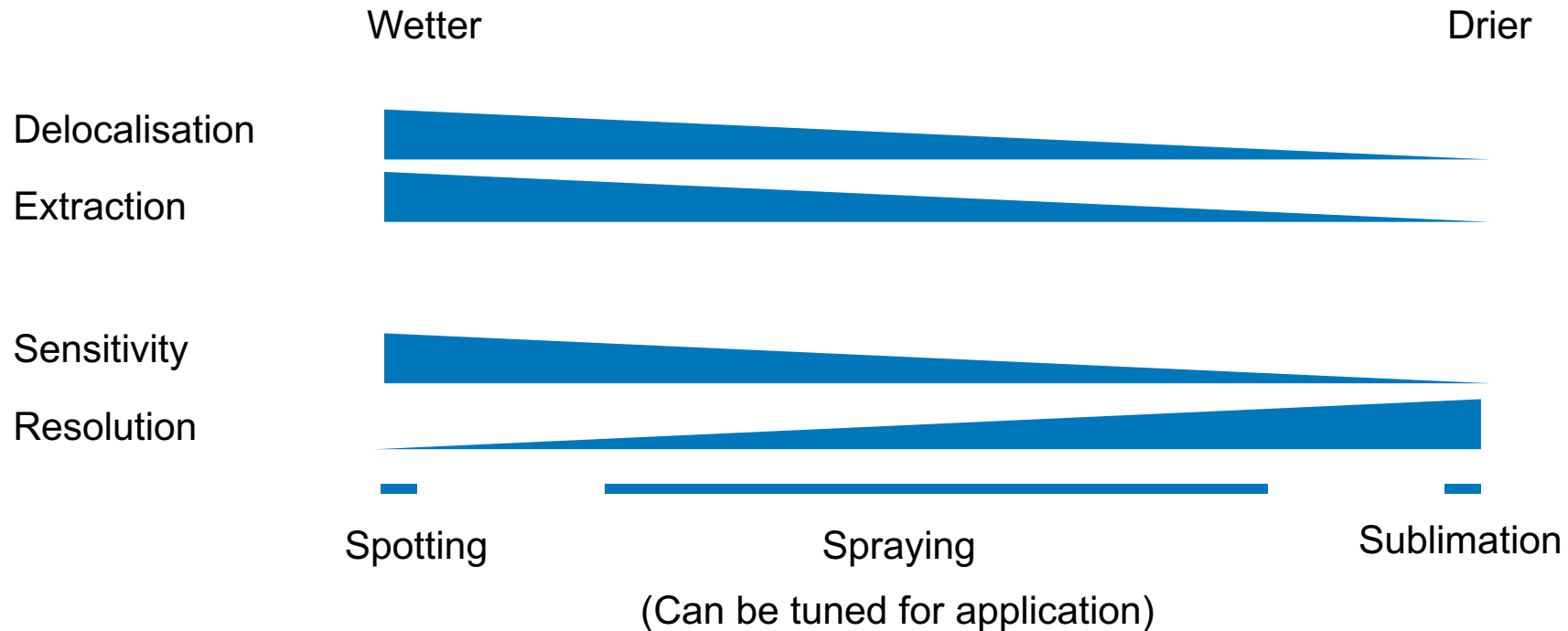
- Multiple passes to build layers.
- Possible delocalisation if tissue gets too wet.
- Highly tuneable with latest sprayer technology (heated spray heads and sample trays).
- Can be used for enzyme application

### Sublimation

- Minimal delocalisation due to matrix being in gas phase
- Low sensitivity due to no liquid extraction

Non-Imaging applications matrix can be mixed 1:1 directly with liquid samples prior to spotting onto target plates.



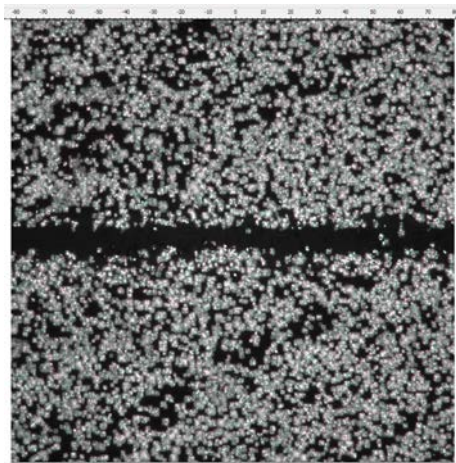


Crylas 2.5KHz Nd: YAG laser

User software control of laser focus via z-motor

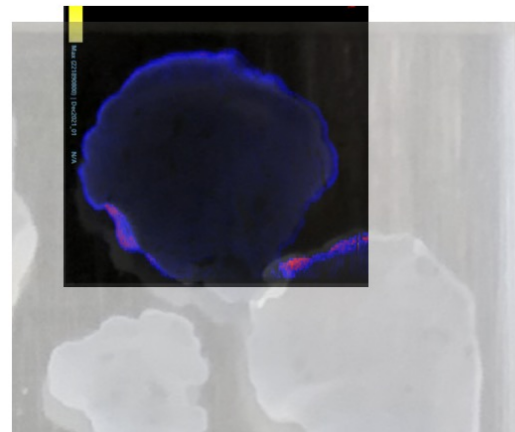
User software control of ND filter control (fine/ coarse)

<10 $\mu$ m to >100 $\mu$ m (defocused laser)



5 $\mu$ m laser focus

Thin film prep

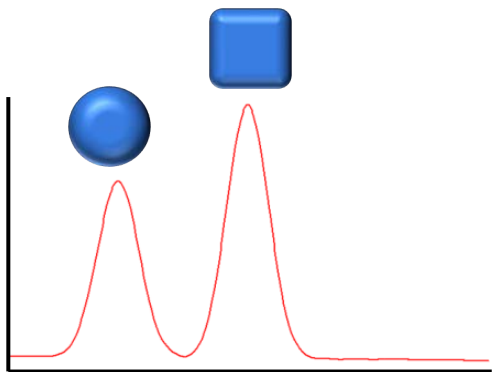


20 $\mu$ m laser focus

## Mass spectrometry – analysis consideration

- Targeted or untargeted
- Ion mobility
- Mass resolution/mass accuracy
- Analysis time (cost\$\$)

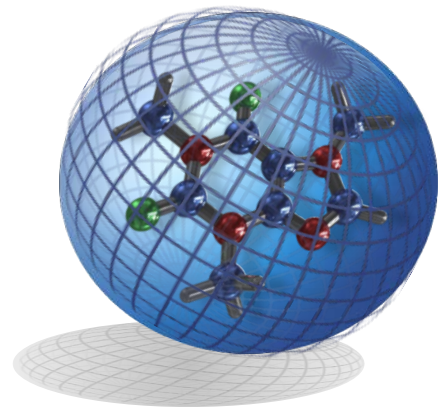
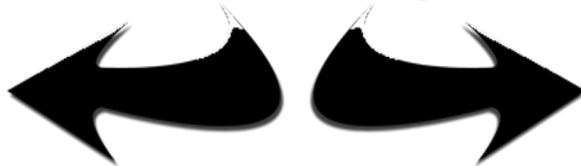
## Benefits of ion mobility



### Orthogonal Separation

- Improved Peak Capacity
  - The ability to see more or increase speed of analysis whilst maintaining resolution
- Improved Selectivity
  - Resolution of isomers
  - Cleaner spectra

# IMS



### Collision Cross Section

- IMS drift time  $\Rightarrow$  CCS
- Robust measurement reflecting,  $m/z$ , charge, conformation and shape
- Matrix independent & System independent